Carotid body denervation eliminates apnea in response to transient hypocapnia

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WE WERE CONCERNED in this study with the role of the carotid chemoreceptors as a cause of hypocapnia-induced central apnea and periodic breathing (PB) in sleep. There is evidence available to support peripheral or central CO2 chemoreceptors as key mediators of this response. A case for carotid chemoreceptors may be made because central apneas often occur within 10–15 s of a transient ventilatory overshoot in sleep (7, 27, 43). The functional response time of the central chemoreceptors appears to be on the order of 25–30 s (10), which would be too slow to account for apneas in response to transient hypocapnia. Furthermore, Bowes et al. (9) showed that a single, passive augmented breath resulted in significant expiratory time (Te) prolongation in vagally blocked awake dogs and that this Te prolongation was dependent on the presence of carotid chemoreceptors. Others have shown in anesthetized animals that carotid sinus nerve activity and ventilation are very sensitive to local changes in Pco2 confined to the carotid body (CB) (19, 26) and that congestive heart failure (CHF) patients with and without Cheyne-Stokes respiration (CSR) can be distinguished on the basis of their ventilatory response to transient CO2, a test that presumably reflects the responsiveness of the carotid chemoreceptors (43).

On the other hand, medullary chemoreceptors are also highly sensitive to H+ concentration changes in their environment, as shown by the ventilatory responses to ventricular-cisternal perfusion in awake goats (18, 36) and to focal acidosis of various chemosensitive areas of the medulla (32). Marked ventilatory depression and even apnea are also caused by ablation of medullary chemosensitive regions in anesthetized cats and goats (20, 40). Furthermore, some studies estimate the time delay from lung to medullary surface to be sufficiently short so as to approximate the delay from ventilatory overshoot to apnea (1, 16, 25). Millhorn et al. (29) have also shown in anesthetized cats that periodic oscillations in phrenic nerve activity corresponded in time and amplitude with those in pH on the medullary surface. PB in CHF patients has also been shown to correspond with an enhanced ventilatory response to hyperoxic CO2, a test that is thought to measure the sensitivity of the medullary chemoreceptors exclusively (24, 45). Finally, our own studies using extracorporeal perfusion of the isolated carotid chemoreceptor in the sleeping dog showed that a progressive CB hypocapnia led to immediate and progressive reductions in tidal volume (Vt) and minute ventilation but that step reductions in CB Pco2 of more than 10 Torr or increases in CB Pco2 of >500 Torr did not affect breath timing or cause apnea (42).

Our present approach was to study the intact and CB denervated dog under conditions that mimicked the occurrence of central sleep apnea and some types of PB commonly seen in sleeping humans. Accordingly, we...
studied the animals in non-rapid eye movement sleep and used mechanical ventilation in the pressure support mode (PSV) to cause transient increases in VT and reductions in arterial PCO\(_2\) (P\(_{ETCO2}\)). It had already been established that similar approaches would produce apnea in the intact sleeping dog (30) and in humans (28, 48) once end-tidal P\(_{CO2}\) (P\(_{ETCO2}\)) was reduced below the apneic threshold.

**METHODS**

Studies were performed over several days during non-rapid eye movement sleep on four unanesthetized, CB-denervated, female mixed-breed dogs (20–25 kg). These dogs were a subset of six dogs reported on in a previous publication when they were neurally intact (30). The dogs were trained to sleep in an air-conditioned (19–22°C), sound-attenuated chamber. They were unrestrained and allowed to pick their own sleeping positions. Throughout all experiments, the dogs’ behaviors were monitored by an investigator seated within the chamber and also by closed-circuit television. The Animal Care and Use Committee of the University of Wisconsin approved the surgical and experimental protocols for this study.

**Chronic instrumentation and CB denervation.** These dogs were chronically instrumented for a prior study (30). Details of this chronically instrumented dog model are described in detail elsewhere (12, 30, 39). Briefly, the dogs were prepared with a chronic tracheostomy and indwelling electromyogram (EMG) electrodes in the crural diaphragm, a five-electrode electroencephalogram montage, and an arterial catheter.

A relatively minor additional surgical procedure was required to achieve CB denervation (CBX). The carotid sinus region was exposed bilaterally, and all tissues surrounding the arteries of the region were removed over a distance of 1–2 cm. General anesthesia and strict sterile surgical techniques were used. Dogs recovered for at least 1 wk before any studies were performed. CBX was confirmed before each experiment by a lack of a significant ventilatory response to intravenous bolus injections of 20–40 μg/kg of sodium cyanide (NaCN). When the dogs were intact, identical doses of NaCN resulted in a transient two- to threefold increase in ventilation.

