Acupuncture plus Low-Frequency Electrical Stimulation (Acu-LFES) attenuates
denervation-induced muscle atrophy

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Abstract

Muscle wasting occurs in a variety of clinical situations including denervation. There is no effective pharmacologic treatment for muscle wasting. In this study, we used a tibial nerve denervation model to test acupuncture plus low frequency electric stimulation (Acu-LFES) as a therapeutic strategy for muscle atrophy. Acupuncture needles were connected to an SDZ-II electronic acupuncture device delivering pulses at 20Hz and 1mA, the treatment was 15 minutes daily for 2 weeks. Acu-LFES prevented soleus and plantaris muscle weight loss and increased muscle cross-sectional area in denervated mice. The abundances of Pax7, MyoD, myogenin and embryonic myosin heavy chain were significantly increased by Acu-LFES in both normal and denervated muscle. The number of central nuclei was increased in Acu-LFES treated muscle fibers. Phosphorylation of Akt was downregulated by denervation leading to a decline in muscle mass; however, Acu-LFES prevented the denervation-induced decline by largely upregulation of the IGF-1 signaling pathway. Acu-LFES reduced the abundance of muscle catabolic proteins forkhead O transcription factor and myostatin, contributing to the attenuated muscle atrophy. Acu-LFES stimulated the expression of macrophage markers (F4/80, IL-1β and arginase-1) and inflammatory cytokines (IL-6, IFNγ and TNFα) in normal and denervated muscle. Acu-LFES also stimulated production of the muscle specific microRNAs miR-1 and miR-206. We conclude that Acu-LFES is effective in counteracting denervation-induced skeletal muscle atrophy and increasing muscle regeneration. Upregulation of IGF-1, downregulation of myostatin and alteration of microRNAs contribute to the attenuation of muscle atrophy in denervated mice.

Keywords
IGF-1, myostatin, macrophage, Akt, FoxO, cytokines
Introduction:

When skeletal muscles are denervated, they undergo atrophy. This occurs in a variety of clinical situations including trauma, motor neuron disease (amyotrophic lateral sclerosis), systemic diseases (alcoholic neuropathy or pernicious anemia) and viral infections such as polio (2, 15, 27, 39). After nerve injury, muscle fibers will progressively undergo an irreversible degeneration process if re-innervation does not occur (40). Denervation lesions result in the loss of muscle proteins, mainly reflecting increased breakdown of myofibrillar proteins, leading to muscle atrophy (45). The clinical consequences of muscle atrophy are fragility and weakness with significantly reduced quality of life. Muscle wasting is an independent index of mortality and morbidity (10, 50). There is currently no reliable pharmacologic treatment to prevent muscle atrophy so effective therapies to treat muscle wasting are needed.

Understanding the underlying mechanisms of skeletal muscle atrophy is critical for development of treatments. Skeletal muscle atrophy and hypertrophy are caused by alterations in a fine balance between protein synthesis and degradation (28). The insulin-like growth factor 1 (IGF-1)/Akt/mammalian target of rapamycin (mTOR) signaling pathway has been shown to be crucial to protein metabolism and muscle regeneration (54). Two major components in this pathway are Akt and forkhead family O (FoxO). Phosphorylation of Akt inhibits FoxO leading to increased protein synthesis and decreased protein degradation. FoxO activation (de-phosphorylation) can induce two important proteolytic pathways: the ubiquitin-proteasome pathway (UPS) and the lysosomal-autophagy pathway, leading to muscle wasting (26). In general, treatments that could upregulate IGF-1 and phosphorylate Akt would increase muscle mass and counteract muscle atrophy (45). In addition, myostatin is considered a negative regulator of muscle mass (52). Therefore, inhibition of myostatin would also reduce muscle atrophy (6, 53).

Many potential therapeutic strategies to limit muscle atrophy have been proposed including the use of pharmaceuticals, antisense oligonucleotides or gene therapy, which have been proven to be partially effective in mice (6) but have met with limited success in humans. Clinical trials for pharmacologic inhibitors of myostatin are ongoing, but none have yielded a reagent for clinical use (32, 41).
The validated treatment for muscle wasting in humans is exercise, which reduces various types of muscle atrophy (1, 9). Previously we discovered that exercise has the ability to prevent a decrease of skeletal muscle mass induced by chronic kidney disease (44). Unfortunately, patients with severe diseases or those with acute illnesses are often bed-ridden and unable to exercise regularly. Thus, there is a medical need to develop new, non-exercise-based therapies that will increase muscle mass and strength and improve patient quality of life and survival.

