Does exercise-induced hypoxemia modify lactate influx into erythrocytes and hemorheological parameters in athletes?

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Submitted 12 September 2003; accepted in final form 27 April 2004

Connes, Philippe, Didier Bouix, Guillaume Py, Corinne Caillaud, Pascale Kippelen, Jean-Frédéric Brun, Alain Varray, Christian Prefaut, and Jacques Mercier. Does exercise-induced hypoxemia modify lactate influx into erythrocytes and hemorheological parameters in athletes? J Appl Physiol 97: 1053–1058, 2004.—This study investigated 1) red blood cells (RBC) rigidity and 2) lactate influxes into RBCs in endurance-trained athletes with and without exercise-induced hypoxemia (EIH). Nine EIH and six non-EIH subjects performed a submaximal steady-state exercise on a cycle-ergometer at 60% of maximal aerobic power for 10 min, followed by 15 min at 85% of maximal aerobic power. At rest and at the end of exercise, arterialized blood was sampled for analysis of arterialized pressure in oxygen, and venous blood was drawn for analysis of plasma lactate concentrations and hemorheological parameters. Lactate influxes into RBCs were measured at three labeled [U-14C]lactate concentrations (1.6, 8.1, and 41 mM) on venous blood sampled at rest. The EIH subjects had higher maximal oxygen uptake than non-EIH (P < 0.05). Total lactate influx was significantly higher in RBCs from EIH compared with non-EIH subjects at 8.1 mM (1,498.1 ± 254.5 vs. 1,145.0 ± 264.4; P < 0.05) and 41 mM (2,562.0 ± 145.0 vs. 1,618.1 ± 149.4; P < 0.01). Monocarboxylate transporter-1-mediated lactate influx also was higher in EIH at 8.1 mM (r = 0.82, P < 0.05) and 41 mM (r = 0.84, P < 0.05) in the two groups together. Plasma lactate concentrations and hemorheological data were similar in the two groups at rest and at the end of exercise. The results showed higher monocarboxylate transporter-1-mediated lactate influx in the EIH subjects and suggested that EIH could modify lactate influx into erythrocyte. However, higher lactate influx in EIH subjects was not accompanied by an increase in RBC rigidity.

mono-carboxylate transporter; endurance; lactate metabolism; hypoxemia; hemorheology

The functional significance of the hypoxia-induced changes is likely an increase of lactate and H+ fluxes from plasma to erythrocyte. During sea-level exercise, some endurance-trained athletes experience arterial hypoxemia [exercise-induced hypoxemia (EIH)] that can be defined as a decrease in both oxygen arterial partial pressure (PaO2) and arterial hemoglobin saturation during exercise (8). Miyachi and Katayama (21) have reported repeated episodes of EIH during training sessions when endurance athletes performed very intense exercise, a result suggesting that athletes develop frequent episodes of mild hypoxemia during training. One can thus think that repeated mild hypoxemia stimulus may lead to increased expression or activity of band 3 and MCT-1 proteins on the red blood cell (RBC) membranes. An increase in lactate and H+ fluxes into RBCs during exercise may also alter the deformability of these cells, because lactate uptake by RBCs increases their rigidity (32). These changes in RBC deformability could, in turn, alter blood rheological properties and then participate to EIH. Several studies have suggested that blood viscosity and its determinants, i.e., RBC rigidity and Hct, could be involved in pulmonary diffusion impairment (6, 35). First, pulmonary diffusing capacity is widely affected by RBC deformability. When deformability is artificially reduced by chemical treatment, diffusion capacity may decrease by 20% (4). Second, high blood viscosity or RBC rigidity causes higher mechanical stress on the blood-gas barrier and may result in membrane disruptions. Aguilaniu et al. (1) showed that 6 wk of a polyunsaturated fatty acid (PUFA) diet led to an upward shift in PaO2 in master athletes. PUFA regimen is known to improve RBC deformability and to reduce blood viscosity (15); however, hemorheological measurements were not performed in this study (1).

In the present study, we hypothesized that EIH subjects have 1) a higher capacity for lactate influx in RBCs and 2) a reduced RBC deformability and increased blood viscosity during exercise. To test this hypothesis, we compared RBC lactate influx characteristics at rest in subjects with and without EIH, and we studied the hemorheological profiles at rest and at the end of a submaximal exercise bout.