**Experimental setup and measurements.** Dogs breathed via auffed endotracheal tube (10.0 mm outer diameter; Shiley, Irvine, CA) that was inserted into the chronic tracheostomy. Airflow was measured via a heated pneumotachograph system (model 3700, Hans Rudolph, Kansas City, MO; model MP-45-14-871, Validyne, Northridge, CA) connected to the endotracheal tube. The pneumotachograph was calibrated before each study with four known flows. Tracheal pressure was measured at a port in the endotracheal tube that was connected to a pressure transducer (model MP-45-14-871, Validyne). The pressure transducer was calibrated before each study by applying six known pressures. Airway PO\(_2\) and PO\(_2\) were monitored by a mass spectrometer (model MGA-1100, Perkin-Elmer, Norwalk, CT) through a second port in the endotracheal tube. Three to six 1-ml arterial samples were obtained at the start of each experiment from the aortic catheter and analyzed for pH, P\(_{O2}\), and P\(_{CO2}\) on a blood-gas analyzer (model ABL-505, Radiometer, Copenhagen, Denmark). The blood-gas analyzer was validated daily with dog blood tonometered with three different combinations of PO\(_2\) and PCO\(_2\) covering the range encountered in the experiments. Samples were corrected for both body temperature and systematic errors revealed by tonometry. The inspiratory and expiratory tubes of the ventilator were connected to the pneumotachograph by using a Y connector. A silent balloon valve was placed between the pneumotachograph and the Y connector such that the dog could breathe spontaneously from room air or be abruptly switched to PSV by inflation of the balloon. All signals were digitized (128-Hz sampling frequency) and stored on the hard disk of a personal computer for subsequent analysis. Key signals were also recorded continuously on a polygraph (Gould ES 2000 or AstroMed K2G). All ventilatory data were analyzed on a breath-by-breath basis by means of custom analysis software developed in our laboratory.

**Use of PSV to characterize the time course of TE responses to hypocapnia.** Dogs breathed room air spontaneously through the open port in the balloon valve (see above). The mechanical ventilator (Veolar, Hamilton Medical) was set in pressure support mode, and the trigger sensitivity was set as low as possible (approximately −2 cmH\(_2\)O). When the balloon was inflated and the low resistance shunt to the room sealed, the ventilator delivered preset levels of inspiratory pressure support whenever the trigger threshold was reached. The expiratory positive airway pressure was set at 0 cmH\(_2\)O. Thus the dog set its own breathing frequency, and increased levels of pressure support resulted in increased VT and decreased P\(_{ETCO2}\). As assessed from the diaphragmatic electromyogram (EMG\(_{dia}\)), 95–100% of the increase in VT during PSV occurred during neural inspiratory time. Each level of pressure support was maintained for 2 min, and then the balloon was deflated and the dog was allowed to breathe spontaneously again (Fig. 1). At least 2 min elapsed before another PSV trial was performed. PSV was begun at a pressure of 3–5 cmH\(_2\)O and increased in steps of 1–2 cmH\(_2\)O in each succeeding trial until apnea and/or PB was achieved (range of 3–35 cmH\(_2\)O). Trials in which there was a state change or sigh were excluded from analysis (see Ref. 30 for details).

**Characterizing P\(_{ETCO2}\) thresholds for TE prolongation/apnea in the intact and CBX dog.** TE was measured from the end of the inspiratory flow to the onset of the next EMG\(_{dia}\) burst. In the intact dog, PB was identified visually by the presence of at least three cycles of hyperpnea and apnea, as judged by EMG\(_{dia}\), with a consistent cycle length, and the apnea lengths had to be at least 3 standard deviations greater than the mean baseline spontaneous TE. It is important to note that the apneic threshold was taken to be the P\(_{ETCO2}\) observed in the breath immediately preceding the initial apneic period only.

