Peripheral vascular decoupling in porcine endotoxic shock

Feras Hatib, PhD,* Jos R Jansen, PhD,** and Michael R. Pinsky, MD***

*Edwards Lifesciences, LLC
Critical Care
Research and Development Department
Irvine, CA

** Department of Intensive Care Medicine
Leiden University Medical Center
The Netherlands

***Department of Critical Care Medicine
University of Pittsburgh School of Medicine
Pittsburgh, PA

Address correspondence to:
Michael R. Pinsky, MD
606 Scaife Hall
3550 Terrace Street
Pittsburgh, PA 15261
e-mail: pinskymr@upmc.edu

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Abstract
Cardiac output measurement from arterial pressure waveforms presumes a defined relationship between the arterial pulse pressure (PP), vascular compliance (C) and resistance (R). Cardiac output estimates degrade if these assumptions are incorrect. We hypothesized that sepsis would differentially alter central and peripheral vasomotor tone decoupling the usual pressure wave propagation from central to peripheral sites. We assessed arterial input impedance (Z), C and R from central and peripheral arterial pressures and aortic blood flow in an anesthetized porcine model (n=19) of fluid resuscitated endotoxic shock induced by endotoxin infusion (7 μg/kg/h increased to 14 and 20 μg/kg/h every 10 minutes and stopped when mean arterial pressure <40mmHg or SvO2 <45%). Aortic, femoral and radial artery pressures and aortic and radial artery flows were measured. Z was calculated by FFT of flow and pressure data. R and C were derived using a two-element Windkessel model. Arterial PP increased from aortic to femoral and radial sites. During stable endotoxemia with fluid resuscitation aortic and radial blood flows returned to or exceed baseline while mean arterial pressure remain similarly decreased at all three sites. However, aortic PP exceeded both femoral and radial arterial PP. Although Z, R and C derived from aortic and radial pressure and aortic flow were similar during baseline, Z increases and C decreases when derived from aortic pressure whereas Z decreases and C increases when derived from radial pressure, while R decreased similarly with both pressure signals. This central-to-peripheral vascular tone decoupling, as quantified by the difference in calculated Z and C from aortic and radial artery pressure may explain the decreasing precision of peripheral arterial pressure profile algorithms in assessing cardiac output in septic shock patients and suggests that different algorithms taking this vascular decoupling into account may be necessary to improve their precision in this patient population.
Introduction

Beat-to-beat estimates of left ventricular (LV) stroke volume and cardiac output from the arterial pressure waveform have been studied since the original work of Hamilton and Remington over 60 years ago (12). Such measurements of cardiac output from the arterial pressure waveform presume a defined relationship between the arterial pulse pressure (PP) profile, vascular compliance (C) and resistance (R) or that if they change, they do so in a uniform fashion. Such cardiac output estimates degrade if these assumptions are incorrect. Recently, commercial devices have evolved to report LV stroke volume and cardiac output from the direct measure of peripheral arterial pressure via an indwelling arterial catheter (3, 5). These devices are presently being used to aid in clinical decision making. However, concern exists as to the accuracy of these devices under conditions wherein peripheral vasomotor tone changes. The arterial pulse is created by LV stroke volume into the central arterial compartment as quantified by a transfer function (12). If arterial elastance and compliance remain constant, then central arterial PP will vary directly with LV stroke volume. Unfortunately, most indwelling arterial catheters sample a more peripheral arterial pressure signal, whose waveform may be altered in unexpected ways as arterial tone, pulse wave velocity and LV contractility vary. For this reason, it has been concluded that after a 1-hr calibration free period recalibration of these devices may be needed (13). Such repetitive recalibration may not be feasible in the clinical setting.

Continuous cardiac output measurement methodologies typically rely on a real-time analysis of the arterial waveform, with the assumption that changes in the waveform morphology can be related to corresponding changes in blood flow and central vasomotor tone. However, remarkable differences between aortic and peripheral arterial pressure have been reported in
patients during the periods shortly after return from cardiopulmonary bypass (22, 18). This resulted in significant outliers in pulse contour cardiac output estimates (14). Also, in the setting of septic shock, profound changes in the vasculature tone occur that may not be uniform across the entire arterial bed. The impact of these vascular tone changes on the accuracy of various arterial pressure contour devices to assess LV stroke volume when vascular reflected waves and central compliance change in unexpected ways is unknown. The relation between blood pressure and flow seen in the central arterial compartment may become dissociated in the periphery under conditions of severe vasomotor shock because of the profound alterations in peripheral vessel compliance and elastance induced by sepsis.

