**PLASMA ARGinine-vasopressin following experimental Stroke: Effect of osmotherapy**

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**Cover Title:** AVP in experimental Stroke
ABSTRACT

Neurohumoral responses have been implicated in the pathogenesis of ischemia-evoked cerebral edema. In a well-characterized animal model of ischemic stroke, the present study was undertaken to 1) study the profile of plasma arginine-vasopressin (AVP), and 2) to determine if osmotherapy with mannitol and various concentrations of hypertonic saline (HS) solutions influence plasma AVP levels. Halothane-anesthetized adult male Wistar rats were subjected to 2 hr of middle cerebral artery occlusion (MCAO) with the intraluminal filament technique. Plasma AVP levels (pg/ml; mean ± SD) were significantly elevated at 24 hr (42 ± 21), 48 hr (50 ± 28), and 72 hr (110 ± 47), and returned to baseline at 96 hr (22 ± 15) following MCAO as compared to sham-operated controls (14 ± 7). Plasma AVP levels at 72 hr were significantly attenuated with 7.5% HS (37 ± 8 pg/mL; 360 ± 11 mOsm/L) as compared to 0.9% saline (NS; 73 ± 6; 292 ± 6 mOsm/L), 3% HS (66 ± 8 pg/mL; 303 ± 12 mosm/L), or mannitol (74 ± 9 pg/mL; 313 ± 14 mOsm/L) treatment. 7.5% HS significantly attenuated water content in the ipsilateral and contralateral hemispheres as compared to surgical shams, NS, 3% HS and mannitol treatments. Peak plasma AVP levels were not associated with direct histopathologic injury to the anterior hypothalamus. Attenuation of brain water content with 7.5% HS treatment coincides with attenuated serum AVP levels, and we speculate that this may represent one additional mechanism by which osmotherapy attenuates edema associated with ischemic stroke.

Key Words: Osmotherapy, Mannitol, Hypertonic saline, Arginine-Vasopressin, Ischemic stroke
Cerebral edema represents a major cause of morbidity and mortality in patients with ischemic stroke, predominantly from lethal intracranial compartmental shifts that result in herniation syndromes and compromise vital brainstem function (10, 15, 16, 29, 32). Over the past several decades, osmotherapy has remained the cornerstone of medical therapy for cerebral edema (10, 28, 31). Acute administration of osmotic agents produces a potent anti-edema action, predominantly on undamaged brain regions with an intact blood brain barrier (BBB), theoretically causing rapid egress of water from the interstitial and extracellular space into the intravascular compartment, resulting in improved intracranial elastance (10, 15, 16, 29, 31, 37, 49). In addition to causing “dehydration” of the brain, osmotic agents exert beneficial non-osmotic cerebral effects, such as augmentation of cerebral blood flow (CBF) (30, 38), modulation of inflammatory molecules (4, 27), scavenging of free radicals, and modulation of cerebrospinal fluid dynamics (formation and reabsorption) (29).

A pathophysiologic role for neurohumoral factors, specifically the nonapeptide arginine-vasopressin (AVP) secreted by the hypothalamo-neurohypophyseal system, has been implicated in a variety of brain injury paradigms including subarachnoid hemorrhage (SAH) (21, 22) and traumatic brain injury (19). Relatively few studies have reported this relationship in ischemic stroke (5, 6). Small (parvocellular) vasopressin-containing neurons in the anterior hypothalamus are known to give rise to a complex fiber system that extends throughout the brain (11, 27). The existence of extrahypophyseal vasopressinergic pathways within the brain has been demonstrated and may allow for independent release and function of central versus systemic AVP (8). AVP influences non-neuronal brain cells (glia), by regulating water balance via adjustment of astrocytic water permeability (27). Recent immunohistochemical and autoradiographic studies
have revealed that its presence is robust in brain and is affected by changes in plasma osmolarity (5, 6, 8, 11, 17, 27, 41).