Acupuncture offers unique clinical advantages over pharmaceutical approaches to many disease states (8, 24). We and others have evidence that acupuncture plus low-frequency electrical stimulation (Acu-LFES) can counteract muscle atrophy (12, 31, 36, 37). Acu-LFES can suppress muscle atrophy induced by hind limb suspension in mice (31) and increase satellite cell proliferation in skeletal muscle (37). However, the effect of Acu-LFES on muscle atrophy resulting from denervation has not been established. Various muscle and nerve stimulation paradigms have been used in the US and around the world during recent years (25), including transcutaneous electrical nerve stimulation (TENS) and functional electrical stimulation (FES). In this study we will refer to Acu-LFES, which delivers electrical stimulation through needles inserted into muscle, as opposed to electrical muscle stimulation (SEMS) which delivers electrical stimulation to the skin surface by use of flat adhered electrodes. SEMS has been used as strength training in healthy people and athletes, a rehabilitation and preventive tool for disease subjects (muscle atrophy or pain release), a testing method for evaluation neural or muscle function, and for post-exercise recovery or cosmetic purposes (3, 4, 17).

The purpose of this study is to test whether Acu-LFES can be used for a therapeutic intervention for muscle atrophy induced by denervation. We investigated the impact of Acu-LFES on myogenesis, IGF-1/Akt, myostatin and FoxO signaling pathways and microRNA expression in denervation-related muscle atrophy. We report here that Acu-LFES counteracts muscle atrophy by influencing muscle regeneration and upregulation of IGF-1 in denervated mice and could provide an additional therapeutic option for treatment of denervation muscle atrophy.
Materials and methods

Animals and denervation model: Male C57/BL6 mice (8 weeks) were used for denervation models and acupuncture treatments. Mice were randomly divided into four groups; normal control, control + Acu-LFES (Acu-LFES), denervation, and denervation + Acu-LFES (denervation/Acu-LFES). Each group had 4 mice and experiments were repeated three times. The denervation animal model and Acu-LFES experiments were approved by the Institutional Animal Care and Use Committee of Emory University. Denervation was induced by transection of the medial and lateral branches of the tibial nerve that innervates the plantar flexor muscles (i.e. gastrocnemius, soleus and plantaris) close to their neuromuscular junction. The cut nerve end was sutured into the quadriceps muscle to ensure that the nerve stumps do not re-innervate.

Acu-LFES treatment: The mice were kept in specially designed restraints so that they would remain in a recumbent position during Acu-LFES treatment. Mice were awake without any anesthesia and appeared to be comfortable throughout the treatments. Acupuncture points selected were according to the WHO Standard Acupuncture guidelines (22). The positive point (anode: Yang Ling Quan, GB34) is in the hollow of the exterior-inferior of the caput fibulae about 6 mm deep. This position is close to the superficial fibular nerve and deep fibular nerve. The negative point (cathode: Zu San Li, ST36) is 5 mm beneath the capitulum fibulae and located laterally and posterior to the knee-joint about 7 mm deep and close to fibular nerve. The impulses were delivered between the 2 acupuncture needles. Disposable sterile needles with a diameter of 0.25 mm (Shen Li Medical & Health Material Co., Ltd., Wujiang, China) were used. The needles were connected into SDZ-II Electronic acupuncture instrument using consistent pulse, electric frequency 20Hz, electric current 1mA. These stimulation conditions were chosen because 1) they were effectively used in a previous study (14), 2) we have found that they are effective in treating muscle atrophy in mice with CKD (12), and 3) the mice do not appear to be uncomfortable at this level of stimulation.

Experimental design: Denervation surgery was performed on day-1, and Acu-LFES was started on day-2. The Acu-LFES was administered for 15 minutes every day for 2-weeks at 10am. Hindlimb muscles (gastrocnemius, soleus and plantaris) were
harvested 48 hours after the last Acu-LFES treatment. The control mice were placed in
these specially designed restraints and remain in a recumbent position without Acu-
LFES treatment for an equal amount of time and for the same number of “treatments”.

**Western blot, muscle histology and antibodies:** Hind limb muscles were
homogenized in gentle lysis buffer (10 mM Tris-HCl, 10 mM NaCl, 2 mM EDTA, 0.5 %
NP-40, 1 % glycerol, and fresh added: 1 mM Na3VO4; 10 μg/ml PMSF; 5 μg/ml aprotinin;
1 μg/ml leupeptin) with phosphatase inhibitors cocktail 1 and 2 (Sigma). Proteins were
subjected to Western blot analysis using previously published methods (55).

For Immunohistology: muscles were embedded under Tissue Freezing Media
(TBS; Fisher, Pittsburgh, PA, USA) in isopentane cooled in dry ice. Cross sections (10
mm) from the mid-belly of different groups of muscle were mounted on gelatin-coated
slides were fixed in 4% paraformaldehyde for 10 min. Tissue was permeabilized in
0.05% Triton X-100 (in PBS) for 10 min, and quench-fixed in 50 mM NH4Cl for another
10 min. Samples were blocked with 5% bovine serum albumin for 1 h, followed by
incubation overnight with first antibody. Sections were further incubated for 60 min with
rhodamine red-labeled antirabbit IgG (diluted 1:100; Jackson Immuno Research Lab,
West Grove, PA). Nuclei were stained by DAPI. Images were visualized with an
Olympus 1X51 inverted fluorescence microscope and captured by DP73-1-51-17MP
color camera. The positive cells number of at least 500 individual myofibers per muscle
was measured using the CellSens Dimension 1.9 with count & measure full Software
(Olympus, Melville, NY, USA). Muscle fiber cross-sectional area was determined in
plantaris muscles using an anti-laminin antibody (1:50 dilution; Sigma-Aldrich).