Circulatory changes in vasomotor tone altering vascular resistance and compliance appear to uniformly occur amongst different cardiovascular stress states, like hemorrhage, hypertension and heart failure and with the use of vasodilators and ACE inhibitors, because they all similarly modulate the adrenergic receptor response. Thus, mathematical models like those used by Pulsion for PiCCOplus®, LiDCO for LiDCOplus® and Edwards for FloTrac®, to calculate stroke volume from arterial pressure waveforms should remain stable in predicting flow from pressure across heart failure, hemorrhage and their therapies whether pressure is measured at a central or peripheral location. What is not known but highly relevant for the assessment of cardiovascular status in the patients with pathological vasoregulatory disease, like sepsis and end-stage liver failure. Prior work have documented the patients with sepsis may have both pathological vasodilation, owing to the excess synthesis of endogenous vasodilators, like nitric oxide, decreased adrenergic responsiveness, presumably due to receptor down-regulation, metabolic block or competing vasoactive substance counter signaling (1, 2, 6, 9, 29). Accordingly, the
accuracy of arterial pressure-derived flow devices can become inaccurate in these common clinical settings. To our knowledge, no analysis of the differential effect of sepsis or sepsis-like states on regional differences in vascular flow characteristics have been reported.

We hypothesized that when acute endotoxic shock in a porcine model resulted in cardiovascular collapse, this would be manifested by a change in vascular impedance characteristics which would decouple the normal pressure pulse propagation from central to peripheral vasculature, thus degrading the ability of peripheral arterial sensing pulse contour devices to estimate cardiac output. To address this issue, we assessed central and peripheral arterial pressure and aortic flow in an acute, anesthetized porcine model in which endotoxin shock was induced by the continuous infusion of low dose endotoxin followed by fluid resuscitation to restore cardiac output to pre-endotoxemic levels.

**Methods**

The experimental protocol for this study was approved by the Ethics Committee for experimental animal research at Edwards Lifesciences, LLC.

*Anesthesia and surgical preparation:* Nineteen female pigs (Yorkshire X) weighing between 75 and 107 kg (mean 89 ±11 kg) were fasted overnight, with free access to water. The animals were anesthetized with intramuscular injection of Telazol (4.4 mg/kg), Ketamine (2.2 mg/kg) and Xylazine (1.1 mg/kg). Neuromuscular blocking drugs were not used. The pigs were then orally intubated, and their lungs were mechanically ventilated (FiO₂ 0.4; positive end expiratory pressure 3 cmH₂O, Narkomed 2B, North American Dräger), with a tidal volume of 10 ml/kg, at a
respiratory rate 12-15 breaths/min, adjusted to maintain arterial PCO₂ between 4.77 to 5.33 kPa. Arterial blood gases were monitored on a periodic basis, and acid-base balance was corrected by infusing bolus sodium bicarbonate or by changing ventilator frequency, as needed. The other ventilator settings were kept unchanged throughout the experiment. The body temperature was maintained at 37 ±1 °C, using blankets and a heating pad. A catheter was placed in the urinary bladder through a cystotomy for measurement of urinary output. Following induction of anesthesia and endotracheal intubation, general anesthesia was maintained with Isoflurane (2%) for 15 min followed by a continuous intravenous infusion of Propranolol (1%) at 10 ml/kg/hour in 12 pigs and by continuous intravenous infusion of Penthotal (1%) at a rate of 15ml/kg/hour in the remaining 7 pigs.

The left external jugular vein was cannulated and a pulmonary artery catheter (Swan-Ganz® CCombo 777HF8, Edwards Lifesciences, Irvine CA) was introduced and by pressure guidance positioned in the pulmonary artery for monitoring continuous cardiac output (CCO) and mixed venous saturation (SvO₂). The positioning of the catheter was verified with fluoroscopy. A central venous catheter for drug and fluid infusion and for blood sampling was inserted into the superior vena cava. Ringer’s lactate solution (10 ml/kg/hr) was used as maintenance fluid balance. The left and right femoral veins were used for additional drug and fluid infusion during the resuscitation phase of the experiment as described below.