We have shown that maintenance of a hyperosmolar state with continuous intravenous (IV) infusion of hypertonic saline (HS) ameliorates cerebral edema associated with experimental ischemic stroke (46, 47). Although the serum osmolarity in patients with acute brain injury is recommended to be 300–320 mOsm/L (34, 35, 45), the optimal serum osmolarity required to exert the most effective osmotic gradient for the anti-edema action with specific osmotic agents remains under investigation. Emerging evidence from experimental studies suggests that a serum osmolarity of > 350 mOsm/L for the treatment of ischemia-evoked cerebral edema may be more beneficial (12, 47).

In the present study, using a well-characterized model of transient focal ischemia, we sought to determine 1) the temporal profile of plasma AVP, and 2) the effect of osmotherapy on plasma AVP levels. Furthermore, we studied the effects of induction and maintenance of a graded hyperosmolar state with HS and mannitol therapy on stroke volume and ischemia-evoked cerebral edema.

MATERIALS AND METHODS

General preparation and animal surgery. The experimental protocol was approved by the Institutional Animal Care and Use Committee and conforms to the National Institutes of Health guidelines for the care and use of animals in research. All techniques are as previously described (46, 47). In brief, adult male Wistar rats (280–320 g; n=159) were anesthetized with halothane (1.0–2.0%) in oxygen-enriched air and allowed to ventilate spontaneously. Using aseptic surgical techniques, the right femoral artery was cannulated to monitor arterial blood pressure and arterial
blood gases, and the femoral vein was cannulated for vascular access. After cannulation, both catheters were exteriorized in the posterior mid-thorax and suture-fixed on to a swivel at the mid-scapular region. This tethering system allows the animal to move freely following emergence from anesthesia while receiving continuous infusion of drugs and desired solutions with a programmable infusion pump at a desired rate. Rectal temperatures (37.5–38.5°C) were maintained with a heating lamp throughout the surgical procedures, and the animals were housed in separate cages at ambient room temperature following emergence from anesthesia.

**Focal ischemia and assessment of stroke volume.** Transient focal ischemia was produced by MCAO (2 hr) using an intraluminal suture technique, as previously described (24), with modifications (3, 46, 47). Adequacy of vascular occlusion and reperfusion was determined by laser-Doppler flowmetry (LDF) over the ipsilateral parietal cortex, as previously described (3, 46, 47) (coordinates: 2 mm posterior and 6 mm lateral to bregma) at baseline, during MCAO, and at 30 min of reperfusion. On successful reperfusion, rats were allowed to emerge from anesthesia in separate cages and given free access to food and water. Rats that did not demonstrate a significant reduction of the LDF signal (≤ 40% of baseline) during MCAO were not included in the final analysis. This degree of reduction in LDF signal is necessary to ensure consistent infarction volume in our model of MCAO (3). Similarly, rats that did not demonstrate return of LDF signal on reperfusion were not included in the final analysis. At 72 hr of reperfusion, rats were deeply anesthetized with 5% halothane and decapitated. The brain was harvested and sliced into seven 2-mm thick coronal sections for staining with 1% triphenyltetrazolium chloride (TTC), as previously described (3). Infarct volume was measured with digital imaging. The infarcted area was numerically integrated across each section and over the entire ipsilateral hemisphere.
Infarct volumes were measured separately in cerebral cortex and caudoputamen (CP) complex and expressed as a percentage of the contralateral structure volume (corrected for swelling), as previously described (23, 46, 47).

**Histopathology**

Brains from rats in the first series of experiments (0.9% saline-treated at 72 hr of reperfusion) were examined by light microscopy. After TTC staining was complete, the coronal slabs containing hypothalamus were cryoprotected in 30% sucrose in phosphate buffer and section at 40 µm on a sliding (freezing) microtome. Every sixth section through the hypothalamus was mounted and stained with hematoxylin and eosin (H&E). Selected adjacent sections were stained with cresyl violet. The distribution of infarct in the H&E and cresyl violet sections corresponded very closely to the distribution of infarction by TTC staining.