Muscle histology: Gastrocnemius muscles were processed for cryosectioning
after placing them in cryo-molds containing embedding medium. The frozen sections
were stained using a Masson modified IMEB trichrome stain kit according to the
manufacturer’s instructions (International Medical Equipment B Inc.; San Marcos, CA).

Primary antibodies (1:1000 dilution except indicated) that we used included: Akt,
p-Akt (Ser473), FoxO1, FoxO3, p-FoxO1/3 (Thr24/Thr32), mTOR, p-mTOR (Ser2448),
P70S6K, p-P70S6K (Thr389), p-Smad3 (Ser423/425), p-Smad2 (Ser465/467) and
Smad2/3 (D7G7) from Cell Signaling (Danvers, MA), GDF8/myostatin from Abcam
(Cambridge, MA), MyoD, Pax7, Myogenin and embryonic myosin heavy chain (eMyHC)
from DSHB product (University of Iowa, Iowa, IA) and GAPDH from Millipore (Burlington, MA). Protein bands were scanned and quantified using the Li-cor Odyssey infrared scanning system (Li-COR Biosciences, Lincoln, Nebraska).

**Enzyme-Linked Immunosorbent Assay (ELISA):** mouse IGF1 in muscle lysates were quantized by IGF1 Mouse ELISA Kit (ab100695, Abcam, Cambridge, MA) and used according to manufacturer's instructions. Plasma cytokine levels were measured by Bioassay Services Team using the cytokine protein ELISA quantification assay (AssayGate, Inc. Ijamsville, MD).

**Reverse transcription and quantitative PCR (q-PCR) for mRNA and microRNA:**

Total RNA was extracted using Tri-Reagent (Molecular Research Inc., Cincinnati, OH) (13). For mRNA, reverse transcription was performed using the M-MLV reverse transcriptase (Invitrogen) and 2 μg denatured RNA according to the manufacturer's instructions. Real-time qPCR was performed with SsoAdvUniversal SYBR Green PCR Reagents (Bio-Rad, Hercules, CA) using the following cycle parameters: 94°C for 2 minutes and 40 cycles at 94°C for 15 seconds, 55°C for 30 seconds, 72°C for 30 seconds with final extension at 72°C for 10 minutes. The quantification cycle (Cq) values was defined as the number of cycles required for the fluorescence signal to exceed the detection threshold. Individual mRNA expression was standardized to 18S rRNA and expression was calculated as the difference between the threshold values of the two genes (2-Δcq). Melting curve analysis was routinely performed to verify the specificity of the reaction. Primers for specific genes were designed to cross intron-exon boundaries and used to generate amplicon in their linear ranges (table 1). For microRNA, the miRCURY LNA™ Universal cDNA Synthesis kit (Exiqon INC., Woburn, MA) was used for reverse transcription of total RNA. The primers were purchased from Exiqon. The miRCURY LNA microRNA PCR SYBR Green master mix (Exiqon INC) was used for qPCR with the following cycle parameters: 95°C for 10 minutes and 45 cycles at 95°C for 10 seconds and 60°C for 60 seconds. Expression of individual microRNA was standardized to the mouse U6 gene.

**Statistical analysis:** Data were presented as mean ± se. To identify significant differences between two groups, comparisons were made by using the t-test. For a comparison of more than two groups, two-way ANOVA was performed with a post hoc
analysis by the Student-Newman-Keuls test. A Chi Square analysis was used to
analyze shifts in the muscle cross sectional area. Differences with P values < 0.05 were
considered significant.
Results:

*Acu-LFES prevents denervation-induced muscle wasting:* Mice (C57BL/6J, 8 weeks) were randomly assigned into four groups: control, Acu-LFES, denervation, and denervation/Acu-LFES (n=12/group). The ratio of individual muscle weight to body weight was measured to identify muscle wasting. The weight ratios of gastrocnemius muscle, but not soleus and plantaris muscles, in the Acu-LFES mice were increased compared with the weight ratio in control mice without Acu-LFES (Table 1). Denervation caused a reduction in relative muscle weight by 37% (soleus), 34% (plantaris) and 20% (gastrocnemius) versus control muscle. Acu-LFES alleviated some of the muscle wasting. The average weight ratio in denervation/Acu-LFES mice was significantly higher versus denervation mice without Acu-LFES in gastrocnemius, soleus and plantaris muscles.