The left and right radial arteries as well as one femoral artery were surgically exposed. In one radial artery and in the femoral artery, fluid-filled catheters were placed for continuous blood pressure recording. A 1-1.5 mm diameter perivascular transit-time ultrasonic flow probe...
(Transonic System, Ithaca, NY) was placed around the other radial artery. Radial flow was continuously recorded (T206 flow meter, Transonic System, Ithaca, NY). A 5 French catheter was inserted using fluoroscopy through the carotid or the femoral artery into the aortic arch for continuous measurement of central aortic pressure. A midline sternotomy was performed, the pericardium separated and a 20-24 mm diameter perivascular transit-time ultrasonic flow probe (Transonic System, Ithaca, NY) was placed around the aortic root.

Measurements: The arterial radial, femoral and aortic catheters were connected to fluid filled direct pressure transducers (PXMK 1063, Edwards Lifesciences, Irvine CA) and radial, femoral and aortic arterial pressures together with an electrocardiogram for heart rate (HR), central venous pressure (CVP), pulmonary artery pressure (PAP) and respiration were measured continuously using a bedside monitor (IntelliVue MP70, Phillips, Boeblingen, Germany). In one experiment to determine if arterial pressure measures were accurately collected using fluid filled catheter, the exact study was repeated but using high fidelity pressure tip-mounted catheters (Mikro-tip®, Millar Instruments Inc., Houston, TX) to measure central aortic and radial arterial pressures. All other instrumentation was similar to that described above. A personal computer-based data acquisition system (DAQCard-AI-16XE-50, National Instruments, Austin, TX) was connected to the analog outputs of both the bedside monitor and the T206 flow meter. All the signals (radial, femoral and aortic pressures and radial and aortic flow signals) were digitized continuously at 500Hz and the data were saved to disc for off-line analysis.

CCO and SvO₂ were measured continuously using the Vigilance® monitor (Edwards LifeSciences, Irvine CA). Bolus injection cardiac output (ICO) was measured intermittently
every 2-3 hours, or as needed using the same monitor. The CCO, ICO and SvO2 data were data
logged to a laptop from the serial ports of the Vigilance monitor using data logger software and
the data were saved on a hard drive.

Experimental Protocol: After surgery, the animals were allowed to recover for a period of 30
minutes during which time no animal displayed arrhythmias, hypotension or on-going metabolic
acidosis. This stabilization period served as baseline for subsequent comparisons. After that the
30 minute stabilization period, an intravenous infusion of endotoxin (LPS E-Coli 055:B5, Sigma,
St. Louis MO) was started at a rate of 7 μg/kg/h and increased stepwise (7, 14, and 20 μg/kg/h)
every 10 min to a final rate of 20 μg/kg/h. The endotoxin infusion was discontinued when MAP
decreased below 40 mmHg, or when SvO2 decreased below 45%. This usually occurred 2 to 3
hours after the start of endotoxin infusion. Fluid resuscitation post-decompensation was
performed with intravenous infusion of 0.9% sodium chloride or lactated Ringers solution and
with 6% Dextran-70 at a rate of 20 mg/kg/hr. Fluids were given until CO ceased to increase. This
usually occurred when CO was between 8 and 18 l/min, with most of the animals reaching their
CO plateau at 10 l/min. The total amount of fluids that the animals received ranged from 10 to 20
liters.

Data Processing: The aortic flow and aortic, femoral and radial artery pressure data were
analyzed in the frequency domain as Fourier series using FFT. A representative example of the
pressure waveforms and Fourier analysis of the measured pressure and flow signals during
baseline and endotoxemic conditions is shown in figure 1. The power spectrum was then
computed and the impedance modulus at each harmonic was calculated as the ratio of the
respectively pressures and flow moduli at that harmonic. Input impedance \(Z\) was calculated using the mean impedance moduli for the 2\(^{nd}\) to 8\(^{th}\) harmonics (20):

\[
Z(\omega) = \frac{P(\omega)}{Q(\omega)}, \text{ for } \omega_2, \omega_3, \ldots \omega_8
\]  

(1)

Where:

- \(P(\omega)\) is the power spectrum of the arterial pressure signal
- \(Q(\omega)\) is the power spectrum of the flow signal
- \(Z(\omega)\) is the arterial input impedance
- \(f\) is the frequency
- \(\omega = 2\pi f\) is the angular frequency

The peripheral resistance (R) was defined as the ratio of pressure to flow at 0 Hz term of the spectrum.