**Assessment of plasma osmolarity, electrolytes and brain edema.** Rats were killed at the end of the experiment by decapitation under deep halothane (5%) anesthesia. A sample of blood (1.0 mL) was drawn by cardiac aspiration. A 0.5 mL sample was sent to the institutional core laboratories for determination of serum electrolytes (sodium, potassium, urea nitrogen and creatinine) and a 0.5 mL sample processed to determine plasma osmolarity (mOsm/L) with a centrifuge and an automated freezing point depression micro-osmometer (Advanced Instruments, Inc., Norwood, MA) as previously described (23, 46, 47). The brain was quickly removed and gently blotted to remove small quantities of adsorbent moisture and dissected through the interhemispheric fissure into ipsilateral and contralateral hemispheres. Brain edema was estimated by comparing wet to dry weight ratios (23, 46, 47). Tissues were weighed with a scale to within 0.1 mg. Dry weight of the entire ipsilateral and contralateral hemispheres was
determined after heating the tissue for 3 days at 100°C in a drying oven. Tissue water content was then calculated as % H₂O = (1-dry wt/wet wt) x 100% (23, 46, 47).

**Assessment of Plasma AVP levels.** At sacrifice, a sample of blood was obtained and analyzed for AVP in the plasma as previously described (40), with modifications utilizing the commercially available DSL-1800 AVP radioimmunoassay (RIA) kit (Diagnostic Systems Laboratories, Inc., Webster, TX).

**Experimental groups.** In the first series of experiments, rats were killed at 24, 48, 72, and 96 hr (n = 10 each) following MCAO, and plasma AVP levels were determined. Surgical shams that underwent all surgical procedures including neck surgery without MCAO were used as controls (n = 5). All rats received a continuous IV infusion of 0.3 mL/hr of 0.9% saline (NS; 308 mOsm/L). In the second series of experiments, rats subjected to transient MCAO were treated in a blinded randomized fashion to receive a continuous IV infusion of 0.3 mL/hr of NS (n = 12), 3% HS (1027 mOsm/L; n = 13), 7.5% HS (2310 mOsm/L; n = 10) or 2 g/Kg of 20% mannitol (1098 mOsm/L; n = 12) IV bolus every 6 hr. Rats treated with mannitol were given a continuous IV infusion of NS (0.3 ml/hr). HS was instituted as a mixture of acetate:chloride (50:50; pH = 6.5–7.0) to avoid hyperchloremic acidosis. Treatments were started at 6 hr of reperfusion and continued until 72 hr of reperfusion. In a third series of experiments, rats were subjected to 2 hr MCAO and treated in a blinded randomized fashion with a continuous IV infusion of 0.3 mL/hr of NS (n = 15), 3% HS (n = 12), 7.5% HS (n = 11) or 20% mannitol (n = 13). Treatments were begun at 6 hr of reperfusion and brain edema determined at 72 hr of reperfusion. Surgical shams were used as controls (n=10). In all three series of experiments, rats were allowed to emerge from anesthesia at 30 min of reperfusion. Rats were housed in separate cages at room
temperature (22–24°C) and during emergence from anesthesia and thereafter until they were euthanized.

**Statistical Analysis.** All values are expressed as means ± SD. Physiologic parameters and mean LDF measurements among groups were subjected to repeated measures analysis of variance (ANOVA). Differences in plasma AVP levels and infract volume among treatment groups were determined by one-way ANOVA with post hoc Newman Keuls test. Mortality rates were compared with logistic regression analysis. The criterion for statistical significance was \( p < 0.05 \).