*Acu-LFES improves muscle regeneration in normal and denervation mice:* In a separate experiment, we compared control (no treatment) and manual acupuncture (insertion of the needles without electrical stimulation). We did not see a statistical difference in the myogenic protein markers (myoD, myogenin and eMyHC or the number of central nuclei and collagen accumulation (data not show) between the muscle from control and manual acupuncture mice. Acupuncture accompanied with electrical stimulation did show a positive effect on denervation-induced muscle atrophy as we show below. To study how Acu-LFES prevents muscle wasting in denervation mice, we first measured protein amounts of myogenesis markers which indicate the regeneration capacity of the muscles. Acu-LFES stimulates protein production under baseline conditions since in the muscle of Acu-LFES mice, the protein expressions of Pax7 (transcription factor to initial myogenesis), MyoD (proliferation marker), myogenin (differentiation marker) and eMyHC (terminal differentiation marker) were significantly increased compared with control mice (Figure 1A). The denervation alone also increased muscle regeneration markers vs. control mice; however, Acu-LFES caused a substantial additional increase in all myogenesis markers (2.5-fold in Pax7, 2.8-fold in myoD, 2.2-fold in myogenin and 2.5 fold in eMyHC) in denervated muscle. Interestingly, the increase in myogenesis markers by Acu-LFES on the denervation background was also significantly higher than Acu-LFES-mediated increase in control muscle without
denervation. These results demonstrate that Acu-LFES counteracts denervation-induced muscle loss by activation of muscle regeneration.

Central nuclei are indicators of muscle regeneration. Upon initiation of myogenesis by Acu-LFES, the central nuclei migrate into the myofibers (Figure 1B). We found increased central nuclei in muscles of both denervation and control mice treated with Acu-LFES. However, the percentage increase in central nuclei was higher in the muscles of denervation/Acu-LFES than of control/Acu-LFES mice. We also found that Acu-LFES prevents denervation-induced decrease in the muscle fiber cross-sectional area. Muscle fiber cross-sectional area was determined in frozen sections of plantaris muscles using an anti-laminin antibody. The size of muscle fibers was significantly smaller in denervation mice without Acu-LFES than in denervation mice with Acu-LFES. Fiber area frequency distribution revealed a clear increase in the percentage of larger fibers (shift to the right) in Acu/LFES–treated denervation mice relative to the distribution curve for denervation alone (Figure 1C). These results show that Acu-LFES effectively reduces muscle atrophy.

**Acu-LFES upregulates IGF-1 in skeletal muscle of normal and denervated mice.** To investigate how Acu-LFES activates the muscle regeneration process, we identified mRNA expression of IGF-1 using quantitative real time PCR (qPCR). Acu-LFES increased IGF-1 expression by 2.2-fold in normal control mice. IGF-1 expression was slightly decreased in denervated muscle at 2 weeks; but was increased 1.9-fold in denervation/Acu-LFES muscle (Figure 2A). Protein levels of IGF-1 were measured by ELISA in muscle lysates. Denervation mice showed 10% lower levels of IGF vs. control mice. Acu-LFES raised IGF-1 protein by 1.6-fold in muscle of denervation mice (Figure 2B). These results indicate that IGF-1 is increased by Acu-LFES treatment in the muscle of normal and denervated mice, which could contribute to increased muscle regeneration.

**Acu-LFES increases Akt phosphorylation and inhibits FoxO activation in denervation mice:** Activation of Akt leads to phosphorylation and inactivation of FoxO which reduces its pro-catabolic functions (46). We assayed phosphorylation of Akt, FoxO1 and FoxO3 in all groups of mice (Figure 2C). Since phosphorylation of these proteins indicates protein activation or inhibition, we expressed the data as the ratio of
phosphoprotein to total protein. Acu-LFES treatment increased the pAkt/Akt ratio 2.2-fold in normal control muscle and 2.9-fold in denervation muscle. The ratio of pFoxO1 thr24/FoxO1 increased 2.5-fold in Acu-LFES treated normal mice and 2.1-fold in denervation muscle. The ratio of pFoxO3 thr32/FoxO3 increased 2.0-fold in Acu-LFES treated normal mice and 2.4-fold in denervation muscle. These results demonstrate that Acu-LFES improves protein metabolism by increasing Akt and FoxO phosphorylation.

**Acu-LFES upregulates phosphorylation of mammalian target of rapamycin (mTOR) and P70S6 kinase (P70S6K) in normal and denervated mice:** Phosphorylated mTOR and P70S6K are the acknowledged active forms of these proteins and upregulated during muscle protein anabolism. The ratio of phospho-mTOR to total of mTOR increased 1.8-fold by Acu-LFES in normal control muscle. Denervation alone also increased mTOR phosphorylation by 1.5-fold. Acupuncture did not promote an additional increase in phospho-mTOR above levels in the denervation muscle. The phosphorylation of the P70S6K increased 1.5-fold in Acu-LFES treated normal mice. Unlike p-mTOR, denervation did not change pP70S6K; however, pP70S6K was increased 2.5-fold by Acu-LFES treatment in the denervation mice. These results suggest that Acu-LFES increases phosphorylation of P70S6K and may lead to increased protein anabolism (Figure 2D).