After CBX, PB was never observed, so it was not straightforward to establish a P\(_{ETCO2}\) threshold for apnea. Our method in the CBX dogs was to choose the longest TE in the 10th to 60th seconds of each PSV trial. We reasoned that apneas that occurred before 10 s of PSV would not be chemoreceptor mediated because CB were lacking. We then associated the chosen TE with the lowest P\(_{ETCO2}\) observed from 0 to the 50th seconds of PSV; i.e., the lowest P\(_{ETCO2}\) that would have been required to produce an increase in TE that was the same as the increase observed at the apneic threshold when that dog was intact. It is important to note that calculated ΔP\(_{ETCO2}\),...
was always within the range of the regression data; we did not extrapolate from the regression line.

\[ \Delta pH \] calculations. The Siggaard-Anderson nomogram was used to estimate changes in pH that accompanied changes in the measured \( \Delta PETCO_2 \) values between spontaneous breathing and the apneic threshold. For changes in arterial pH, we assumed that the dogs' blood buffer slopes were parallel to the normal human blood buffer slope and passed through the measured eupneic PaCO_2-pH point determined for each dog on each day. Changes in cerebrospinal fluid (CSF) pH for a given change in \( \Delta PETCO_2 \) between eupnea and apnea were estimated by assuming no change in CSF HCO_3\(^{-}\) concentration ([HCO_3\(^{-}\)]). We also assumed that CSF [HCO_3\(^{-}\)] was equal to the measured eupneic arterial [HCO_3\(^{-}\)] measured on that day, that CSF PCO_2 was equal to PETCO_2 + 6 Torr (41), and that \( \Delta PETCO_2 \) equaled the change in CSF PCO_2.

Statistical analyses. Significance of group mean data was determined with a paired t-test. Differences were considered significant at \( P < 0.05 \).

**RESULTS**

**CB denervation.** CB denervation resulted in hypoventilation and respiratory acidosis in all dogs, although the changes in breathing frequency and VT employed varied between dogs (Table 1). The absence of significant CO_2 retention post-CBX in dog Ni is noteworthy. Despite the small CO_2 retention, this dog was clearly denervated because she showed no ventilatory response to doses of NaCN that promoted vigorous hyperpnea when the dog was intact.

**Time course of response to PSV: intact vs. CBX.** Figure 1A shows a typical intact PSV trial at a level of pressure support (15 cmH_2O) that produced sufficient hypocapnia to elicit PB. In this dog, 3 trials at PSV levels of 15–20 cmH_2O elicited PB, whereas 11 trials at PSV levels of 5–19 cmH_2O did not. PSV was initiated...
not consistently prolonged (i.e.,
6.0
than spontaneous baseline, but PB did not occur. In contrast, 32.8
apnea occurred (typically on the second PSV breath).
levels of PSV, increased VT, and hypocapnia in the
port breath with a VT that remained twofold greater
About 10
–
required in the CBX dogs before clear, hypocapnia-
control value on the
rst PSV breath and remained
emergent: one peak averaging slightly longer than spontaneous eupneic Te and the other centered on
apnea duration. This distribution of Te is predictable
from a breathing pattern consisting almost entirely of
doublets (Fig. 1). In contrast, although PSV-induced
hypocapnia in the same dogs after CBX resulted in Te
prolongations over time, PB was never observed (Figs.
1 and 2). When viewed as a histogram, these data
present a single broad peak at a mean Te about mid-
way between the two Te peaks of the intact dog (Fig. 3).

Central vs. peripheral chemoreceptor sensitivity
to hypocapnia. When the dogs were intact, apnea occurred
(Te = 8.4 ± 1 s) after the second ventilator
(10–11 s of PSV); this required a decrease in
PETCO2 of 5.1 ± 0.4 Torr. After CBX, as shown above
(Figs. 1 and 2), PB never occurred in response to
PSV-induced hypocapnia. However, given time, Te was
always prolonged in the CBX dogs, and this allowed us
to calculate an apneic threshold (see METHODS). After
CBX, an apneic equivalent in length to that seen intact
(8.4 ± 1 s) required a twofold decrease in
PETCO2 of 10.1 ± 2.1 Torr (range = 8.2–12.9 Torr; significantly
different from intact, P < 0.02), relative to intact, and this
apnea occurred after ~33 s of PSV (Fig. 4). These
differences in ΔPETCO2 between intact and CBX coincided with changes in the slopes of the ventilatory
responses below eupnea, i.e., from apnea to eupnea
(ΔVT/ΔPETCO2, where VT is inspiratory minute ventilation,
in l·min
–1·Torr
–1). For dogs G, J, N, and Ni, respectively, the intact slopes were 0.74, 0.75, 0.59, and 0.38; slopes after CBX were 0.28, 0.26, 0.21, and 0.24, respectively.

When the dogs were intact, apnea required an increase in estimated pHa of 0.036 ± 0.005 and estimated
CSF pH of 0.052 ± 0.005. After CBX, an apnea equivalent to that seen in intact animals required increases in
pHa of 0.068 ± 0.016 (significantly different from intact dogs, P < 0.01), and estimated CSF pH of 0.094 (significantly different from intact, P < 0.01).