The arterial system was modeled with a two-element Windkessel (11). The two elements of this model are the constant arterial compliance (C), representing elastic property of the arterial tree and a peripheral resistance (R), which drains the capacitor. The aortic flow curve (Q) together with one of the three pressure curves (P) was used as input of this model to characterize the parameters of the model.

The accompanying differential equation is:

\[
C \frac{d}{dt} P(t) + \frac{P(t)}{R} = Q(t)
\]  

(2)

In the frequency domain the corresponding input impedance is given by:
The reactive component $X_c(\omega)$ of the impedance of the Windkessel model was estimated based on the input impedance $Z(\omega)$ and the peripheral resistance estimated in the previous steps above:

$$X_c(\omega) = \frac{R \cdot Z(\omega)}{R - Z(\omega)}, \text{ for } \omega_2, \omega_3, \ldots, \omega_8 \tag{4}$$

Total arterial compliance, $C$, was then estimated by:

$$C = \frac{1}{\omega \cdot X_c(\omega)}, \text{ for } \omega_2, \omega_3, \ldots, \omega_8 \tag{5}$$

Theoretically, in the 2-element Windkessel model, $C$ should be the same throughout all harmonics. However, since the cardiovascular system cannot be simply modeled by a linear model and has also a transmission line of behavior an effect of reflected waves exists. So, $C$ is not consistent for all harmonics, and could either increase or decrease slightly with the increase of the harmonic number. Thus, the total $C$ was computed as the average compliance from the 2nd to 8th harmonics. These harmonics were chosen so that this analysis would be consistent with previous analyses of impedance (8,20).

Statistical analysis: Group mean data ±SD were derived for all the P, Q, R, C and Z parameters. Differences in parameters among animals and between control and endotoxic state within animals were compared by a three-way analysis of variance and a post hoc Student-Neuman-Keules test. Differences corresponding to a P value <0.05 were considered significant.

Results
All animals tolerated the surgical procedure well without any post-operative instability, arrhythmias or ongoing intrathoracic bleeding. Data for the one animal with pressure reading using the Millar catheters were indistinguishable from those of the other 19 animals. Radial arterial flow signals were highly variable with ripples, reflections and high frequency artifacts at baseline and in 8 animals displayed no flow during endotoxemia making analysis of radial arterial flow from these data impossible. We attribute these changes to the surgical trauma associated with isolation of the arteries during the initial instrumentation. Thus, radial arterial flow data are only reported for 11 of 19 animals and not used for any hemodynamic calculations.

The hemodynamic variables during the control and 30 minutes post-resuscitation in the endotoxic state are summarized in table 1 and the derived vascular Z, C and R parameters for those times are summarized in table 2. Figure 1 displays the regional pressure waveforms forms and Z for one animal for baseline and endotoxemic states. Figure 2 displays the time activity data for all measured variables for one animal and figure 3 displays its paired instantaneous calculated Z, R and C values.

Three primary findings are described in figure 2 and Table 1. First, at baseline PP progressively increases from aortic to femoral to radial arterial sampling sites. Although mean arterial pressure (MAP) was not different, < 1 mmHg, whereas the increases in PP were large (10-15 mmHg). The observed increase in PP is due primarily to an increase in systolic arterial pressure. Second, following the start of the endotoxin infusion, arterial hypotension starts to develop after about 20 minutes and is paralleled by a decrease in both aortic and radial arterial flow. Importantly, as MAP decreases aortic PP tends to remain constant whereas both femoral and radial arterial PP progressively decrease and at some point either at the end of decompensation or early during
fluid resuscitation aortic PP exceeds both femoral and radial arterial PP and remains so during resuscitation. The decrease in peripheral PP is due primarily to a greater decrease in systolic pressure in the peripheral sites as diastolic pressure tends to decrease to a similar amount at all sites. Third, during fully developed endotoxic shock following fluid resuscitation aortic and radial arterial blood flows return and usually exceed baseline values while all measures of MAP remain decreased, thus characterizing the hypotensive hyperdynamic septic shock state. Furthermore, during this fully developed hypotensive hyperdynamic state aortic PP markedly exceeds both femoral and radial arterial PP to levels similar to that seen in the opposite direction during control conditions. These dynamic trends shown for one animal in this example persisted across all animals (Table 1).