MATERIALS AND METHODS

Subjects

Fifteen male endurance-trained subjects (runners, cyclists, and triathletes) were assigned to one of two groups (EIH or non-EIH) by

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assessing the decrease in oxyhemoglobin saturation [oxyhemoglobin saturation measured by pulse oximetry (SpO₂)], during a first exercise test. A minimum drop of 3–4% in SpO₂ for at least the last three steps of an incremental exercise test is required to conclude that the drop is significant (25). ΔSpO₂ was calculated as the difference between SpO₂ measured at rest and during the last step of the incremental exercise. Group assignment was also confirmed by the decrease in PaO₂, i.e., ΔPaO₂, measured during a subsequent submaximal test. Nine subjects with EIH were included in the EIH group (24 ± 1 yr, 73.4 ± 2.2 kg, 180 ± 3 cm, ΔPaO₂ = −12.40 ± 1.17 Torr, ΔSpO₂ = −4.9 ± 1.1), and six subjects without EIH were assigned to the non-EIH group (26 ± 2 yr, 72.7 ± 2.2 kg, 182 ± 2 cm, ΔPaO₂ = −2.50 ± 1.50 Torr, ΔSpO₂ = −1.9 ± 0.4).

The major exclusion criteria were tobacco use and muscle, joint, and cardiorespiratory diseases.

**Experimental Protocol and Procedures**

The local ethics committee approved the study, and each subject gave informed, written consent to participate. The subjects performed two exercise tests on a cycle ergometer (Ergoline type) separated by 1 wk. The first one was a progressive and maximal exercise test to determine the maximal cardiorespiratory parameters and SpO₂. The second was a submaximal steady-state exercise test. Arterialized blood was sampled at rest and at the end of exercise for analysis of PaO₂, and venous blood was sampled at the same times for analysis of plasma lactate concentrations ([Lac]) and hemorheological parameters. Lactate influxes into RBCs were measured on blood sampled at rest.

**Exercise tests.** The incremental maximal exercise test began with a 5-min warm-up at 60% of VO₂max, a 25°C, 10 min, 2000 g), Plasma and buffy coat were removed by aspiration. Thirty volumes of chloride buffer [150 mM NaCl and 10 mM sodium tricine, pH 8.0, at 37°C, osmolality ~315 mosmol/kgH₂O, volume (ml) = 30 × 2.5 × pre-Hct] were added to the pellet, which was mixed by inversion and incubated in a water bath for 30 min at 37°C to ensure complete removal of endogenous lactate (10, 30, 31). After incubation, the RBCs were sedimented at room temperature (25°C, 10 min, 2000 g), and the supernatant was removed by aspiration. The cell pellet was then washed twice with chloride buffer and suspended in a volume of HEPES buffer (90 mM NaCl, 50 mM HEPES, pH 7.4, 37°C, osmolality ~267 mosmol/kgH₂O) equivalent to a 30%Hct level (packed cell volume) to obtain the stock cell suspension for influx measurements. This suspension was divided into two tubes, each containing 1 ml. One of the two tubes contained no lactate transport blockers. The second tube contained 1 mM of p-chloromercuribenzenesulfonic acid (PCMB), which is known to inhibit the monocarboxylate-specific carrier at this concentration (30, 31). The tubes were then incubated in a water bath for an additional 30 min at 37°C. Part of the stock cell suspension was used for Hct determination (post-Hct).

All samples for lactate influx measurements were run in triplicate. At time t = 0, a 25-μl sample of stock cell suspension was added to a 13 × 100-mm test tube containing 75 μl of HEPES influx buffer at 37°C. The HEPES influx buffer contained 1-[U-14C]lactate (sodium salt, specific activity 50 μCi/mmol) at three [Lac] values of 2, 10, and 50 mM. The HEPES influx buffer was adjusted to pH 7.4. Because of dilution with the stock cell solution, the actual [Lac] values were 1.6, 8.1, and 41 mM. The cells were exposed to the influx buffer and mixed at 37°C for 20 s (30, 31). At that time, 5 ml of an ice-cold stop solution [150 mM NaCl, 10 mM sodium-2-(N-morpholino)ethanesulfonic acid, pH 6.5] were then added to each test tube to stop influx. The sample was spun for 15 min at 2,000 g and 4°C, and the supernatant was removed by aspiration. Control blanks were run in duplicate at the three [Lac] values for both total and MCT-1-mediated lactate uptake, to correct for any residual extracellular radioactivity and for any transmembrane lactate exchange that might have occurred, despite ice-cold stop solution. An additional wash phase was conducted by adding 5 ml of stop solution to each sample. The cells

**Hemorheology.** Seven milliliters of venous blood were collected in Vacutainer tubes (Becton Dickinson) containing EDTA as the anticoagulant for the hemorheological measurements. Hct was measured by the micromethod after blood centrifugation. Measurements of blood viscosity and plasma viscosity were performed with a falling ball viscometer (MT 90 Medicatex, Saint Benoit, France) (12). The coefficient of variation for this method ranged from 0.6 to 0.8% (14).

