Glycemic index of a meal fed before exercise alters substrate use and glucose flux in exercising horses

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Jose-Cunilleras, Eduardo, Kenneth W. Hinchcliff, Richard A. Sams, Steven T. Devor, and Jon K. Linderman. Glycemic index of a meal fed before exercise alters substrate use and glucose flux in exercising horses. J Appl Physiol 92: 117–128, 2002.—In a randomized, balanced, crossover study each of six fit, adult horses ran on a treadmill at 50% of maximal rate of oxygen consumption for 60 min after being denied access to food for 18 h and then 1) fed corn (51.4 kJ/kg digestible energy), or 2) fed an isocaloric amount of alfalfa 2–3 h before exercise, or 3) not fed before exercise. Feeding corn, compared with fasting, resulted in higher plasma glucose and serum insulin and lower serum nonesterified fatty acid concentrations before exercise (P < 0.05) and in lower plasma glucose, serum glycerol, and serum nonesterified fatty acid concentrations before exercise (P < 0.05). Feeding corn, compared with feeding alfalfa, resulted in higher carbohydrate oxidation and lower lipid oxidation during exercise (P < 0.05). Feeding a soluble carbohydrate-rich meal (corn) to horses before exercise results in increased muscle utilization of blood-borne glucose and carbohydrate oxidation and in decreased lipid oxidation compared with a meal of insoluble carbohydrate (alfalfa) or not feeding. Carbohydrate feedings did not produce a sparing of muscle glycogen compared with fasting.

glycogen; corn; alfalfa

MUSCLE GLYCOGEN AND blood-borne glucose are important substrates for contracting skeletal muscle during moderate-intensity exercise bouts. Carbohydrate (CHO) metabolism in exercising muscle and the factors that influence CHO metabolism during exercise in horses are assumed to be similar to humans. However, this may not necessarily be the case. Horses have a greater aerobic capacity than humans, and, therefore, at the same exercise intensity relative to the maximal rate of oxygen consumption (VO2 max), the rate of oxygen consumption (VO2) and the energy expenditure are much higher in horses than in humans (46). In addition to the greater aerobic capacity of horses, the anatomy and physiology of their gastrointestinal tract is adapted to a herbivorous diet based on ingestion of structural CHOs of plants (mainly cellulose and hemicellulose) and bacterial fermentation of structural CHOs in the hindgut with subsequent absorption of volatile fatty acids (VFAs; acetate, propionate, and butyrate). Horses obtain a substantial proportion of their energy requirements from absorption of VFAs during resting conditions as well as during low-intensity exercise; however, the contribution of VFAs to energy expenditure during exercise is unknown. Thereby, the observations made in human subjects regarding CHO metabolism in exercising skeletal muscle and the influence of nutritional status on CHO metabolism may not apply to horses because of the aforementioned physiological differences between horses and humans. Among the numerous factors that influence CHO metabolism in exercising muscle, there is evidence, in studies in human subjects, that diet composition and interval before exercise can affect both uptake of glucose by muscle and the rate of muscle glycogen utilization. In studies performed in human subjects, ingestion of glucose or a high-glycemic meal (HGM) before exercise results in enhanced CHO oxidation (CHOox) and utilization of blood-borne glucose, and the rate of glycogenolysis is enhanced (18) or unchanged (8, 17, 22, 23). In humans, ingestion of a HGM or CHO supplementation before and/or during endurance and moderate-intensity exercise has resulted in enhanced (1, 2, 4, 5, 13, 16, 23, 37, 38, 45, 49), decreased (44), or unchanged submaximal exercise performance (8, 9, 18, 40, 47). Current recommendations of feeding before and during endurance exercise in human athletes include ingestion of CHO solutions to maintain the rate of CHOox (21). Studies performed in horses have determined the effect of meal composition and interval before exercise on the blood concentrations of substrates and hormones and muscle glycogen utilization during exercise (7, 24, 25, 29, 30, 34, 43); however, the relative contributions of CHOox, in the form of blood-borne glucose
and muscle glycogen utilization, and lipid oxidation to energy production during exercise have not been determined. In brief, ingestion of a HGM 2–4 h before a moderate-intensity exercise bout results in a transient decrease in plasma glucose concentration during exercise, attenuation of exercise-induced increase of nonesterified fatty acid (NEFA) concentration, and increased serum insulin concentration during exercise compared with horses withheld from food or fed a hay meal. These alterations in plasma and serum concentrations of substrates and hormones have been hypothesized to be deleterious for performance in horses, presumably because of impaired substrate use during exercise. However, none of these studies has determined the effect of meal composition before exercise on substrate use during exercise or on measurements of exercise performance in horses.