**Acu-LFES decreases myostatin in normal and denervation mice:** Decreased myostatin is an indicator of increasing muscle mass. To identify whether Acu-LFES changes the expression of myostatin, mRNA and protein levels of myostatin were measured in all groups of mice. Acu-LFES decreased myostatin expression by 35% in normal control mice. Denervation also decreases myostatin expression; however, Acu-LFES had an additional 23% decrease in denervation muscle (Figure 3A). Myostatin protein was 55% decreased by Acu-LFES in normal control mouse. We did not see a significant change after two weeks in denervation muscle; but, Acu-LFES reduced the myostatin protein level by 53% in denervated muscle (Figure 3B). Myostatin induces muscle atrophy by activation of the transcription factors SMAD2 (Mothers against decapentaplegic homolog 2) and SMAD3 (33). We found that phosphorylation (activation) of SMAD2 and SMAD3 was repressed by Acu-LFES in denervation muscle consistent with the observed decrease in myostatin. These results indicate that
one of the ways that Acu-LFES ameliorates denervation-induced muscle atrophy is by suppressing myostatin, pSmad2 and pSmad3.

**Acu-LFES activates macrophages and increases inflammatory cytokines in normal and denervated muscle:** An early step in muscle regeneration is activation of macrophages and upregulation of inflammatory cytokines. We examined the markers of macrophage levels. We assayed mRNA expression of general macrophage marker F4/80, an M1 macrophage marker IL-1β and an M2 macrophage marker arginase-1 by qPCR. F4/80 expression was elevated 5-fold in the muscle of Acu-LFES mice, 2.3-fold in the denervation muscle and 4.4-fold in denervation/Acu-LFES mice versus controls. IL-1β expression was elevated 3.6-fold in the muscle of Acu-LFES mice, 1.9-fold in the denervation muscle and 4.1-fold in denervation/Acu-LFES mice versus controls. Arginase-1 expression was increased 6.1-fold in the muscle of Acu-LFES mice, 3.2-fold in the denervation muscle and 6.3-fold in denervation/Acu-LFES mice versus controls (Figure 4A). These data indicate that Acu-LFES upregulates the M1 and M2 macrophage expression after 2-weeks treatments. The levels of increase were higher in M2 than M1 macrophages.

Since macrophages influence myogenesis through cytokines, we examined several inflammatory cytokines in the muscle of all groups of mice. The largest change was observed in IL-6 expression which was increased 7-fold in the muscle of Acu-LFES mice vs. control. Denervation also increased IL-6 expression by 2-fold and denervation/Acu-LFES has additional increase to a total of 10.5-fold compared with control mice. IFN\(_\gamma\) expression showed a 4.5-fold increase by Acu-LFES, and no change in denervation group and 3.8-fold increase in denervation/Acu-LFES group versus control mice. TNF\(\alpha\) mRNAs also showed an increased expression by Acu-LFES in normal and denervation mice (Figure 4B). To confirm that the protein levels of cytokines mirrored the altered RNA levels in response to Acu-LFES, we measured the levels of 8 cytokines using ELISA assay in plasma and muscle lysate for all groups of mice. Acu-LFES did not significantly increase the plasma cytokine levels except for IL-10 (Figure 4C), which indicates that cytokine response to Acu-LFES occurs locally. However, Acu-LFES significantly upregulated all 8 cytokines in the muscle of normal mice: 2-fold in IL-6; 2-fold in TNF\(\alpha\); 1.4-fold in IFN\(\gamma\); 4-fold in TGF\(\beta\); 1.6-fold in IL-1β;
1.6-fold in IL-4; 1.9-fold in IL-13 and 11-fold in IL-10; but Acu-LFES did not significantly increase the muscle lysates cytokine levels except for IL-6 in denervated muscle (Figure 4D).

**Acu-LFES has no significant impact on the abundance of collagen in the muscle of normal mice:** We used Masson Trichrome staining of muscle frozen sections from each condition to assess collagen content. Collagen showed a trend toward increased amount in Acu-LFES treated normal muscle. However, using the CellSens Dimension 1.9 with full count & measure Software (Olympus, Melville, NY, USA) calculation, there were no statistic significant differences in the area of blue color staining in each group to indicate the increase in collagen accumulation (Figure 5).