Plasma was obtained via centrifugation at 2,000 g and 4°C for 5 min (Jouan, Saint Nazaire, France). The index of RBC rigidity (Tk) was calculated according to the equation of Dintenfass (11).

\[
\mu_b = \mu_p [1 - (Tk \cdot Hct)]^{-2.5}
\]

where \(\mu_b\) is blood viscosity (mPa/s), \(\mu_p\) is plasma viscosity (mPa/s), and Hct is in percent.

**Plasma [Lac].** Venous blood (5 ml) was sampled in heparinized tubes (Becton Dickinson). Plasma was obtained via centrifugation at 2,000 g and 4°C for 5 min in a refrigerated centrifuge (Jouan). Plasma was then isolated and frozen at −80°C until assay. Plasma [Lac] was determined by using an enzymatic method (Roche Diagnostics kit, Mannheim, Germany).

**RBC lactate transport.** For lactate influx into RBCs, 5 ml of venous blood were collected at rest in heparin tubes (heparin, 0.2 U/ml), stored in ice, and prepared before lactate influx measurements. The techniques for RBC preparation and lactate influx measurement were modified from previously published methods (30, 31). The initial Hct (pre-Hct) was determined for all blood samples. One-half (2.5 ml) of each blood sample was transferred to a 50-ml conical tube, depleted of lactate, and washed by using the following procedure. First, the RBCs were isolated by centrifugation at room temperature (25°C, 15 min, 2000 g). Plasma and buffy coat were removed by aspiration. Thirty volumes of chloride buffer [150 mM NaCl and 10 mM sodium tricine, pH 8.0, at 37°C, osmolality ~315 mosmol/kgH₂O, volume (ml) = 30 × 2.5 × pre-Hct] were added to the pellet, which was mixed by inversion and incubated in a water bath for 30 min at 37°C to ensure complete removal of endogenous lactate (10, 30, 31). After incubation, the RBCs were sedimented at room temperature (25°C, 10 min, 2000 g), and the supernatant was removed by aspiration. The cell pellet was then washed twice with chloride buffer and suspended in a volume of HEPES buffer (90 mM NaCl, 50 mM HEPES, pH 7.4, 37°C, osmolality ~267 mosmol/kgH₂O) equivalent to a 30%Hct level (packed cell volume) to obtain the stock cell suspension for influx measurements. This suspension was divided into two tubes, each containing 1 ml. One of the two tubes contained no lactate transport blockers. The second tube contained 1 mM of p-chloromercuribenzenesulfonic acid (PCMB), which is known to inhibit the monocarboxylate-specific carrier at this concentration (30, 31). The tubes were then incubated in a water bath for an additional 30 min at 37°C. Part of the stock cell suspension was used for Hct determination (post-Hct).

All samples for lactate influx measurements were run in triplicate. At time t = 0, a 25-μl sample of stock cell suspension was added to a 13 × 100-mm test tube containing 75 μl of HEPES influx buffer at 37°C. The HEPES influx buffer contained 1-[U-14C]lactate (sodium salt, specific activity 50 μCi/mmol) at three [Lac] values of 2, 10, and 50 mM. The HEPES influx buffer was adjusted to pH 7.4. Because of dilution with the stock cell solution, the actual [Lac] values were 1.6, 8.1, and 41 mM. The cells were exposed to the influx buffer and mixed at 37°C for 20 s (30, 31). At that time, 5 ml of an ice-cold stop solution [150 mM NaCl, 10 mM sodium-2-(N-morpholino)ethanesulfonic acid, pH 6.5] were then added to each test tube to stop influx. The sample was spun for 15 min at 2,000 g and 4°C, and the supernatant was removed by aspiration. Control blanks were run in duplicate at the three [Lac] values for both total and MCT-1-mediated lactate uptake, to correct for any residual extracellular radioactivity and for any transmembrane lactate exchange that might have occurred, despite ice-cold stop solution. An additional wash phase was conducted by adding 5 ml of stop solution to each sample. The cells
were spun again, and the supernatant was removed. The RBC pellet was lysed and deproteinized with 0.5 ml of 4.2% perchloric acid followed by centrifugation of the sample for 15 min (2,000 g) at 4°C. A 0.4-ml sample was then placed into scintillation vials containing 5 ml of aqueous counting fluid and counted in a liquid scintillation counter (Tri-Carb Liquid Scintillation Analyzer, model 2200).