Analysis of Z, R and C from aortic pressure and aortic flow and from radial artery pressure and aortic flow during the evolution of hypotensive hyperdynamic shock reveals striking time and loci-specific changes (Figure 3, Table 2). During baseline Z and R is similar with both aortic and radial pressures as input of the model whereas calculated C tends to be less with peripheral pressures as input, though not significantly so. However, with the induction of endotoxic shock, the calculated Z separates increasing with aorta pressure as input and decreasing with the radial arterial pressure as input. Since flow and mean arterial pressure are similar for both measurement sites R though decreasing remains common in both central and peripheral sites. Accordingly, endotoxemia induces a dissociation in calculated C from central and peripheral pressure, such that the central compartment behaves as if it is getting stiffer (less compliant) whereas with the peripheral compartment becomes more compliant. These differences in Z and C with central and peripheral pressures become even more apparent following fluid resuscitation (Table 3).
This study of acute porcine endotoxemia has two main findings. First, like other studies of acute endotoxic shock associated with fluid resuscitation previously described by us, we report the development of a hypotensive hyperdynamic state (23, 24, 25). Furthermore, though not shown, arterial blood gas analysis revealed the progressive onset of metabolic acidosis. Thus, we believe our model of acute porcine endotoxic shock is representative of the previously described pathophysiology. It is within the differences between baseline and developed hyperdynamic hypotension that our analyses focus. Second, we describe the novel finding of a switch in the usual amplification of the arterial PP from central to peripheral vascular loci with the development of acute endotoxic shock. The etiology of this PP reversal appears to be due to a differential effect of acute endotoxemia on the central and peripheral derived vascular compliance. Presumably the endotoxin-induced expression of vascular endothelial inducible nitric oxide synthetase (iNOS) results in pathological peripheral vasodilation (30). If this were the case, why then do we only see peripheral loss of vascular stiffness despite a generalized decrease in Ohmic resistance? Our study does not allow us to answer that question. However, since central aortic vessels have little smooth muscle relative to their vascular walls whereas peripheral vascular has much reactive smooth muscle architecture it is reasonable that they may be differentially affected by vasodilating substances during endotoxemic shock. Since stroke volume is constant across the arterial tree, if one could simultaneously measure both central and peripheral arterial PP, noting dynamic changes in their ratios would suggest changes in regional vascular compliance. Unfortunately, dual measures of central and peripheral PP are rarely available in clinical practice.
Technical limitations: Extrapolation of these data to clinical studies should be done with caution because an acute animal preparation under general anesthesia and challenged with an endotoxin infusion for only two to three hours is only a rough approximation of an awake critically ill patient in circulatory shock with varying levels of sympathetic tone and pre-existing cardiovascular abnormalities and associated exogenous vasopressors infusions. Still, this model has been widely used to assess the circulatory state of humans in septic shock since pigs have a cardiovascular response similar to that of humans. Thus, we predict that if similar measures of central and peripheral pressures and flow are made in patients in septic shock similar directional changes in peripheral vascular compliance and loss of PP amplification will be seen. Accordingly, to the extent that any acute animal model is useful in understanding human pathophysiology, these data should be relevant. In this study we used fluid filled catheters to measure P and calculated Z. To the extent that over damping or under-damping of these catheters occur or their frequency response is inadequate to assess the pressure harmonics, our reported changes in Z may be suspect. But as noted in the methods, in one animal we used high-fidelity pressure-tipped catheters to measure central aortic and peripheral P and found identical MAP, PP and Z values as with the other animals. Thus, these data can collectively be shown to reflect their purported changes in regional vascular tone.