If similar mechanisms regulate CHO metabolism in working skeletal muscle in horses and in humans, it is plausible that preexercise ingestion of a HGM in horses may enhance glucose availability to skeletal muscle and alter the hormonal milieu so that \( \text{CHO}_\text{ox} \) is maintained and lipid oxidation is suppressed, compared with ingestion of a low-glycemic meal (LGM) or not feeding before exercise. To date, however, this hypothesis has not been tested. The present study was, therefore, undertaken to determine the effects of glucose supply (preexercise ingestion of a HGM (corn) vs. preexercise ingestion of a LGM (hay) vs. withholding feed) on stable isotope tracer-determined whole body glucose appearance and uptake in horses during moderate-intensity exercise. It was hypothesized that ingestion of a HGM before exercise would result in enhanced whole body glucose appearance and uptake. A further objective of this study was to determine the effects of glucose supply on whole body rates of \( \text{CHO}_\text{ox} \) and lipid oxidation and relative contribution of these fuels to energy expenditure as well as to the rate of muscle glycogenolysis. It was hypothesized that ingestion of a HGM before exercise would result in enhanced \( \text{CHO}_\text{ox} \), attenuated lipid oxidation, greater relative contributions to energy expenditure from blood-borne glucose, and attenuated muscle glycogenolysis (“muscle glycogen-sparing effect”).

**MATERIALS AND METHODS**

All animal experiments were conducted after approval by the Institutional Laboratory Animal Care and Use Committee of The Ohio State University and were performed in compliance with their guidelines and recommendations. **Experimental design.** The effects of feeding a meal of corn or alfalfa before exercise on glucose kinetics, whole body substrate utilization, and muscle glycogen utilization during moderate-intensity exercise were examined in a balanced, randomized, three-way crossover study. Six horses underwent 60 min of treadmill exercise at a workload requiring 50% \( \text{VO}_2\text{max} \) in each of three experimental conditions: 1) 90 min after consuming a meal of cracked corn (1.7 kg in a 450-kg horse; 12.4 kcal/kg (51.8 kJ/kg) digestible energy; grain trial); 2) 90 min after consuming a meal of an isocaloric amount of alfalfa cubes (3.0 kg in a 450-kg horse; hay trial); and 3) after having feed withheld for 18 h (feed-withholding trial). In preliminary studies, it was determined that horses offered hay were not able to consume the entire meal in 60 min. For this reason, horses were allowed to eat the grain meal from 0800 to 0900 (60 min) and the hay meal from 0700 to 0900 (120 min). Trials were separated by 7 days, and the order of the trials was randomized for individual horses but balanced among treatments.

**Horses.** Six Standardbred horses (5 mares and 1 gelding), 4–11 yr (7.8 ± 2.5 SD) yr of age and 404–485 kg (425 ± 30 SD) kg) body mass, were studied. All horses were housed indoors during the experimental period, fed a diet of timothy grass-alfalfa hay, alfalfa cubes, mixed pelleted grain, and cracked corn, and had unlimited access to a salt-mineral block and water. All horses were conditioned and undertaking regular treadmill exercise for 6 wk before the study. Horses were not exercised the day before an experimental trial, and, after an experimental trial, they received 4 days of light treadmill exercise (30 min of trotting at 4–4.5 m/s with the treadmill set at 4° incline).

**Preliminary testing.** For each horse, \( \text{VO}_2\text{max} \) and the relationship between rate of \( \text{VO}_2 \) and speed were determined during an incremental exercise test 1 wk before the first experiment. The incremental exercise test consisted of the horse running on a high-speed treadmill (Sato, Uppsala, Sweden) inclined at 4° for 90 s at 4 m/s; the treadmill speed was then increased by 1 m/s every 90 s until the horse was no longer able to maintain its position on the treadmill. \( \text{VO}_2 \) was measured every 10 s during the exercise test. \( \text{VO}_2\text{max} \) was defined as the value at which \( \text{VO}_2 \) reached a plateau, despite further increases in speed. A plateau was defined as a change in \( \text{VO}_2 \) of < 0.4 mL·kg\(^{-1}\)·min\(^{-1}\) with an increase in speed. The running speed that elicited 50% \( \text{VO}_2\text{max} \) was calculated for each horse using linear regression analysis of speeds below \( \text{VO}_2\text{max} \).