**Acu-LFES enhances the expression of muscle specific microRNA (myomiRs):** Muscle specific microRNAs also influence muscle regeneration. To identify whether Acu-LFES stimulates the expression of muscle specific microRNAs, we measured microRNAs by qPCR in muscle after 2 weeks of Acu-LFES. The expressions of miR-1 and miR-206 were significantly increased by Acu-LFES in control mice (2.0-fold for miR-1 and 2.4 fold for miR-206). Denervation resulted in a 20% decrease in miR-1 expression but the change was not statistically significant due to the animal to animal variability. Acu-LFES caused increases of 2.2-fold for miR-1 expression and 2.8 fold for miR-206 expression in denervation mice (Figure 6). The expressions of both miR-133a and miR-133b also showed a trend toward increased expression by Acu-LFES in normal and denervation muscle, but the changes were not statistically significant (data not show). These data indicate that Acu-LFES stimulates the expressions of miR-1 and -206.
Discussion:
Numerous investigations have focused on identifying new therapeutic approaches to attenuate muscle wasting. Trials using intramuscular electrodes have been investigated to reduce denervation-induced muscle atrophy. Willand et al. showed that a 1-hour per day regimen of electric stimulation using intramuscular stimulating electrodes effectively increases muscle mass and force (47, 48). However, it is not easy to translate this invasive method to clinical use. Acu-LFES is a therapy that can decrease skeletal muscle atrophy induced by hind limb suspension in mice and has been used clinically for multiple disorders in humans (7, 31). Previously, we performed Acu-LFES on mice with chronic kidney disease-induced muscle atrophy and found that this treatment improved muscle weights and function (12). In the current study we found that Acu-LFES shows significant benefits for attenuating muscle atrophy and increasing muscle regeneration capacity. There are several potential advantages of Acu-LFES over surface electrical stimulation. First, since acupuncture needles cross the skin barrier, Acu-LFES subjects have more tolerance for higher current intensities than SEMS subjects, because skin has more sensory nerves. In addition, the electrical stimulation of Acu-LFES is delivered through metal acupuncture needles directly into the muscle, and close to the nerve. Because of the proximity, Acu-LFES yields a stronger stimulation at a given power than surface (SEMS) stimulation at the same power setting. The needle approach elicits a multiple muscle response that mimics the muscle action in treadmill training. SEMS does not elicit the same multiple muscle response. Lastly, muscle regeneration is an injury-regeneration process (11). In Acu-LFES the insertion of needles causes subtle myofiber injuries leading to activation of quiescent satellite cells and initiates muscle regeneration. The evidence for this injury-regeneration response is the presence of increased central nuclei and increased myogenesis markers (MyoD, myogenin, eMyHC) in the muscle of Acu-LFES treated mice.

In general, muscle wasting is the loss of both muscle cells and protein content. Acu-LFES is a therapy that can address muscle wasting from two directions, promoting the regeneration of the muscle cells and blocking the loss of the muscle protein (44, 54). The first step of muscle regeneration is to activate myogenic stem cells, also known as
satellite cells. This process involves a cascade of transcription factors that promote different steps in the regeneration process. Initiation of myogenesis is triggered by the transcription factor Pax7(21). Proliferation and differentiation involves a complex interaction of multiple proteins of which MyoD is an essential element. Differentiation of myoblasts into myotubes requires myogenin to promote fusion of myogenic precursor cells to either new or previously existing fibers. Myosin is the main component of myofibrils (thick filaments), best known for their role in muscle contraction. Myosin heavy chain protein is a marker indicating mature muscle (38). Each of these components and markers for muscle regeneration is upregulated in the muscle of normal and denervated mouse in response to Acu-LFES.

In denervated muscle the number of satellite cells is also increased in both animals (16) and in humans (30) suggesting that, although muscle mass is reduced, muscle regeneration is stimulated. This regeneration capacity is a constant response to the denervated state since myoD, myogenin and eMyHC are upregulated even 3 and 9 months post-denervation (18). In the current study, we confirmed that satellite cells were activated in denervated muscle suggesting increased regenerative capacity but muscle wasting still occurred as evidenced by decreased muscle weight (Table 1) and muscle cross-sectional area (Figure 1). Therefore, factors other than increased regeneration potential may be required to attenuate muscle atrophy in denervated muscle.

There are a few possible targets of Acu-LFES that could contribute to the attenuation of muscle atrophy. IGF-1 is a growth factor that actives satellite cells to start muscle regeneration. In addition, an increase in IGF-1 signaling will counteract muscle atrophy by promoting protein synthesis and prohibiting protein degradation induced by muscle injury and diseases (20, 42, 44, 54, 55). The expression of IGF-1 in denervated muscle changes in two phases. Expression of IGF-1 is increased at 3 days after denervation but then declines later (51). Overexpression of IGF-1 in muscle reduces the amount of myofiber atrophy by 30% in the muscle of denervated mice (35). In the current study we found that after 2 weeks of Acu-LFES, IGF-1 mRNA and protein were upregulated relative to the level of IGF-1 in untreated denervated muscle. An increase in IGF-1
leads to inhibition of the pro-catabolic factor FoxO, by increasing FoxO phosphorylation, providing a means of preserving muscle mass and function.