Thus, with CBX, baseline PCO2 was increased and pH was reduced in both arterial blood and (presumably) CSF. The changes in PCO2 and pH required in the CBX
dog to cause the same apneic length as was observed when the dog was intact both increased about twofold.

### Table 1. Mean apneic values during spontaneous breathing in NREM sleep before and after CBX

<table>
<thead>
<tr>
<th>Dog</th>
<th align="right">Ti, s</th>
<th align="right">Te, s</th>
<th align="right">VT, liter</th>
<th align="right">fb, breaths/min</th>
<th>pHa</th>
<th align="right">PCO2, Torr</th>
<th align="right">Pao2, Torr</th>
<th align="right">[HCO3−], mEq/l</th>
</tr>
</thead>
<tbody>
<tr>
<td>G intact</td>
<td align="right">1.5 ± 0.2</td>
<td align="right">4.5 ± 0.8</td>
<td align="right">0.32 ± 0.03</td>
<td align="right">10.4 ± 1.7</td>
<td>7.394 ± 0.008</td>
<td align="right">35.7 ± 0.9</td>
<td align="right">98 ± 3</td>
<td align="right">21.3 ± 0.7</td>
</tr>
<tr>
<td>G CBX</td>
<td align="right">1.6 ± 0.2</td>
<td align="right">5.9 ± 0.9</td>
<td align="right">0.28 ± 0.04</td>
<td align="right">8.2 ± 1.1</td>
<td>7.340 ± 0.007</td>
<td align="right">46.6 ± 2.6</td>
<td align="right">88 ± 7</td>
<td align="right">24.6 ± 1.4</td>
</tr>
<tr>
<td>J intact</td>
<td align="right">1.4 ± 0.2</td>
<td align="right">4.0 ± 0.9</td>
<td align="right">0.29 ± 0.02</td>
<td align="right">11.6 ± 2.6</td>
<td>7.369 ± 0.004</td>
<td align="right">39.9 ± 0.1</td>
<td align="right">110 ± 3</td>
<td align="right">22.2 ± 0.1</td>
</tr>
<tr>
<td>J CBX</td>
<td align="right">1.3 ± 0.1</td>
<td align="right">3.1 ± 0.7</td>
<td align="right">0.19 ± 0.04</td>
<td align="right">14.2 ± 2.5</td>
<td>7.340 ± 0.011</td>
<td align="right">51.1 ± 1.1</td>
<td align="right">85 ± 6</td>
<td align="right">26.8 ± 1.3</td>
</tr>
<tr>
<td>N intact</td>
<td align="right">1.4 ± 0.1</td>
<td align="right">2.9 ± 0.2</td>
<td align="right">0.22 ± 0.03</td>
<td align="right">14.4 ± 0.9</td>
<td>7.356 ± 0.025</td>
<td align="right">36.5 ± 0.5</td>
<td align="right">97 ± 5</td>
<td align="right">23.1 ± 1.0</td>
</tr>
<tr>
<td>N CBX</td>
<td align="right">1.4 ± 0.2</td>
<td align="right">3.0 ± 1.1</td>
<td align="right">0.18 ± 0.03</td>
<td align="right">15.4 ± 4.7</td>
<td>7.354 ± 0.002</td>
<td align="right">45.9 ± 0.8</td>
<td align="right">88 ± 3</td>
<td align="right">24.9 ± 0.2</td>
</tr>
<tr>
<td>Ni intact</td>
<td align="right">1.5 ± 0.3</td>
<td align="right">3.4 ± 0.6</td>
<td align="right">0.16 ± 0.04</td>
<td align="right">12.8 ± 1.2</td>
<td>7.359 ± 0.019</td>
<td align="right">39.7 ± 0.3</td>
<td align="right">111 ± 3</td>
<td align="right">21.8 ± 0.9</td>
</tr>
<tr>
<td>Ni CBX</td>
<td align="right">1.3 ± 0.1</td>
<td align="right">3.7 ± 0.5</td>
<td align="right">0.17 ± 0.02</td>
<td align="right">12.1 ± 1.0</td>
<td>7.341 ± 0.013</td>
<td align="right">45.3 ± 1.8</td>
<td align="right">95 ± 9</td>
<td align="right">23.9 ± 0.7</td>
</tr>
<tr>
<td>Intact mean</td>
<td align="right">1.5 ± 0.1</td>
<td align="right">3.7 ± 0.6</td>
<td align="right">0.25 ± 0.06</td>
<td align="right">12.3 ± 1.5</td>
<td>7.377 ± 0.014</td>
<td align="right">38.0 ± 1.9</td>
<td align="right">104 ± 7</td>
<td align="right">21.7 ± 0.4</td>
</tr>
<tr>
<td>CBX mean</td>
<td align="right">1.4 ± 0.1</td>
<td align="right">3.9 ± 1.2</td>
<td align="right">0.21 ± 0.04</td>
<td align="right">12.5 ± 2.7</td>
<td>7.343 ± 0.006*</td>
<td align="right">47.2 ± 2.3*</td>
<td align="right">89 ± 4*</td>
<td align="right">25.1 ± 1.1*</td>
</tr>
</tbody>
</table>