**Diet composition.** Horses were fed a basal diet of mixed alfalfa and grass hay and pelleted concentrate to maintain ideal body weight (condition score 5–6 out of 9). Corn and alfalfa cubes were fed beginning 4 wk before the start of the experimental trials so that horses were accustomed to the diet. The diet consisted of 4.8 kg of alfalfa hay cubes, 2.9 kg of mixed hay, 1.4 kg of pelleted concentrate, and 1 kg of cracked corn per day for a 450-kg horse. The amounts of hay and grain were given following the guidelines published by the National Research Council for horses in moderate work (28). The equation used for estimation of daily requirements of digestible energy (Mcal of DE/day) was

\[
\text{DE} = 1.5 \cdot (1.4 + 0.03 \cdot \text{BW})
\]

where DE is the daily requirement of digestible energy (Mcal/day) and BW is the body weight (kg). The roughage and concentrates in the diet were analyzed by a commercial laboratory (Holmes Laboratory, Millersburg, OH) (Table 1). **Experimental protocol.** Before each experimental trial, food was withheld for 18 h, and the horses were confined to their stalls for 24 h. All experiments began at 0700 when horses were offered alfalfa cubes or at 0800 when horses were offered cracked corn. After aseptic preparation and injection of local anesthesia of the overlying skin, catheters (14 gauge, 5.25 in.; Angiocath, Becton Dickinson) were inserted into the right and left jugular veins for isotope infusion and blood sample collection, respectively. Thereafter, a blood sample was obtained for subsequent determination of background isotopic enrichment. In the grain trial, horses were fed, at 0800 in their stall, a meal of cracked corn corresponding to one-quarter of the daily energy requirements as previously established for horses on moderate work (28). In the hay trial, horses were fed, at 0700 in their stall, an isocaloric meal.
of alfalfa cubes. In the feed-withholding trial, horses were not fed before exercise. In all trials, horses were kept undisturbed (with the exception of blood sampling from the jugular catheter) in the stall until 0900. Horses were then moved to stockades in a temperature-controlled building. Glucose kinetics was determined by a primed (17.5 µmol/kg), continuous (0.22 ± 0.01 µmol·kg⁻¹·min⁻¹) infusion of [6,6-²H]glucose (99% enriched; Cambridge Isotopes, Cambridge, MA) in 0.9% saline. The [6,6-²H]glucose was infused for 90 min with a calibrated infusion pump while the horses stood quietly in the stocks. Fifteen minutes before initiation of exercise, a sample of the middle gluteal muscle was obtained by percutaneous biopsy. Thereafter, the horses were positioned on an inclined treadmill (4° incline), and a loose-fitting face mask was secured around the sides of the treadmill to maintain an air flow through the system. Flow through the system was measured (0.66 ± 0.02 µmol·kg⁻¹·min⁻¹) during the warm-up. During the exercise test, fans mounted 0.5 m in front and to the sides of the treadmill were used to maintain an air velocity of ∼4 m/s over the horse. Ambient conditions were similar for all trials (room temperature and relative humidity were 15–20°C and 50–60%, respectively).

**Respiratory exchange measurements.** V̇O₂, carbon dioxide production (V̇CO₂), and respiratory exchange ratio (RER) were measured with an open-circuit calorimeter (Oxymax-XXL, Columbus Instruments, Columbus, OH) as previously described (19). Flow through the system was ∼1,500 l/min STP when the horse was stationary and 9,000 l/min during running. Data for expired O₂ (electrochemical cell, Columbus Instruments) and CO₂ (single-beam nondispersive infrared sensor, Columbus Instruments) concentrations were measured continuously and were reported at 10-s intervals. The gas-analysis system was calibrated before the start of each exercise test by using gas mixtures with oxygen and carbon dioxide concentrations that spanned the measurement range. The overall accuracy of the system for the measurement of V̇O₂ was verified for each run by the nitrogen-dilution method, and measured values were corrected for any discrepancy (10). The accuracy of the system for the measurement of RER was verified for each run by burning of propane, which has a respiratory quotient of 0.6, and measured values were corrected for any discrepancy. Standard equations were used to calculate V̇O₂ and V̇CO₂, and RER was obtained by dividing V̇CO₂ by V̇O₂.