We used fluid-filled catheters to measure arterial pressure instead of pressure-tipped catheters. The Edwards Direct Pressure Transducer has a flat frequency response up to 100Hz. The pressure catheters used to measure redial, femoral and aortic pressure were short (60 in) and stiff. The natural frequency of the catheter-tubing-transducer units ranged from 42Hz to 45Hz with the
damping coefficient of 0.16, which was a better than the frequency response than previously reported (8, 20). This response is considered adequate, as most of the energy of the signal is contained in the first 7 harmonics (20).

Another potential limitation was the use of colloid for fluid resuscitation with a mixture of crystalloid and colloid solutions which could alter blood viscosity. We specifically used a 3:1 volume ratio of crystalloid to colloid so as to minimize any change in blood viscosity due to fluid resuscitation. Although our method minimized changes in blood viscosity relative to pure crystalloid resuscitation, we did not measure blood viscosity in these studies. Still, our resuscitation parallels clinical practice making these findings as consistent as possible to the clinical setting. Finally, decreases in arterial pressure during acute endotoxemia could independently alter central arterial compliance independent of endotoxin-induced changes in vasomotor tone owing to the curvi-linearity of the central vascular compliance relation. But, if anything, systemic hypotension would under-fill the aorta increasing its compliance, and we observed a marked decrease in central aortic compliance during endotoxemia. Thus, the changes we report cannot be explained by differences in mean arterial pressure.

In our data analysis we used a linear two-element Windkessel model, although arterial compliance has been reported non-linearly related to arterial pressure (16, 17, 26) and heart rate (15). However, the use of a linear compliance instead of a nonlinear compliance suffices in most Windkessel models (10). Therefore, we chose the more simple approach with a linear Windkessel model. Different methods exist to estimate arterial compliance. A recent review showed nine methods based in the Windkessel approach (33). The most important methods are; the decay time method (11); the stroke volume over PP method (7); the area methods (17, 27);
the parameter estimation method (31) and the impedance method in the frequency domain (8, 19, 21, 34). We used the generally accepted, robust and well described impedance method. Furthermore, we used a two-element Windkessel model for the whole heart beat. This model is known for its good properties to describe the flow-pressure relation for the diastolic part of the heart beat. However, it has been shown that this model is inadequate to describe the flow-pressure relation during the systolic part of a heart beat (4). A more accurate model is a three element Windkessel model, as has been used in model flow pulse contour (32). Still, using the three element model though reporting cardiac output more accurately, would not fundamentally change the conclusion that resistance remains similar but impedance and compliance differentially change with pressure measured at different loci during sepsis.

Clinical implications: These data have important clinical relevance in the use of arterial waveform analyses to derive estimates of CO in critically ill patients. Presently, all commercially-available devices that estimate cardiac output from the arterial PP do so based on a series of assumptions regarding aortic compliance and PP or pulse power transmission to the periphery. Although our study did not attempt to study the accuracy of any of these devices in this model, it is clear that if our data reflect the potential degree of vascular compliance and impedance change that can rapidly occur during sepsis. Thus, algorithm calibration based on healthy vasculatures without recalibration following significant non-homogeneous vascular tone changes may display a reporting bias, either under or over estimating true CO. The degree of bias by each of the commercially-available devices is unknown. Furthermore, devices that use only internal calibration approaches or one initial calibration and then follow CO trends over hours may be prone to systematic measurement errors due to regional changes in vascular reactivity.
There are two potential solutions to this calibration mismatch. First, as recommended by at least one manufacturer, if clinical conditions rapidly change, a recalibration is required. Recall that in our study the profound changes in impedance and compliance occurred only after the development of profound hypotension, not during the initial stages of changing pressures and flow. Thus, the clinical signs of a significant vascular change should be similarly obvious and alert the bedside clinician to recalibrate the device. Second, once vascular decoupling occurred in our animal preparations, it remained remarkably constant up until the terminal event. Thus, if one could characterize the specific dynamic qualities of the decoupled vasculature one could create a second algorithm to match the decoupled state and use it for the calculation of CO, thus minimize the reporting bias. One would need to know, however, when the decoupled state occurred, its specific characteristics and when it resolved in order for such a two-algorithm approach to be used effectively. It is unknown however, if different patients existing in this decoupled vascular state would display similar enough flow characteristics to allow a single second algorithm to apply for all. But this two algorithm approach would be a simpler solution to the vascular decoupling effect of estimating CO from arterial pressure recordings.
Figure legends

Figure 1. An example of the Fourier Analysis for one animal of central aortic flow (QAo) and the three measured arterial pressures, aortic pressure (PAo), femoral pressure (Pfemoral) and radial pressure (Pradial) for one animal under baseline (left) and endotoxic shock (right) conditions.