**RESULTS**

In the first series of experiments, physiologic parameters, including mean arterial blood pressure (MABP), pH, arterial carbon dioxide (\( P_aCO_2 \)), oxygen (\( P_aO_2 \)), and rectal temperatures were within normal ranges in all experimental groups at baseline, during MCAO, and at 30 min of reperfusion (*Table 1*). LDF signal during MCAO was not different among different treatment groups (*Table 1*). Mortality rates prior to completion of the experiment were as follows: 0/5 in surgical-sham controls, 3/13 in rats for 24-hr endpoint, 5/15 in rats for 48-hr endpoint, 6/16 in rats for 72-hr endpoint, and 2/12 in rats for 96-hr endpoint. Plasma AVP levels were significantly elevated at 24 hr (42 ± 21 pg/mL; \( n = 10 \)), 48 hr (50 ± 28 pg/mL; \( n = 10 \)), and 72 hr (110 ± 47 pg/mL; \( n = 9 \)), but not at 96 hr (22 ± 15 pg/mL; \( n = 10 \)) following MCAO, as compared to sham-operated controls (14 ± 7; \( n = 5 \)) (*Table 1*). Plasma osmolarity was not different among experimental groups (*Table 1*). TTC-determined infarct volume (corrected for brain swelling) was not different among experimental groups, and plasma AVP levels did not demonstrate correlation with infarct volume (data not shown). Histopathologic examination of the hypothalamus in rats (\( n = 4 \)) treated with NS for 72 hr of reperfusion did not demonstrate any
neuronal injury in anterior hypothalamic nuclei associated with AVP production (paraventricular nuclei, supraoptic nuclei, suprachiasmatic nuclei, medial nuclei of the diagonal band of Broca) (Figure 1), although there were varying degrees of ischemic damage in the lateral hypothalamus.

In the second series of experiments, physiologic parameters (MABP, pH, $P_aCO_2$, $P_aO_2$, and rectal temperatures) were within normal ranges in all experimental groups at baseline, during MCAO, and at 30 min of reperfusion (data not shown). Mortality rates prior to completion of the experiment were as follows: 3/12 in rats treated with NS, 3/13 in rats treated with 3% HS, 4/12 in rats treated with mannitol, and 1/10 in rats treated with 7.5% HS. One rat with NS treatment, 2 with 3% HS, and 1 with 7.5% HS treatment did not meet the LDF criterion for successful MCAO or reperfusion. Thus, 8 rats in each of the experimental groups were included in the final analysis. Average LDF-signal reduction from baseline during MCAO was not different among experimental groups (NS: 28 ± 9%; 3% HS: 27 ± 10%; 7.5% HS: 27 ± 9%). At the completion of treatments (72 hr of reperfusion), plasma AVP levels were significantly attenuated with 7.5% HS (360 ± 11 mOsm/L), as compared to NS (292 ± 6 mOsm/L), 3% HS (303 ± 12 mOsm/L), or mannitol (313 ± 14 pg/mL) treatment (Figure 2). TTC-determined infarct volume, corrected for brain swelling, was significantly attenuated in CP complex in rats treated with 7.5% HS (cortex: 44 ± 6%; CP: 59 ± 6%), as compared to NS (cortex: 44 ± 8%; CP: 78 ± 5%), 3% HS (cortex: 39 ± 8%; CP: 66 ± 6%), and mannitol (cortex: 38 ± 8%; CP: 70 ± 5%), but there were no differences in total hemispheric infarct volume in various treatment groups (NS: 20 ± 5%; 3% HS: 19 ± 4%; 7.5% HS: 17 ± 4%; 20% mannitol: 18 ± 4%).