Total lactate influx into the RBCs was determined at the three [Lac] values (1.6, 8.1, and 41 mM). For this determination, neither the stock cell solution nor the HEPES influx buffer contained any lactate transport blocker. Influx into PCMBs-treated RBCs was measured at the same three concentrations of lactate. Because PCMBs treatment inhibits the monocarboxylate pathways, influx into the PCMBs-treated RBCs was the sum of lactate transport by the band 3 pathway and nonionic diffusion. Therefore, total lactate influx minus influx into the PCMBs-treated cells represents the MCT-1-mediated lactate influx. Control blanks radioactivity was subtracted from the radioactivity measured in samples.

The percentage of contribution from the MCT-1 pathway was calculated by dividing MCT-1-mediated lactate influx by the corresponding total lactate influx. Lactate influx was calculated in nanomoles of lactate per milliliter of cells (erythrocyte) per minute: the 25 μl of stock solution were multiplied by its Hct fraction (0.30) to obtain a packed cell volume. Total lactate influxes into RBCs and MCT-1-mediated lactate influxes were determined.

Statistical Analysis

Values are presented as means ± SE. Subject characteristics and VO₂max values were compared between groups by using an unpaired Student’s t-test. Lactate influxes into RBCs were compared with a two-way ANOVA with repeated measures: two groups (EIH subjects and non-EIH subjects) × three [Lac] values (1.6, 8.1, and 41 mM). The relationships between the drop in PaO₂ (ΔPaO₂) and the total lactate influx values and between VO₂max and the total lactate influx values were evaluated for each subject by using a Pearson correlation. Hemorheological parameters and plasma [Lac] were compared by using a two-way ANOVA: two groups × sample time (at rest or at the end of submaximal exercise). Pairwise contrasts were used when necessary to determine where the significant differences occurred. The level of significance was set at α = 0.05.

RESULTS

VO₂max

VO₂max was significantly higher in the EIH subjects (65.33 ± 2.09 ml·kg⁻¹·min⁻¹) compared with the non-EIH subjects (60.27 ± 0.85 ml·kg⁻¹·min⁻¹) (P < 0.05).

Hemorheological Parameters

The values of the hemorheological parameters were identical in the two groups at rest (Table 1). During exercise, blood viscosity, plasma viscosity, and Hct increased significantly in both groups (P < 0.001) and reached similar values. Neither a group nor an exercise effect was observed for RBC rigidity.

### Table 1. Hemorheological parameters at rest and at the end of submaximal exercise in EIH and non-EIH subjects

<table>
<thead>
<tr>
<th></th>
<th>EIH Subjects</th>
<th>Non-EIH Subjects</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Rest</td>
<td>End of exercise</td>
</tr>
<tr>
<td>Tm, °C</td>
<td>33.1±0.13</td>
<td>38.3±0.17*</td>
</tr>
<tr>
<td>TM, mPa/s</td>
<td>1.37±0.01</td>
<td>1.46±0.01*</td>
</tr>
<tr>
<td>Hct, %</td>
<td>42.1±0.8</td>
<td>45.9±0.9*</td>
</tr>
<tr>
<td>Tk, mPa/s</td>
<td>0.69±0.02</td>
<td>0.69±0.01</td>
</tr>
</tbody>
</table>

Values are means ± SE. EIH, exercise-induced hypoxemia; Tm, blood viscosity; TM, plasma viscosity; Hct, hematocrit; Tk, red blood cell rigidity coefficient. *Significantly different between rest and the end of exercise (P < 0.001).

### Fig. 1. Total lactate influx into red blood cells (RBCs) from 2 groups at different lactate concentration ([Lac]) values. Values at 8.1 and 41 mM were higher in exercise-induced hypoxemia (EIH) subjects. Values are means ± SE. *P < 0.05; tP < 0.001.

Plasma [Lac]

Plasma [Lac] values were similar for both groups at rest: 2.15 ± 0.11 and 2.32 ± 0.19 mM for EIH and non-EIH subjects, respectively. At the end of submaximal exercise, the plasma [Lac] was also identical in the two groups: 4.88 ± 1.12 and 5.32 ± 1.34 mM for EIH and non-EIH subjects, respectively.