Values are means ± SD. NREM, non-rapid eye movement; CBX, carotid body denervation; Ti, inspiratory time; Te, expiratory time; VT, tidal volume; fb, breathing frequency; PCO2, arterial PCO2; Pao2, arterial PO2; [HCO3−], HCO3 concentration; G, J, N, Ni, dog designation. *Significantly different from intact mean (P < 0.05).
Hyperoxia. In three of the four dogs, we completed several trials of PSV under hyperoxic conditions (PETCO₂ of ~300–350 Torr) when the dogs were still neurally intact. Steady-state hyperoxia had no consistent effect on spontaneous breathing pattern or PETCO₂. As shown in Fig. 5, hyperoxia did not prevent rapid development of apneas in response to PSV-induced hypocapnia, as prolonged TE (>3 SD than control mean) occurred by the second or third PSV breath. However, as shown in the histogram in Fig. 3, relative to normoxia, 1) the average apneic lengths in hyperoxia were shorter (6.0 ± 1.6 vs. 8.4 ± 1 s), 2) PB was transient and not tightly organized in doublets as seen in intact dogs, and 3) periodicity ceased after ~40 s of PSV, although TE was prolonged (see Fig. 5). Thus the effects of hyperoxia on hypocapnia-induced PB were very similar to those following CBX except for the rapidity with which TE was prolonged after the ventilatory overshoots.

Neuromechanical effects of PSV. PSV in both intact and CBX dogs had an immediate effect on TE and EMGdia. In the intact dog, the first breath of PSV prolonged TE by 10.220.33.4 on April 9, 2017 http://jap.physiology.org/ Downloaded from
a small fraction of the apnea length observed after the second or third breath (4.7 ± 1.1 s). After CBX, the first breath of PSV also prolonged T\text{E} to 122 ± 16% (not significantly different from intact) and reduced EMG\text{dia} to 85 ± 10% (not significantly different from intact) of the spontaneously breathing, eupneic values (Table 2).

**DISCUSSION**

Our study used intact and CBX sleeping dogs subjected to varying levels of pressure support ventilation to address the role of the carotid chemoreceptors in hypocapnic-induced apnea and PB. By contrasting the time course of responses of intact vs. CBX animals to PSV, we determined that the presence of carotid chemoreceptors was required to produce the apnea normally caused by a transient hyperventilation and hypocapnia and that these receptors were also required for the PB pattern produced by PSV. Hyperoxia in the intact animal did not (unlike CBX) prevent the immediate apnea in response to transient hyperventilation but (like CBX) prevented prolonged apneas and a distinct periodic pattern. Finally, the level of hypocapnia required to cause apnea in the CBX animal was found to be about two times that required for apneas of similar length in the intact animal. Implications of these findings to human sleep apnea are discussed.

**Critical role of carotid chemoreceptors in hypocapnic-induced apnea.** Our data in sleeping dogs point strongly to an obligatory role for carotid chemoreceptors in causing the central apneas that commonly occur after transient ventilatory overshoots, which drive Pa\text{CO}_2 below the apneic threshold. First, apneas occurred within 10–12 s or two breaths of the onset of PSV and the transient rise in V\text{T} and fall in PET\text{CO}_2. More to the point, denervation of the carotid chemoreceptors prevented the occurrence of apnea within its normal short latency after the ventilatory overshoot. Apneas still occurred in the CBX animal but not until 33 s after the ventilatory overshoot, and PB never occurred. So, carotid chemoreceptors appear to be obligatory in causing the apneas during the time period in which they commonly occur as a result of brief, transient, ventilatory overshoots. This means that hypocapnia-induced apnea required an ~5 Torr reduction in Pa\text{CO}_2, relative to eupnea, at the carotid chemoreceptor.