Myostatin is a member of the TGF-β superfamily of secreted proteins and produced primarily in skeletal muscle (19). Myostatin is a potent negative regulator of muscle mass (53). In published accounts, myostatin infusion into mice led to a 33% decrease in body weight and a 35–50% decrease in muscle mass, whereas genetic or functional knockout of myostatin resulted in increased muscle mass of mice, sheep, cattle, or humans (34, 56). In denervated muscle, myostatin mRNA and protein expression gradually increased (1 to 14 days), and then gradually declined over a later response phase (14 to 56 days) (5, 23). In our two week denervated mice, the levels of myostatin mRNA were decreased, but the protein amounts were not different from control levels. However Acu-LFES reduced both myostatin mRNA expression and protein abundance in both control and denervated muscle, which could contribute to the positive effect of Acu-LFES against muscle atrophy. In agreement with our study, Takaoka et al. reported that electrical acupuncture treatment suppressed myostatin expression, which led to muscle hyperplasia in mice (37).

MicroRNAs offer another level of control at a translational level contributing to regulation of muscle wasting (43). Direct application of miR-1, miR-133 and miR-206 to muscle by injection induced myoD, Pax7 and myogenin protein production leading to increase muscle regeneration in a rat skeletal muscle injury model (29). Williams et al. provided evidence that miR-206 is required for efficient regeneration of neuromuscular synapses after acute nerve injury (49). In the current study, we found that miR-206 was decreased in denervated muscle. Acu-LFES not only increased miR-1 and -206 in normal control mouse muscle, but also reversed the decrease of these microRNAs in denervated muscle.

In conclusion, Acu-LFES ameliorated denervation-induced skeletal muscle atrophy by multiple mechanisms, including improvements in muscle regeneration, upregulation of IGF-1 and downregulation of myostatin signaling pathways and raising the expression of microRNAs. Our work contributes to a better understanding of the mechanisms involved in Acu-LFES treatment of muscle wasting.
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Figure legend:
Figure 1: Acu-LFES increases muscle regeneration.
(A) Western blot of Pax7, myoD, myogenin and eMyHC in hindlimb (gastrocnemius, soleus and plantaris) muscle lysates from normal control, Acu-LFES, denervation or denervation/Acu-LFES mice. The bar graph compares the mean densities of all bands in each group expressed as a fold-change from levels in normal control mice which is represented by a line at 1-fold. All band densities were normalized to the density of the GAPDH band (Bars: mean ± s.e.; n=12/group; *=p<0.05 vs. control and #=p<0.05 vs. denervation).

(B) Shown are representative cross-sections from plantaris muscles after staining for laminin (red, cell perimeter) and counterstaining with DAPI (blue, nuclei) are shown in control, Acu-LFES, denervation or denervation/Acu-LFES mice. The white arrows point to central nuclei inside of myofibers which indicates satellite cell migration. The bar graph shows the number of nuclei that have migrated to the center of the cells (central nuclei) per 500 muscle fibers (Bars: mean ± s.e.; n=9; *=p<0.05 vs. control and #=p<0.05 vs. denervation).

(C) A representative cross-sectional area of plantaris muscle from control, denervation or denervation/Acu-LFES mice are compared. Cryosections of plantaris muscle were immunostained with anti-laminin antibody. The frequency distribution of fiber cross-sectional area in control (grey bar), denervation (open bar) and denervation/Acu-LFES (black bar) mice is presented as percent cross section area of fibers. The cross section area is significantly different between denervation and denervation/Acu-LFES group according to a Cai Square analysis (P<0.05, n=6).

Figure 2: Acu-LFES upregulates IGF-1 mRNA and protein in the muscle of denervated mice.
(A) Total RNAs were isolated from hindlimb muscles of control, Acu-LFES, denervation or denervation/Acu-LFES mice. The expression of IGF-1 mRNA was assayed by real time qPCR. The bar graph shows mean mRNA from the muscles of each group of mice. Results are normalized to 18S RNA (Bars: mean ± s.e.; n=12/group; *=p<0.05 vs. control and #=p<0.05 vs. denervation).
(B): IGF-1 protein levels were measured by ELISA in hindlimb muscle lysates from normal control, Acu-LFES, denervation or denervation/Acu-LFES mice. The bar graph shows the mean of IGF-1 protein level in each group. IGF-1 levels in the muscle lysates were normalized to the total protein concentration (Bars: mean ± s.e.; n=9; *=p<0.05 vs. control and #=p<0.05 vs. denervation).

(C): The protein metabolism related proteins Akt, p-Akt, FoxO1, p-FoxO1, FoxO3 and pFoxO3 were measured by Western blotting in hindlimb muscle lysates from control, Acu-LFES, denervation or denervation/Acu-LFES mice. Since the phosphorylation of these proteins indicates protein activation or inhibition, we expressed the data as the ratio of each phospho-protein to total protein. The bar graph shows the fold change of the each protein band ratio compared with levels in control mice (represented by a line at 1-fold). All band densities were normalized to the density of GAPDH (Bars: mean ± s.e.; n=12/group; *=p<0.05 vs. control and #=p<0.05 vs. denervation).