In the third series of experiments, physiologic parameters (MABP, pH, $P_aCO_2$, $P_aO_2$, and rectal temperatures) were within normal ranges in all experimental groups at baseline, during
MCAO, and at 30 min of reperfusion (data not shown). Mortality rates prior to the end of the experiment (between 48 and 72 hr; surgical shams – 0/10; NS – 5/15; 20% mannitol – 3/13; 3% HS – 2/12; 7.5% HS – 1/11) were significantly higher in NS-treated rats as compared to surgical shams. Thus 10 rats in each treatment group were included in the final analysis in this series of experiment. All rats met LDF criterion for successful MCAO. Water content in the ipsilateral and contralateral hemispheres was comparable in surgical sham controls. Treatment with 7.5% HS attenuated cerebral edema in both hemispheres at 72 hr of reperfusion (serum osmolarity 358 ± 20 mOsm/L) compared to that achieved with NS (308 ± 8 mOsm/L), 20% mannitol (336 ± 14 mOsm/L) and 3% HS (332 ± 12 mOsm/L) (Figure 3). Serum sodium was significantly elevated with 7.5% HS treatment while potassium, urea nitrogen, and creatinine were within normal limits in all treatment groups (Table 2).

**DISCUSSION**

This study demonstrates two novel and important findings: 1) A distinct profile in plasma AVP levels following experimental focal ischemia that is independent of brain injury volume; and 2) institution and maintenance of a hyperosmolar state with HS to levels, well beyond those suggested and reported in the literature, attenuate ischemia-evoked brain edema as well as lower plasma AVP levels. Furthermore, our histopathologic studies did not demonstrate direct injury to the anterior hypothalamus at the time when peak plasma levels of AVP are demonstrated. These data may have implications for the role of AVP in ischemic stroke and highlight the interaction of osmotherapy with plasma AVP levels following large ischemic strokes.

**Cerebral edema following ischemic stroke.** We previously demonstrated (46, 47) that increases in water content in the ipsilateral as well as contralateral hemispheres following large
experimental ischemic stroke are responsive to treatment with HS at concentrations used in the present study. The mechanisms of cerebral edema following focal ischemia are complex. In addition to the classic, simplistic pathobiology of ischemia-evoked cerebral edema that includes a cytotoxic component (secondary to post-ischemic energy failure) and a vasogenic component (secondary to breakdown of the BBB) (10, 15, 16, 29, 32), other mechanisms are under investigation. Some of these include impedance of cerebral venous return from cerebral swelling; intrahemispheric diaschisis or hypometabolism (2, 25, 36); role of inflammatory mediators (1, 15); neurohumoral responses, including AVP release (5, 6, 11, 17, 25, 41); induction of proteins, such as the vascular endothelial growth factor (VEGF) (48); and upregulation of water channels, predominantly aquaporin-4 (44).

**AVP in ischemic stroke.** In the present study, we focused our attention on neurohumoral responses and the role of AVP in the pathophysiology of ischemic stroke. AVP is robustly present in the brain and regulates water balance by adjusting water permeability in glial cells (5). Several lines of evidence suggest that AVP may play an important role in events that follow cerebral ischemia. Elevated AVP levels have been reported in experimental ischemia (5), as well as in the serum and cerebrospinal fluid (CSF) from patients with ischemic stroke (6, 41). Intracerebroventricular (ICV) injections of AVP exacerbate acute ischemic brain edema, while ICV injection of AVP antiserum significantly decreases cerebral edema (17). It has been suggested that AVP may play an important role in inhibiting the \( \text{Na}^+ - \text{K}^+ \) ATPase activity of the cerebral cell membrane via AVP receptors mediated by cAMP and cGMP (17). Indirect evidence with the use of pharmacologic agents that inhibit AVP release, support a therapeutic target for the treatment of cerebral edema. For example, the kappa-opioid receptor agonist
RU51599 and AVP inhibitor with potent aquaretic activity, characterized by pure water diuresis, reduces brain edema following experimental stroke (20), and a selective nonpeptide V₁ receptor antagonist (OPC-21268) has been shown to reduce cerebral edema associated with cold lesion (8), which simulates traumatic brain injury. We did not discern the source of AVP in our model of focal ischemia; however, we did not demonstrate any direct neuronal injury to the anterior hypothalamic nuclei which are the major source of this hormone. Vasopressin-containing neurons in the anterior hypothalamus give rise to a complex intrinsic fiber system that can modulate aquaporin-mediated water flux and hence play a crucial role in brain water and ion homeostasis (27).