Is CB hypocapnia sufficient by itself to cause apnea? On the one hand, there is ample evidence demonstrating substantial sensitivity of the carotid chemoreceptors to changes in pH and CO\text{2} (see Introduction). On the other hand, there are also several lines of evidence

![Fig. 3. Histograms of all PSV breaths from 1 dog when intact breathing room air (dashed line), intact breathing 100% O\text{2} (thin dotted line), and after CBX breathing room air (solid line). Only data from trials with apnea and/or PB are shown for the intact condition (normoxia or hyperoxia); for CBX animals, trials were included if their initial \Delta PET\text{CO}_2 was equal to or greater than that observed during PB when a given dog was intact. Note that when dogs were intact and breathing room air there were two clear TE peaks, the first only slightly longer than the eupneic control and the second representing the apneic TE (see RESULTS for details). In contrast, both the normoxic CBX trials and the intact hyperoxic trials produced a single, broad TE peak between the two intact peaks. Apnea observed early in the hyperoxic trials was not sustained, which thus explained the similarity of CBX and hyperoxic TE distribution.](image)

![Fig. 4. This figure illustrates the reduction in PET\text{CO}_2 required to cause an apnea of 8.4 ± 1-s duration while intact and after CBX in each of the 4 dogs during NREM sleep. Filled bars, intact dogs; open bars, CBX dogs. Note that, on average, \Delta PET\text{CO}_2 was about twofold greater after CBX than when intact. Ventilatory response slopes below eupnea after CBX averaged only 40% of the intact slope values (see RESULTS). G, J, N, Ni, dog designations.](image)
to suggest that additional mechanisms beyond just CB hypocapnia, per se, contribute to the apnea after transient hyperventilation.

First, our own previous data in intact sleeping dogs showed that progressive step changes in hypocapnia of CB PCO2 of $-3$ to $-15$ Torr applied via extracorporeal perfusion to the isolated carotid chemoreceptor caused an immediate (within 5 s) dose-dependent reduction in VT and minute ventilation; however, apnea or substantial changes in breath timing did not occur. Second, even extreme hyperoxia (CB PCO2 $>500$ Torr) applied to the isolated carotid chemoreceptor did not cause apnea (42). So, how can the carotid chemoreceptors be required for apneas that occur after a transient ventilatory overshoot but not cause apnea when subjected to CB-specific transient hypocapnia (or even hyperoxia)?

One key difference between the studies is that the transient hypocapnia in the present study was produced by an increased VT (via PSV), whereas the hypocapnia presented to the isolated CB via extracorporeal perfusion was not accompanied by a ventilatory overshoot. The increase in VT may exert two additional types of potential inhibitory effects on respiratory motor output. First, within-breath oscillations in PaCO2 and pH would be enhanced with an augmented VT, which in turn would be sensed by carotid chemoreceptors and therefore affect respiratory motor output, depending on when in the respiratory cycle the peak or nadir of the Pco2 oscillation arrived at the CB, i.e., a “gating” effect on respiratory motor output (14, 17). For example, to cause TE prolongation, the trough of the PaCO2 (and alkaline peak of arterial blood pH) would have to arrive at the carotid chemoreceptors during early expiration. On the other hand, even substantial oscillations in arterial pH caused only 30% prolongations of TE in anesthetized dogs (14) and removal of all normal oscillations in PaCO2 in anesthetized (44) or awake (42) animals did not influence ventilatory output significantly, at least under conditions of resting CO2 production.

Second, the feedback effects of increased VT and lung stretch also would have accompanied ventilatory overshoot but not specific CB hypocapnia via CB perfusion. Certainly, lung stretch by itself could not have caused apnea because apnea did not occur immediately after the ventilatory overshoot in the CBX animal (see Fig. 2) nor did apnea occur when hypocapnia was prevented [via increased inspired CO2 fraction (FICO2)] during the