**Sampling procedures.** Blood samples were obtained at ~60 and ~30 min for hay trials and at 0, 30, 60, 90, 120, 135, 150, 155, 165, 180, 195, and 210 min for all trials (where the “0”-min sample was collected at 0800 when the first sample was obtained in grain and feed-withholding trials). Blood samples were divided (6-ml aliquots) into four different tubes for subsequent analysis. Two aliquots of each sample were placed in evacuated tubes containing EDTA. These samples were later analyzed for plasma isotopic enrichment, hematocrit, plasma total protein, NEFA, and glycerol concentrations. A 6-ml aliquot was placed in a tube containing sodium fluoride-potassium oxalate for subsequent determination of plasma glucose and lactate concentration. The final aliquot was placed in a tube containing no additive for measurement of serum immunoreactive insulin (IRI) concentration. Plasma or serum was obtained by centrifugation (1,500 g for 20 min at 4°C) within 30 min of collection and frozen at −80°C until analysis.

**Muscle biopsy samples** were collected percutaneously from the middle gluteal muscle via the needle-biopsy technique (26). Muscle biopsies were obtained 15 min before commencement of exercise and within 3 min of cessation of exercise. Muscle samples were immediately placed in liquid nitrogen and stored at −80°C until analysis.

**Plasma isotopic enrichment.** Plasma [6,6-²H]glucose enrichment was determined as previously described and performed by our laboratory (14). The intra- and interassay coefficients of variation were 1.6 ± 0.1 and 2.7 ± 0.4, respectively. To control for between-day variability in plasma [6,6-²H]glucose enrichment determination, all samples for a given horse (feed-withholding, hay, and grain trials) were analyzed during the same analytic session. All samples were analyzed in duplicate.

**Plasma biochemical analyses.** Plasma glucose concentration was measured spectrophotometrically using a commercial kit that employs the hexokinase reaction (glucose hexokinase kit, Sigma Diagnostics, St. Louis, MO), and plasma lactate concentration was measured by using an automated lactate oxidase method (Sprint 1500 lactate analyzer, Yellow Springs Instruments, Yellow Springs, OH). Plasma NEFA concentration was determined by using a commercial kit that employs an enzymatic colorimetric method (NEFA test kit, Wako Chemicals, Dallas, TX). Plasma glycerol concentration was measured by using an enzymatic spectrophotometric method (triglycerides kit 337A [without triglyceride hydrolase step], Sigma Diagnostics). Plasma glucose, NEFA, and glycerol concentrations were measured by using a microplate reader spectrophotometer (Versamax, Molecular Devices, Sunnyvale, CA). Intra- and interassay coefficients of variation for measurement of plasma glucose, lactate, NEFA, and

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**Table 1. Result of analysis of the dietary components**