In the present study, AVP levels were significantly elevated and continued to rise to a maximum at 72 hr following focal ischemia. This temporal profile corresponds to and coincides with the maximal cerebral edema typically seen 48–72 hr following ischemic stroke in our animal model (46, 47). While previous studies have demonstrated that the degree of elevation in AVP is proportional to the severity of ischemic brain injury (6), we did not observe a correlation between infarct volume and plasma AVP levels in our study. Our histopathologic studies did not demonstrate direct injury to the anterior hypothalamus, suggesting that there are other sources of AVP following ischemic brain injury.

**Osmotherapy and AVP.** HS solutions are being increasingly utilized clinically as a therapeutic modality for cerebral edema in a variety of brain injury paradigms (10, 16, 30, 31, 37–40, 49). Owing to its better toxicity profile, and because sodium chloride (reflection coefficient = 1.0) (13) is completely excluded from brain with an intact BBB, it has been proposed that HS may be a more favorable osmotic agent than the conventional agent, mannitol (reflection coefficient =
0.9). Furthermore, HS may be a more desirable agent for maintaining a “euvolemic hyperosmolar” state in a variety of brain injury paradigms (10, 16, 46, 47). We have previously demonstrated that HS therapy, when instituted at the onset of reperfusion following transient focal ischemia (2 hr), worsens TTC-determined infarct volume at the 24-hr endpoint (9), but attenuates cerebral edema when treatment is delayed for 24 hr following experimental stroke (46, 47). While we have no direct evidence for mechanism(s) responsible for this set of observations, the mechanism of this detrimental effect was not due to changes in regional CBF (9). Little is known about the differential response of neurons and glia to HS solutions during an “evolving” cerebral infarction. We have postulated that a hyperosmolar state impedes the recovery of neurons from ischemia during the early phases of its evolution. This is based on in vitro studies that have demonstrated that hypertonic-hyperoncotic saline differentially affects healthy and glutamate-injured hippocampal neurons and astrocytes (18). There may be competing effects of HS solutions in ischemic stroke, and the beneficial osmotic effects on stroke-associated cerebral edema may be dependent on timing of the onset and duration of therapy in relation to “maturation” of the lesion following ischemic stroke. Based on our the observed beneficial effects of HS when therapy was delayed from the onset of ischemia (46,47) we instituted osmotherapy at 6 hr of MCAO and continued treatment until the period of peak plasma levels of AVP (72 hr). Continuous 3% HS and 7.5% HS therapy were utilized to maintain a constant osmotic gradient to cause egress of water from the brain. In contrast to our previous study (9), while total hemispheric infarct volume was not different at 72 hr of reperfusion, injury was significantly attenuated in the deep subcortical regions (CP complex) with 7.5% HS therapy in the present study. We did not measure regional CBF in our experiments but it is plausible that
HS accentuates regional CBF in the subcortical regions during delayed reperfusion, thereby attenuating injury in the CP complex. These results also suggest that outcomes with the use of osmotic agents depend on timing of onset and duration of treatment following ischemic stroke. While the literature recommends that serum osmolarity be raised to 300–320 mOsm/L in patients with brain injury (34, 45), we have demonstrated (44) beneficial effects on ischemia-evoked brain edema with HS therapy with plasma osmolarity of > 350 mOsm/L. Although we did not discern any differences in mortality rates in our experimental groups, ischemia-evoked cerebral edema was significantly attenuated in the ipsilateral and contralateral cerebral hemispheres, more significantly so with 7.5% HS compared to mannitol and 3% HS treatments. In the present study, we observed attenuation of plasma AVP levels only with 7.5% HS therapy (plasma osmolarity in the range of 350–360 mOsm/L). We utilized 3% HS to create a graded hyperosmolar state and as a control for 20% mannitol because the two solutions have comparable osmotic load (1027 versus 1098 mOsm/L, respectively). While our study did not discern the mechanisms of attenuated levels of AVP with 7.5% HS treatment, we speculate that institution of osmotherapy with target plasma osmolalities > 350 mOsm/L in our well-characterized model of ischemic stroke, attenuates brain and plasma AVP levels, which consequently leads to decreased cerebral edema.