Lactate Influx into RBCs

Total lactate influxes into RBCs are illustrated in Fig. 1. At 1.6 mM [Lac], no difference between groups was observed. Total lactate influx was greater in the EIH subjects at 8.1 mM (P < 0.05) and 41 mM [Lac] (P < 0.001) compared with the other group. MCT-1-mediated lactate influx was higher in the EIH subjects at 8.1 mM (P < 0.05) and 41 mM [Lac] (P < 0.001; Fig. 2).

The fractional contribution of MCT-1 to total lactate influx decreased with external [Lac] (Table 2).

### Table 2. Relationships between ΔPaO₂ and total lactate influx values

<table>
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<tr>
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<th>EIH Subjects</th>
<th>Non-EIH Subjects</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Rest</td>
<td>End of exercise</td>
</tr>
<tr>
<td>VO₂max, %</td>
<td>0.84±0.05</td>
<td>0.82±0.05</td>
</tr>
</tbody>
</table>

Values are means ± SE. ΔPaO₂, change in arterial oxygen tension.

DISCUSSION

This study mainly showed that, in the resting condition, total and MCT-1-mediated lactate influxes into RBCs were higher in the EIH subjects at 8.1 and 41 mM [Lac]. In addition, there was a strong correlation between ΔPaO₂ and total lactate influx. The hemorheological profile was the same in the two groups at rest.
and at the end of exercise. These results suggest that EIH can modify lactate fluxes into RBC without any significant change in cell deformability.

Lactate Influx

We have observed higher total lactate influx into the RBCs from EIH subjects at moderate (8.1 mM) and high [Lac] (41 mM) than in the other group. We are mindful that the use of another [Lac], as done by Skelton et al. (30), closer to the physiological range would have reinforced our results. From Figs. 1 and 2, we can reasonably think that the use of a [Lac] around 16 mM would have given similar results, i.e., higher total or MCT-1-mediated lactate influx in EIH subjects. The results obtained at the tested concentrations showed in EIH subjects an increased influx via MCT-1, the major pathway of lactate influx into RBCs (30). This could be related to the greater aerobic physical fitness in EIH subject. Indeed, Skelton et al. (31) showed that the lactate influx via MCT-1 was faster in RBC obtained from animals with high-oxidative capacity (dogs and horses) than from animals with low-oxidative capacity (goats and cattle). However, in the present study, we did not find any significant relationship between $V'O_2\max$ and total lactate influx. Lactate uptake by RBCs may, nevertheless, play a role in enhancing physical performance. The accumulation of lactate and hydrogen ions in muscle during exercise may inhibit muscular function and cause fatigue (13, 16). High-RBC uptake of these ions could help in establishing a gradient between plasma and interstitial fluid (9, 28). Transport of these metabolites away from the exercising muscles into the blood would reduce the potential for muscle acidosis and delay the onset of fatigue.

Recently, Juel et al. (18) found that band 3 and MCT-1 expressions on RBC membranes from humans were increased after 2 and 8 wk of hypoxia exposure. RBC MCT-1 and band 3 contents increased dramatically (+330 and +150%, respec-

Table 2. Fractional contribution (%) of the monocarboxylate transporter-1 pathway to total lactate influx at three lactate concentrations

<table>
<thead>
<tr>
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<th>1.6 mM</th>
<th>8.1 mM</th>
<th>41 mM</th>
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<tbody>
<tr>
<td>EIH subjects, %</td>
<td>89.7±0.6</td>
<td>92.5±1.4</td>
<td>80.8±4.6</td>
</tr>
<tr>
<td>Non-EIH subjects, %</td>
<td>93.8±3.7</td>
<td>88.0±0.8</td>
<td>70.2±3.2</td>
</tr>
</tbody>
</table>

Values are means ± SE. No significant difference was observed between groups at any concentration.
example, Vandewalle et al. (34) observed higher plasma con-
increased to the same value in both groups at the end of
logical parameters involved in blood viscosity regulation had
depends on several parameters, including plasma viscosity,
more speci-
ned that the MT 90 viscometer measures with a shear
rate of 1,000 s⁻¹. Based on the Dintenfass equation, it was thus
possible to calculate Tk in our study because this Tk becomes
expression of the proteins involved or their activity. The
underlying physiological mechanism is still unknown but may
be related to erythropoietin (Epo) secretion. In a recent study
(7), our laboratory showed that 4 wk of treatment with recom-
binant human Epo led to a signi-
expression might also explain why Juel et al. (18) found an
increase in MCT-1 expression on RBC membrane but did not
find such a change in muscle samples. Indeed, to our knowl-
edge, there is no Epo receptor in mature human muscular cells.