<table>
<thead>
<tr>
<th></th>
<th>Alfalfa Cubes</th>
<th>Cracked Corn</th>
<th>Mixed Hay</th>
<th>Pelleted Concentrate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dry matter, %</td>
<td>86.1</td>
<td>85.9</td>
<td>87.3</td>
<td>86.4</td>
</tr>
<tr>
<td>Crude protein, %</td>
<td>13.4 (15.6)</td>
<td>8.04 (9.36)</td>
<td>16.2 (18.5)</td>
<td>12.7 (14.7)</td>
</tr>
<tr>
<td>ADF, %</td>
<td>34.0 (39.8)</td>
<td>2.76 (3.21)</td>
<td>34.4 (39.4)</td>
<td>11.5 (13.3)</td>
</tr>
<tr>
<td>NDF, %</td>
<td>43.3 (50.2)</td>
<td>7.48 (8.7)</td>
<td>40.3 (46.2)</td>
<td>20.6 (23.8)</td>
</tr>
<tr>
<td>Lignin, %</td>
<td>6.11 (7.09)</td>
<td>0.15 (0.17)</td>
<td>5.27 (6.04)</td>
<td>2.02 (2.34)</td>
</tr>
<tr>
<td>Starch (estimated), %</td>
<td>3.26 (3.79)</td>
<td>64.1 (74.6)</td>
<td>3.69 (4.23)</td>
<td>61.8 (71.5)</td>
</tr>
<tr>
<td>Crude fat, %</td>
<td>2.33 (2.7)</td>
<td>3.44 (4.0)</td>
<td>2.36 (2.7)</td>
<td>3.02 (3.5)</td>
</tr>
<tr>
<td>Ash, %</td>
<td>8.08 (9.38)</td>
<td>0.86 (1.0)</td>
<td>5.74 (6.58)</td>
<td>5.22 (6.04)</td>
</tr>
<tr>
<td>Calcium, %</td>
<td>1.01 (1.17)</td>
<td>0.02 (0.02)</td>
<td>1.12 (1.28)</td>
<td>0.53 (0.61)</td>
</tr>
<tr>
<td>Phosphorus, %</td>
<td>0.28 (0.32)</td>
<td>0.21 (0.25)</td>
<td>0.23 (0.26)</td>
<td>0.54 (0.62)</td>
</tr>
<tr>
<td>DE, Mcal/kg feed</td>
<td>1.85 (2.16)</td>
<td>3.32 (3.87)</td>
<td>2.09 (2.40)</td>
<td>2.73 (3.15)</td>
</tr>
</tbody>
</table>

Values are as sampled and, in parentheses, as a dry matter basis. ADF, acid detergent fiber; NDF, neutral detergent fiber; DE, digestible energy.
glycerol were ~1.0 and ~2.5%, respectively. Hematocrit was measured by the microhematocrit technique. Plasma total protein was measured by refractometry (Cambridge Instruments, Buffalo, NY). To control for between-day variability, all samples for a given horse (feed-withholding, hay, and grain trials) were analyzed during the same analytic session. All samples were analyzed in duplicate.

**Plasma hormone analyses.** Serum IRI was determined in duplicate by use of a commercially available RIA (insulin kit, Coat-a-Count Diagnostics, Los Angeles, CA) that has been validated for horse blood (33). Intra- and interassay coefficients of variation were ~6 and ~8%, respectively.

**Muscle glycogen.** Muscle samples were dissected free of any blood and connective tissue, and duplicate samples, of 20 mg each, were analyzed. The samples were extracted and analyzed for muscle glycogen concentration (as glucosyl units per kg of wet weight) according to the procedure of Pansonneau and Launderdale (32). Bovine liver glycogen (G0885, Sigma Diagnostics) was used to obtain known standard concentrations of 200 and 50 mM, as a quality control for the acid hydrolysis procedure. Intra- and interassay coefficients of variation were 9.4 and 12.3%, respectively.

**Calculations of glucose kinetics.** Glucose rate of appearance (Ra) and rate of disappearance (Rd) at rest were calculated by using the steady-state tracer dilution equation (48)

\[Ra = Rd = F \cdot \frac{[IE/IE_p] - 1}{E}\]

where \(F\) is the infusion rate of the isotope (\(\mu\)mol·kg\(^{-1}\)·min\(^{-1}\)); \(IE\) and \(IE_p\) are the stable isotopic enrichment of the infused and plasma, respectively; and \(-1\) accounts for the tracer’s contribution to the turnover rate of the substrate (48). The rate of infusion was calculated by multiplying the infusion pump rate by the concentration of glucose in the plasma (48). The rate of infusion was calculated by multiplying the infusion pump rate by the concentration of glucose in the plasma (48).

The rate of infusion was calculated by multiplying the infusion pump rate by the concentration of glucose in the plasma (48). The rate of infusion was calculated by multiplying the infusion pump rate by the concentration of glucose in the plasma (48).

\[Ra = \frac{dC_m}{dt} \left(1 + \frac{dE}{dt}\right) - \frac{C_m}{1 + \frac{dE}{dt}}\]

where \(V_d\) is the effective volume of distribution, \(E\) is the plasma isotopic enrichment, \(C_m\) is the measured plasma concentration of the tracee, and \(dE/dt\) and \(dC_m/dt\) are maximum rates of change in enrichment and glucose concentration, respectively, as a function of time. With use of this fixed, one-compartment model of Steele, it is assumed that 1) the apparent glucose space is 25% of body weight and 2) 65% of this space represents the rapidly mixing portion of the glucose pool. Therefore, the effective \(V_d\) for glucose was assumed to be 162 ml/kg. Glucose metabolic clearance rate (MCR) was calculated by dividing glucose \(Rd\) by the plasma glucose concentration. Glucose \(Ra\) was assumed to represent glucose absorption from the gastrointestinal tract plus hepatic glucose production, although a small contribution from renal glycogenolysis and gluconeogenesis is possible.