**Table 2. Mean Ti, Te, and EMGdia of the first breath of all PSV trials when dogs were intact and after CBX**

<table>
<thead>
<tr>
<th>Dog</th>
<th>Intact Ti, % of spontaneous</th>
<th>CBX Ti, % of spontaneous</th>
<th>Intact Te, % of spontaneous</th>
<th>CBX Te, % of spontaneous</th>
<th>Intact EMGdia, % of spontaneous MEA</th>
<th>CBX EMGdia, % of spontaneous MEA</th>
</tr>
</thead>
<tbody>
<tr>
<td>G</td>
<td>94 ± 15</td>
<td>103 ± 14</td>
<td>102 ± 22</td>
<td>95 ± 15</td>
<td>76 ± 16*</td>
<td>70 ± 18*</td>
</tr>
<tr>
<td>J</td>
<td>104 ± 12</td>
<td>112 ± 15*</td>
<td>117 ± 16*</td>
<td>127 ± 32*</td>
<td>74 ± 14*</td>
<td>83 ± 14*</td>
</tr>
<tr>
<td>N</td>
<td>111 ± 10*</td>
<td>133 ± 20*</td>
<td>117 ± 30*</td>
<td>138 ± 20*</td>
<td>69 ± 17*</td>
<td>98 ± 20</td>
</tr>
<tr>
<td>Ni</td>
<td>119 ± 18*</td>
<td>115 ± 14*</td>
<td>146 ± 31*</td>
<td>128 ± 32*</td>
<td>91 ± 17*</td>
<td>89 ± 15*</td>
</tr>
<tr>
<td>Mean</td>
<td>107 ± 9</td>
<td>116 ± 11</td>
<td>121 ± 16</td>
<td>122 ± 16</td>
<td>78 ± 8</td>
<td>85 ± 10</td>
</tr>
</tbody>
</table>

Values are means ± SE. PSV, pressure support ventilation; MEA, mean electrical activity.
PSV-induced ventilatory overshoot (30). However, during the ventilatory overshoot, rapid decreases in PaCO₂ and carotid sinus nerve activity occur coincident with lung stretch. Bajic et al. (5), using recordings from medullary respiratory neurons, showed that these combined carotid chemoreceptor-mechanoreceptor inputs have a clear additive interaction at the level of the central respiratory controller. Perhaps, then, this interactive effect was required to cause apnea. This is a hypothesis that may be tested by determining the effects of vagal blockade on the response to transient hyperventilation.

Our findings do not rule out a significant contribution from central chemoreceptors to the hypocapnia-induced apnea in the intact animal. The recent work of Okada et al. (35), Ballantyne and Scheid (6), and Richardson et al. (37), for example, supports the idea of a perivascular location for many chemosensitive medullary neurons. Older studies have shown, on the basis of medullary surface pH measurements, that pH can change very rapidly, approximating the delay from ventilatory overshoot to apnea (1, 16, 25). Taken together, these data are consistent with rapidly responding central chemoreceptors. That said, it is important to recognize that most evidence suggests that the functional (i.e., respiratory motor output) response of the central chemoreceptors alone is relatively slow (25–30 vs. 10–15 s after ventilatory overshoot in an intact animal (Refs. 9, 10, present study)]. These data all come from studies that utilized CBX. However, whatever mechanisms may be responsible for the slow central response time in CBX animals, CB are required to produce apneas in response to transient hypocapnia. So, although we believe the most straightforward interpretation of available data indicates that central chemoreceptors respond to hypocapnia relatively slowly, we acknowledge the possibility that in the intact animal the sensitivity and/or response time of central chemoreceptors might be significantly enhanced by tonic or phasic input from the carotid chemoreceptors and thereby contribute significantly to the observed apnea (see Relative CO₂ sensitivities of the peripheral and central chemoreceptors below).

In summary, our past and present data in intact and CBX sleeping dogs, respectively, demonstrate that, although carotid chemoreceptors are essential to explain apneas caused by the hypocapnia normally resulting from transient ventilatory overshoots, the effects of CB hypocapnia, per se, are not sufficient to cause these apneas. Additional interactive effects with carotid chemoreception appear to be necessary to explain the apneas in the intact animal.

Relative CO₂ sensitivities of the peripheral and central chemoreceptors. The degree of hypocapnia (or change in pH) required in the CBX animal (and therefore presumably of the medullary chemoreceptors) was about twice that in the intact animal to achieve the same apnea length. We note that absolute values for PETCO₂ at the apneic threshold were relatively hypercapnic after CBX, because the eupneic PETCO₂ was elevated. However, it is not this absolute value but rather the magnitude of the change in PETCO₂ that is required to produce apnea that defines the sensitivity of the CO₂ response below eupnea (see Potential limitations of the experimental model below). Although our ability to quantify the apneic threshold (i.e., the ∆PETCO₂ from eupnea to apnea) in the CBX dog in the absence of PB was not as precise as in the intact dog, our approach did provide a conservative estimate of the differences in apneic threshold between intact and CBX conditions (see Methods).