Figure 2. A time series display of all the measured hemodynamic variables for one animal from the initial post-operative baseline through endotoxin infusion (red vertical line and horizontal arrow) and resuscitation (green vertical line and horizontal arrow). Abbreviations: AOR, aorta; FEM, femoral; RAD, radial; MAP, mean arterial pressure; SYS, systolic arterial pressure; DIA, diastolic arterial pressure; PP, pulse pressure. See text for discussion.

Figure 3. A time series display of all the calculated aortic and radial arterial calculated hemodynamic variables for the same animal as in figure 2 from the initial post-operative baseline through endotoxin infusion (red vertical line and arrow) and resuscitation (green vertical line and arrow). Abbreviation: AOR, aorta; RAD, radial; Z, impedance (mm Hg·min·L⁻¹); R, resistance (mm Hg·min·L⁻¹); C, compliance (ml·mm Hg⁻¹). See text for discussion.

Table legends

Table 1. Pressures are in mmHg, heart rate in beats/min and cardiac output in L·min⁻¹.

Table 2. Z, Arterial input impedance; R, resistance; C, compliance.
Table 3. Pressures in mm Hg, Z, input impedance (mmHg·min·L⁻¹); R, resistance (mmHg·min·L⁻¹); C, compliance (ml·mmHg⁻¹), abbreviations as in Table 2.
References:


### TABLE 1: Hemodynamic Data (n=19)

<table>
<thead>
<tr>
<th></th>
<th>Baseline mean ± SD</th>
<th>Endotoxemia mean ± SD</th>
<th>ANOVA Baseline to Endotoxemia</th>
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<tbody>
<tr>
<td><strong>Pulse Pressure</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aortic</td>
<td>30.7 ± 6.2</td>
<td>44.2 ± 13.5</td>
<td>p=0.0001; F=23.7</td>
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<tr>
<td>Femoral</td>
<td>40.9 ± 7.0</td>
<td>27.3 ± 13.0</td>
<td>p=0.001; F=15.7</td>
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<tr>
<td>Radial</td>
<td>43.2 ± 9.0</td>
<td>33.8 ± 20.4</td>
<td>p=0.08; F=3.2</td>
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<tr>
<td><strong>Mean Arterial Pressure</strong></td>
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<td></td>
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<tr>
<td>Aortic</td>
<td>75.4 ± 17.7</td>
<td>50.5 ± 16.0</td>
<td>p&lt;0.0001; F=32.6</td>
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<td>Femoral</td>
<td>76.4 ± 18.0</td>
<td>47.8 ± 17.4</td>
<td>p&lt;0.0001; F=37.0</td>
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<tr>
<td>Radial</td>
<td>76.2 ± 16.9</td>
<td>47.0 ± 18.4</td>
<td>p&lt;0.0001; F=40.4</td>
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<td><strong>Systolic Pressure</strong></td>
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<tr>
<td>Aortic</td>
<td>99.0 ± 21.2</td>
<td>80.3 ± 22.9</td>
<td>p=0.003; F=11.5</td>
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<tr>
<td>Femoral</td>
<td>106.6 ± 23.3</td>
<td>65.5 ± 25.0</td>
<td>p&lt;0.0001; F=37.0</td>
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<tr>
<td>Radial</td>
<td>108.2 ± 20.4</td>
<td>69.1 ± 30.7</td>
<td>p&lt;0.0001; F=33.1</td>
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<td><strong>Diastolic Pressure</strong></td>
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<tr>
<td>Aortic</td>
<td>68.2 ± 20.4</td>
<td>34.6 ± 12.3</td>
<td>p&lt;0.0001; F=67.2</td>
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<tr>
<td>Femoral</td>
<td>66.3 ± 20.2</td>
<td>36.8 ± 10.9</td>
<td>p&lt;0.0001; F=58.6</td>
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<tr>
<td>Radial</td>
<td>65.1 ± 20.5</td>
<td>35.4 ± 12.4</td>
<td>p&lt;0.0001; F=40.8</td>
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<tr>
<td><strong>Heart Rate</strong></td>
<td>85.4 ± 15.2</td>
<td>126.5 ± 23.2</td>
<td>p&lt;0.0001; F=53.7</td>
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<tr>
<td><strong>Cardiac Output</strong></td>
<td>6.7 ± 1.5</td>
<td>8.3 ± 3.6</td>
<td>p=0.08; F=3.4</td>
</tr>
</tbody>
</table>