Rates of energy expenditure and whole body substrate oxidation. Total energy expenditure (TEE) and absolute rates of \(\text{CHO}_{\text{ox}}\) and lipid oxidation were calculated as follows (11, 12)

\[\text{TEE (kcal/min)} = 3.9 \dot{V}O_2 + 1.1 \dot{V}CO_2\]

\[\text{CHO}_{\text{ox}} (g/min) = 4.585 \dot{V}CO_2 - 3.225 \dot{V}O_2\]

where \(\dot{V}O_2\) is in liters per minute and it was assumed that protein oxidation made a negligible contribution to \(\dot{V}O_2\) and \(\dot{V}CO_2\) (i.e., nonprotein RER). The calculated values were based on respiratory gas exchange values averaged over 5-min intervals. \(\text{CHO}_{\text{ox}}\) in grams per minute was converted to micromoles per kilogram per minute by dividing the molecular weight of glucose (mol wt 180.16) and the horse’s body weight. Similarly, rates of fat oxidation were converted to micromoles per kilogram per minute by dividing the molecular weight of palmitate (mol wt 256.43) and the horse’s body weight. Muscle glycogen (plus lactate) oxidation was calculated as the difference between total \(\text{CHO}_{\text{ox}}\) and glucose \(Rd\). Coggan et al. (3) reported that, in human subjects, >95% of glucose \(Rd\) is oxidized during submaximal exercise. Therefore, glucose \(Rd\) provides a reasonable estimate of plasma glucose oxidation during exercise. Finally, the absolute and relative contributions by plasma glucose, other \(\text{CHO}\) sources (muscle glycogen and lactate), and lipid to the TEE during the 0- to 30- and the 35- to 60-min periods of exercise were estimated using standard calorific equivalents (4.2 kcal/g CHO, 9.0 kcal/g lipid).

**Statistical analyses.** Values are means ± SE. The data for all dependent measures were analyzed using a two-way ANOVA for repeated measures, with treatment (feed with- holding, hay, and grain) and time as independent factors. Percent data were subject to arcsine transformation before ANOVA. The null hypothesis was rejected at \(P \leq 0.05\) for the main effects (treatment and time) and \(\alpha = 0.10\) for the interaction. Significant differences identified by ANOVA were isolated using the Tukey post hoc test. The only dependent variable analyzed with a paired Student’s \(t\)-test was the difference in the relative feed consumptions between hay and grain meals. The Sigmanstat 2.0 software package (Jandel Scientific, San Rafael, CA) was used for statistical computations.

**RESULTS**

Individual values for \(\dot{V}O_2_{\text{max}}\) ranged from 109 to 125 ml·kg\(^{-1}\)·min\(^{-1}\) (mean \(117 ± 2.7\) ml·kg\(^{-1}\)·min\(^{-1}\)). Mean running speed on a 4° inclined treadmill during the exercise protocol was 4.9 ± 0.2 m/s (range 4.3–5.5 m/s), which corresponded to a relative workload of 50 ± 0.7% of \(\dot{V}O_2_{\text{max}}\) (range 46.9–52.4%).

**Preexercise feed consumption.** Before exercise in hay meal trials, horses were offered 2.9 ± 0.1 kg (6.3 ± 0.2 lb.; range 2.7–3.3 kg) and consumed 82 ± 8% (range 64–100%) of the meal. In grain meal trials, horses were offered 1.6 ± 0.04 kg (3.5 ± 0.1 lb.; range 1.5–1.8 kg) and all horses consumed all of the meal. The difference in caloric intake between the two meals was ~10 ± 3.9 kcal DE/kg of body weight (2.4 ± 0.9 kcal/kg) (range 0–19 kcal/kg). The meal consumption in hay trials had a tendency to be lower than in grain trials (\(P = 0.06\)).