**Hemorheological Measurements**

Blood viscosity was assessed by using a falling ball viscom-
eter, the MT 90 Medicatex. This method was shown to be valid
for the assessment of both plasma and blood viscosities in
humans (12, 14). Moreover, Doffin et al. (12) specifically
demonstrated that the MT 90 viscometer measures with a shear
rate of 1,000 s⁻¹. Based on the Dintenfass equation, it was thus
possible to calculate Tk in our study because this Tk becomes
more specific as the shear rate rises (11). Blood viscosity
depends on several parameters, including plasma viscosity,
Hct, and RBC rigidity (11, 26). In our study, the hemorhoe-
ological parameters involved in blood viscosity regulation had
increased to the same value in both groups at the end of
exercise, and thus blood viscosity also showed the same
increase. This increase in plasma viscosity during exercise
could be related to a rise in plasma protein concentration. For
example, Vandewalle et al. (34) observed higher plasma con-
centrations of α1-globulins, α2-globulins, β-globulins, and
γ-globulins at the end of a moderate exercise.

Several factors could be responsible for the change in Hct
during exercise, including fluid shift, water loss, and RBCs
released from the spleen (17, 19, 22, 33), but we did not
determine the relative contribution of each factor in this study.

RBC rigidity did not change with exercise in either group.
Usually, RBC deformability decreases with exercise (5, 29).
The increased plasma [Lac] and decreased pH that occur
during exercise lead to water loss by RBCs (osmotic process),
which, in turn, results in greater rigidity (20, 32). In our study,
however, the rise in plasma [Lac] during exercise was slight in
all subjects, which can explain the lack of RBC rigidity change.
The subjects exhibiting EIH had higher RBC lactate transport
activity at 8.1 and 41 mM [Lac], but, in vivo, the plasma [Lac]
reached lower values during exercise (<6 mM at the end of
exercise). These results could also explain the lack of RBC
rigidity difference between the two groups at the end of
exercise.

**EIH**

We investigated blood rheology to clarify its relationship
with EIH. Because the measurements were performed at a
high-shear rate, we were able to hypothesize about the effects
of blood rheological properties in small vessels like the capil-
laries, even though the blood was drawn from the antecubital
vein. At a high-shear rate, blood viscosity depends primarily on
erthrocyte rigidity and then on plasma viscosity and Hct,
whereas, at a low-shear rate, blood viscosity depends mostly on
aggregation properties and Hct.

Previous studies have suggested a potential role of RBC
rigidity in pulmonary hemodynamics and pulmonary diffusion
limitation. Weiss et al. (35) perfused pony lungs with pentox-
ifylline-treated RBCs, which are known to be very deformable.
They observed a 27% decrease in filtration pressure, resulting
in a 10% decrease in pulmonary arterial pressure. In master
athletes, Aguilaniu et al. (1, 2) observed a decrease in EIH
severity after a PUFA diet. Because a PUFA diet is known to
improve RBC deformability, a potential contribution of blood
parameters to EIH genesis cannot be excluded. In our study,
we do not understand why hemorheology did not contribute to EIH during the
submaximal exercise performed at 85% VO₂ max, because no difference between groups was noted for any of the hemorho-
logical parameters. Nevertheless, blood viscosity may contrib-
ute to EIH during very intense exercise, i.e., >90% VO₂ max,
because we observed a greater increase in blood viscosity in the endurance-trained subjects with EIH compared with those
without EIH during a progressive and maximal exercise (6).

In conclusion, higher total lactate transport activity and
greater MCT-1-mediated lactate influx were found in the RBCs
from the EIH subjects, but erythrocyte deformability and blood
viscosity were similar in the two groups of endurance-trained
subjects. We suggest that the higher lactate influx into RBCs
may be more closely related to hypoxemia than to aerobic
capacity. Further studies are needed to identify the underlying
physiological process.

**ACKNOWLEDGMENTS**

The authors express their sincere gratitude to all of the athletes
who participated in the present study. We are also grateful to the staffs from the Laboratoire de Physiologie des Interactions (Hôpital Arnaud de Villeneuve, Montpellier, France) and Service de Médecine Nucleaire (Hôpital Lapeyronie, Montpellier, France) for their vital contribution. We thank C. Carmen for help in writing the English manuscript.

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LACTATE INFUX INTO ERYTHROCYTES IN HYPOXEMIC ATHLETES