Our study has some limitations. Although we instituted IV fluids to ensure euvolemia in our experiments, we have limited measures of hydration status in our animal model. We cannot comment on the diuretic effects of HS because we did not monitor urinary output in our rat model. It is plausible that HS has diuretic effects, resulting from attenuated plasma AVP levels at the higher serum osmolalities. However, urea nitrogen and creatinine levels were within normal
limits in various treatment groups indicating no systemic dehydration in any of the experimental groups. We did not assess behavioral outcomes in our study because of technical considerations as a result of the tethering system for institution of continuous IV infusions in our experimental model. We cannot comment on the possible rebound effects on cerebral edema following cessation of osmotherapy in our experimental paradigm. During prolonged elevation of plasma osmolarity, with ongoing osmotherapy for brain edema over days, excess brain electrolytes are replaced by organic solutes (“idiogenic osmoles”; myo-inositol, taurine, glycercylphosphorylcholine, and betaine) for volume regulation and cotransported along with sodium from the extracellular to the intracellular compartment (29,43). This osmotic compensation as well as penetration and accumulation of osmotic agents into the brain tissue particularly in regions of disrupted BBB may explain prolonged cerebral edema and delayed “rebound edema” when osmotherapy is withdrawn (29). While this phenomenon has been well described with mannitol use, it has not been studied with HS therapy but is theoretically possible. We did not measure regional brain AVP levels, and it is plausible that extra-CNS sites of AVP production are reflected in our data.

CONCLUSIONS

In conclusion, our data demonstrate that plasma AVP levels are elevated following ischemic stroke, with peak levels corresponding to the time period of maximal ischemia-evoked cerebral edema. Osmotherapy to target levels > 350 mOsm/L attenuates plasma AVP levels that may further augment anti-edema effects. The precise mechanism(s) of the role of AVP and clinical significance of agents that attenuate its release following ischemic stroke require further study for the treatment of accompanying cerebral edema.
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REFERENCES


FIGURE LEGENDS

Figure 1. Cresyl violet staining of hypothalamus demonstrating relationship of the largest studied MCA infarct to hypothalamic nuclei. A) A coronal section through the forebrain shows the paraventricular nuclei separated by the third ventricle. The control side is on the left. B) The supraoptic nucleus on the left (control) side is shown along with the adjacent optic tract. C) The neurons in the supraoptic nucleus on the right (ischemic) side show normal neuronal morphology, although there are reactive cells in the adjacent neuropil.

Figure 2. Plasma AVP (mean ± SD) levels in rats treated with 0.9% saline (NS; n = 8), 3% HS (n = 8), 20% mannitol (n = 8), and 7.5% HS (n = 8). Treatments were initiated at 6 hr and continued until 72 hr of reperfusion following 2 hr MCAO. *p < 0.05 versus NS, 3% HS, and mannitol treatment.

Figure 3. Water content in the ipsilateral (ischemic) and contralateral (non-ischemic) hemispheres in surgical sham controls (n=10) and rats subjected to 2 hr MCAO and treated with 0.9% saline (NS) (n=10), 3% HS (n=10), 20% mannitol (n=10), and 7.5% HS (n=10). Treatments were initiated 6 hr of reperfusion and continued for 72 hr. * and † p < 0.01 versus corresponding hemisphere in experimental groups that received NS. ¶ and # p < 0.05 versus corresponding hemisphere in rats treated with mannitol.
Table 1. Physiologic variables at baseline, during ischemia (2 hr MCAO), and 30 min of reperfusion in various treatment groups