These differences in ∆PETCO₂ (eupnea – apnea) and in ∆V̇ETCO₂/PETCO₂ imply that the carotid chemoreceptor sensitivity to hypocapnia (at least the sensitivity that was elicited by transient changes in PCO₂ resulting from a ventilatory overshoot) is substantially greater than that of the central chemoreceptors. Cross et al. (14) and Borison et al. (8) also showed little change in breath timing in response to increases in CO₂ and reductions in pH in CBX vs. intact anesthetized animals. Furthermore, in our studies in the isolated, intact, perfused CB, sustained CB hypocapnia alone caused hypoventilation that persisted for >2 min, even in the face of substantial systemic (PaCO₂ of 5–7 Torr) and presumably central nervous system hypercapnia (42). Only when CB hypocapnia was suddenly removed was the ventilatory response to systemic hypercapnia manifested in a marked ventilatory overshoot. Apparently then, the inhibitory effects of a persistent CB hypocapnia masked the excitatory influence of a substantial brain extracellular fluid acidosis.

On the other hand, CBX preparations, as presently used by us and others (8), removes the normal tonic input to the medullary pattern generator. Nattie (31) and Cohen (13) recently emphasized the importance of maintaining tonic inputs intact when quantification of chemoreceptor drives was attempted. We also showed qualitatively different ventilatory responses to systemic hypoxia in CBX animals vs. animals with the isolated CB intact and maintained normoxic and normocapnic via extracorporeal perfusion (15). Accordingly, our present estimates of central chemoreceptor sensitivity to hypocapnia in the CBX animals may underestimate the actual gain and thus its relative importance to apnea and PB.

In summary, we propose that both past and present findings point to a relatively high sensitivity to hypocapnia at the level of the carotid chemoreceptors that may exceed that of the medullary chemoreceptors. To test this hypothesis, additional experiments using the intact, isolated CB perfusion model are needed to quantify the sensitivity of the central chemoreceptors to systemic hypcapnia in the presence of normal tonic input from the carotid chemoreceptors.

Potential limitations of the experimental model. We used PSV to produce graded increases in V̇E and thereby graded decreases in PETCO₂. This method readily produces PB in normal, intact human subjects (29, 31, 50), and the ∆PETCO₂ (eupnea – apnea) at which PB occurs is quite reproducible. However, this method also causes some neuromechanical inhibition of ventilatory drive with the primary effect
being a reduction in the amplitude of the EMGdia and minor effects on breath timing (see Table 2). That these effects observed on the first cycle of PSV (Fig. 1, A and B) are not attributable to chemoreceptor inhibition is shown by the almost identical changes obtained in intact vs. CBX dogs (see Table 2). Similar, relatively small changes in breath timing and EMG amplitude occurred throughout 1–2 min of PSV that was applied to the intact animal when $F_{\text{ICO}}$ was raised to prevent $P_{\text{ETCO}}$ below the apneic threshold, two PSV breaths (or 10–12 s) were sufficient to cause apnea (46). Finally, increased $F_{\text{ICO}}$ in sleeping humans with PB was shown to eliminate apnea within 10–15 s of the onset of its application (3, 7, 47).

Second, the magnitude of the ventilatory responsiveness to transient (single breath) increases in $F_{\text{ICO}}$ was shown to correlate positively with the presence of PB in CHF and in idiopathic central sleep apnea (43). The fact that the degree of peripheral chemosensitivity (as was estimated from the transient hypercapnia test) accounted for <50% of the variance in the apnea hypopnea index among all patients (43) may be reflective of the importance of both the chemoreceptor-driven propensity to ventilatory overshoot as well as the relative sensitivity of the hypocapnia-induced apneic threshold in causing central sleep apnea.

Third, we found that, unlike CBX, hyperoxia did not prevent significant $T_{\text{E}}$ prolongation immediately after transient ventilatory overshoots. These findings are consistent with recordings of carotid sinus nerve activity in the anesthetized cat (26) and the effects of CBX in the awake cat and dog (22, 38), which showed that CB chemoreceptors remained sensitive to $CO_2$ even in the presence of hyperoxia. On the other hand, with hyperoxia, hypocapnia-induced apneas were significantly shorter than in normoxic hypocapnia, and, like CBX, clear PB was not sustained. The findings are analogous to the effects of supplemental oxygen treatment in patients with central apnea and CSR, in whom the number of apneas and hypopneas was significantly reduced but not eliminated (2, 21, 27).

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