All pressures in mmHg, heart rate in beats·min⁻¹, and cardiac output in L·min⁻¹.
### TABLE 2: Derived Arterial Vascular Parameters (n=19)

<table>
<thead>
<tr>
<th></th>
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<th>Baseline mean ± SD</th>
<th>Endotoxemia mean ± SD</th>
<th>ANOVA Baseline to Endotoxemia</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Input Pressure</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Z</td>
<td>Aortic</td>
<td>1.5 ± 0.6</td>
<td>2.7 ± 1.2</td>
<td>p=0.0003; F=23.5</td>
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<tr>
<td></td>
<td>Radial</td>
<td>1.3 ± 0.4</td>
<td>1.0 ± 0.6</td>
<td>p=0.03; F=5.8</td>
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<tr>
<td>R</td>
<td>Aortic</td>
<td>10.7 ± 3.3</td>
<td>6.9 ± 3.5</td>
<td>p=0.004; F=11.9</td>
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<tr>
<td></td>
<td>Radial</td>
<td>10.4 ± 3.2</td>
<td>6.4 ± 3.3</td>
<td>p=0.004; F=11.8</td>
</tr>
<tr>
<td>C</td>
<td>Aortic</td>
<td>34 ± 11</td>
<td>20 ± 18</td>
<td>p=0.02; F=5.8</td>
</tr>
<tr>
<td></td>
<td>Radial</td>
<td>30 ± 18</td>
<td>79 ± 64</td>
<td>p=0.01; F=8.9</td>
</tr>
</tbody>
</table>

Z, input impedance (mmHg·min·L⁻¹); R, resistance (mmHg·min·L⁻¹); and C, compliance (ml·mmHg⁻¹).
TABLE 3: The Effect of Endotoxemia on the difference between aortic and radial measured and derived variables (n=19)

<table>
<thead>
<tr>
<th></th>
<th>Baseline Aortic-Radial mean ± SD</th>
<th>Endotoxemia Aortic-Radial mean ± SD</th>
<th>ANOVA Baseline to Endotoxemia</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pulse Pressure</td>
<td>-12.4 ± 5.0</td>
<td>10.4 ± 15.6</td>
<td>p=0.0001; F=26.1</td>
</tr>
<tr>
<td>Mean Arterial Pressure</td>
<td>-0.8 ± 1.7</td>
<td>3.5 ± 4.1</td>
<td>p=0.0003; F=19.3</td>
</tr>
<tr>
<td>Z</td>
<td>0.1 ± 0.7</td>
<td>1.7 ± 1.4</td>
<td>p=0.0002; F=27.6</td>
</tr>
<tr>
<td>R</td>
<td>0.3 ± 0.7</td>
<td>0.4 ± 0.5</td>
<td>p=1.7; F=0.2</td>
</tr>
<tr>
<td>C</td>
<td>4.6 ± 14</td>
<td>-60 ± 50</td>
<td>p=0.009; F=9.4</td>
</tr>
</tbody>
</table>

Pressures in mm Hg, Z, input impedance (mmHg·min·L⁻¹); R, resistance (mmHg·min·L⁻¹); C, compliance (ml·mmHg⁻¹), abbreviations as in Table 2
Fourier Analysis of Aortic Flow and Arterial Pressures

Arterial Pressure

[f, Hz]  [t, sec]  [mmHg]

Baseline

Endotoxic Shock

$Q_{Ao}$

$P_{Ao}$

$P_{femoral}$

$P_{radial}$
Fig. 2
Fig. 3