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<th>Surgical Shams (n=10)</th>
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<td><strong>Rectal Temperature (°C)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>37.2 ± 0.9</td>
<td>37.1 ± 1.2</td>
<td>37.5 ± 1.3</td>
<td>37.6 ± 1.2</td>
<td>37.4 ± 0.9</td>
</tr>
<tr>
<td>Ischemia</td>
<td>38.3 ± 0.8</td>
<td>38.4 ± 1.1</td>
<td>38.3 ± 1.2</td>
<td>38.3 ± 1.1</td>
<td>38.3 ± 1.1</td>
</tr>
<tr>
<td>Reperfusion</td>
<td>38.1 ± 1.0</td>
<td>38.0 ± 0.9</td>
<td>37.8 ± 0.8</td>
<td>38.0 ± 1.1</td>
<td>37.9 ± 1.2</td>
</tr>
<tr>
<td><strong>Blood Glucose (mg/dL)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>88 ± 20</td>
<td>87 ± 18</td>
<td>85 ± 22</td>
<td>100 ± 24</td>
<td>99 ± 16</td>
</tr>
<tr>
<td>Ischemia</td>
<td>82 ± 28</td>
<td>88 ± 26</td>
<td>84 ± 32</td>
<td>94 ± 20</td>
<td>82 ± 22</td>
</tr>
<tr>
<td>Reperfusion</td>
<td>94 ± 32</td>
<td>84 ± 28</td>
<td>86 ± 30</td>
<td>95 ± 18</td>
<td>95 ± 30</td>
</tr>
<tr>
<td><strong>Serum Osmolarity (mOsm/L)</strong></td>
<td>290 ± 9</td>
<td>286 ± 10</td>
<td>284 ± 12</td>
<td>288 ± 9</td>
<td>294 ± 14</td>
</tr>
<tr>
<td><strong>Plasma AVP (ng/mL)</strong></td>
<td>14 ± 7</td>
<td>42 ± 21*</td>
<td>50 ± 28*</td>
<td>110 ± 47*</td>
<td>22 ± 15</td>
</tr>
</tbody>
</table>

Values are presented as mean ± SD; MABP = mean arterial blood pressure. *p < 0.05 versus surgical sham controls.
Table 2. Serum electrolytes in various experimental groups at 72 hr of reperfusion following 2 hr MCAO.

<table>
<thead>
<tr>
<th></th>
<th>NS</th>
<th>Mannitol</th>
<th>3% HS</th>
<th>7.5% HS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Urea nitrogen (mg/dl)</td>
<td>15.8 ± 8.2</td>
<td>15.5 ± 3.0</td>
<td>15.0 ± 3.4</td>
<td>24.0 ± 7.7</td>
</tr>
<tr>
<td>Creatinine (mg/dl)</td>
<td>0.22 ± 0.04</td>
<td>0.22 ± 0.04</td>
<td>0.24 ± 0.05</td>
<td>0.30 ± 0.09</td>
</tr>
<tr>
<td>Sodium (mEq/l)</td>
<td>148 ± 4*</td>
<td>144 ± 4*</td>
<td>147 ± 4*</td>
<td>172 ± 13</td>
</tr>
<tr>
<td>Potassium (mEq/l)</td>
<td>4.7 ± 0.4</td>
<td>4.1 ± 0.6</td>
<td>4.2 ± 0.5</td>
<td>4.3 ± 0.5</td>
</tr>
</tbody>
</table>

Data in mean ± SD; * P < 0.001 vs. 7.5% HS

Normal range: BUN: 12-25.8 mg/dL; creatinine: 0.39-2.29 mg/dL; sodium: 129-150 mEq/L; potassium: 4.6-6.0 mEq/L (33).
Figure 2

Plasma AVP (pg/mL)

- NS
- Mannitol
- 3% HS
- 7.5% HS

* Indicates significant difference.
Figure 3

![Graph showing water content in different groups of animals. The graph compares Ipsilateral Hemisphere and Contralateral Hemisphere for Surgical Sham, Mannitol, 3% HS, and 7.5% HS controls.](image-url)