(D) Acu-LFES improves protein anabolic markers in skeletal muscle of denervated mice. The protein synthesis related markers mTOR, p-mTOR, p70S6K and p-p70S6K were measured by Western blotting in hindlimb muscle lysates from normal control, Acu-LFES, denervation or denervation/Acu-LFES mice. The bar graph shows the fold change of the each protein band ratio compared with levels in control mice (represented by a line at 1-fold). All band densities were normalized to the density of GAPDH (Bars: mean ± s.e.; n=12/group; *=p<0.05 vs. control and #=p<0.05 vs. denervation).

Figure 3: Acu-LFES downregulates myostatin mRNA and protein in the muscle of denervated mice.

(A) Total RNAs were isolated from hindlimb muscles of control, Acu-LFES, denervation or denervation/Acu-LFES mice. The expression of myostatin was assayed by real time qPCR. The bar graph shows mRNA from the muscles of each group of mice. Results are normalized to 18S RNA (Bars: mean ± s.e.; n=12/group; *=p<0.05 vs. control and #=p<0.05 vs. denervation).

(B): Myostatin, SMAD 2/3 and phosphorylated SMAD 2/3 were measured by Western blotting in hindlimb muscle lysates from control, Acu-LFES, denervation or denervation/Acu-LFES mice. The data expressed as the ratio of each phospho-protein
to total protein. The bar graph shows the fold change of the each protein band ratio compared with levels in control mice (represented by a line at 1-fold). All band densities were normalized to the density of GAPDH (Bars: mean ± s.e.; n=12/group; *=p<0.05 vs. control and #*=p<0.05 vs. denervation).

Figure 4. Acu-LFES increases macrophage expression and upregulates of cytokines.
(A & B) Total RNA was isolated from hindlimb muscles of control, Acu-LFES, denervation or denervation/Acu-LFES mice. Real time qPCR was used to assay for (A) general macrophage marker, F4/80, M1 macrophage marker, IL-1β and M2 macrophage marker, arginase-1, and (B) inflammation cytokines IL-6, IFNγ and TNFα. The bar graph shows mRNA levels in each group of muscles expressed as a fold change from the normal controls (line indicates controls set as 1-fold). Results are normalized to 18S mRNA (Bars: mean ± s.e.; n=12/group; *=p<0.05 vs. control and #*=p<0.05 vs. denervation).
(C) Plasma was obtained from normal control, Acu-LFES, denervation or denervation/Acu-LFES mice. Plasma cytokine levels were measured using a cytokine ELISA quantification assay. The bar graphs provide concentrations of eight cytokines compared with normal control levels (dashed lines). Bars: mean ± s.e.; n=9/group; *=p<0.05 vs. control and #*=p<0.05 vs. denervation.
(D) Muscle lysates were obtained from control, Acu-LFES, denervation or denervation/Acu-LFES mice. Muscle cytokine levels were measured by cytokine ELISA and normalized by protein concentration. The bar graphs provide concentrations of eight cytokines compared with normal control levels (dashed lines). Bars: mean ± s.e.; n=9/group; *=p<0.05 vs. control and #*=p<0.05 vs. denervation.

Figure 5: collagen accumulation appears in the muscle of Acu-LFES mice.
Muscle samples were collected 14 days after acupuncture. Muscle frozen sections were stained with Masson modified IMEB Trichrome to detect collagen deposition in the gastrocnemius muscle of normal control, Acu-LFES, denervation or denervation/Acu-LFES mice. The red color indicates muscle fiber; the blue indicates collagen and the black indicates nuclei. The bar graph compares the area (μm²) of blue staining per 500
muscle fibers in each group expressed as a fold-change from levels in control mice which is represented by 1 (Bars: mean ± s.e.; n=6/group).

Figure 6: Acu-LFES increase miR-1 and miR-206 microRNA in the muscle of control and denervated mice. Total RNA was isolated from hindlimb (gastrocnemius, soleus and plantaris) muscles of control, Acu-LFES, denervation or denervation/Acu-LFES mice, and then assayed for specific microRNA. The expressions of miR-1 (A) and miR-206 (B) were measured using real time qPCR with LNA™-enhanced oligonucleotide primers. The bar graph shows microRNA levels in each group expressed as a fold-change from levels in normal control mice which is represented by a line at 1-fold. All band densities were normalized to the density of U6 RNA (Bars: mean ± s.e.; n=9/group; *=p<0.05 vs. control and #=p<0.05 vs. denervation).
Table 1: Muscle and body Weights

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<td>Body weight (g)</td>
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Data are presented as mean ± s.e.; P < 0.05 is significant (*vs. control, #vs. denervation), n=12/group.
Table 2: primer sequence

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Figure 1
Figure 2
Figure 3
Figure 4A&B
Figure 4 (C+D)
Figure 5
**Figure 6**

(A) and (B) show the expression levels of miR-1 and miR-206, respectively, under different conditions: Acu-LFES, Denervation, and Denervation/Acu-LFES. The bars represent the fold change in expression compared to the control. Asterisks denote statistical significance: * p<0.05 vs. Control and # P<0.05 vs. denervation.