**Plasma glucose concentration.** Feeding type before exercise significantly affected plasma glucose concentrations before and during exercise. Before ingestion of a meal, plasma glucose was similar among the three trials (Fig. 1A). During the 150-min period, after the horses were allowed to eat a grain meal equivalent to...
one-quarter of the daily energy requirements, glucose concentration steadily increased from 4.4 \pm 0.1 \text{ mM} before the meal to 6.6 \pm 0.7 \text{ mM} at 150 min (Fig. 1A). Plasma glucose concentration remained unchanged in horses fed the hay meal and in which the feed was withheld. During exercise in feed-withholding trials, plasma glucose concentration increased steadily to reach a peak of 9.0 \pm 0.1 \text{ mM}, whereas in grain trials, plasma glucose concentration decreased from preexercise values during the first 15 min of exercise and were subsequently similar to values in hay trials. Plasma glucose concentrations were higher \((P < 0.05)\) in feed-withholding trials than in hay or feed-withholding trials during the second half of exercise.

Serum insulin. Feeding type before exercise significantly altered serum IRI concentrations before and during exercise. Allowing horses to eat a hay meal induced an insulinemic response in which IRI increased over 60\% (127.1 \pm 17.5 \text{ pM} at 30 min) from IRI concentrations before feeding (72.7 \pm 6.1 \text{ pM} at -60 min; \(P < 0.001\); Fig. 1B). Serum IRI concentrations steadily increased after a grain meal from 67.2 \pm 4.9 pM at 0 min to reach a peak of 175.6 \pm 17.4 \text{ pM} 150 min after the meal was offered \((P < 0.001)\). During exercise, serum IRI decreased in all trials but in grain meal trials remained elevated above values in feed-withholding trials for the first 15 min of exercise \((P < 0.05)\). No differences in serum IRI concentration were found among trials between 30 and 60 min of exercise.

NEFA and glycerol. Plasma glycerol concentrations were affected by feeding type during exercise but not before exercise. Plasma glycerol concentrations during the second half of exercise were significantly lower in grain trials than in hay or feed-withholding trials. Plasma glycerol concentration increased steadily during exercise in all trials from \(-0.1 \pm 0.0 \text{ mM}\) to reach a peak of 1.22 \pm 0.1, 1.13 \pm 0.1, and 0.96 \pm 0.1 \text{ mM} at 60 min of exercise for feed-withholding, hay, and grain trials, respectively \((P < 0.001; 150 \text{ vs. } 210 \text{ min}; \text{Fig. } 2A)\). Feeding type before exercise altered plasma NEFA concentrations before and during exercise. During the 120-min period in which horses were allowed to eat hay, plasma NEFA steadily decreased from 0.90 \pm 0.2

Fig. 1. Plasma glucose (A) and serum immunoreactive insulin (B) at rest and during 60 min of exercise at 50 \pm 0.7\% of maximal oxygen uptake after horses had feed withheld (withholding feed), were fed a hay meal (hay), or were fed an approximately isocaloric grain meal (grain) 90 min before exercise. Values are means \pm SE for 6 horses. *Grain meal significantly different from withholding feed, \(P < 0.05\). # Hay meal significantly different from withholding feed, \(P < 0.05\). & Grain meal significantly different from hay meal, \(P < 0.05\).

Fig. 2. Plasma glycerol (A) and nonesterified fatty acids (NEFA; B) at rest and during 60 min of exercise at 50 \pm 0.7\% of maximal oxygen uptake after withholding feed, feeding hay, or feeding grain 90 min before exercise. Values are means \pm SE for 6 horses. *Grain meal significantly different from withholding feed, \(P < 0.05\). # Hay meal significantly different from withholding feed, \(P < 0.05\). & Grain meal significantly different from hay meal, \(P < 0.05\).
mM before the hay meal to 0.27 ± 0.1 mM at 60 min (P < 0.001). NEFA concentration gradually decreased after both hay and grain meal trials, whereas in feed-withholding trials, it initially remained unchanged and gradually increased (Fig. 2B). Plasma NEFA concentration decreased sharply at commencement of exercise in feed-withholding trials (0.95 ± 0.2 mM immediately before exercise and 0.40 ± 0.1 mM at 5 min of exercise; P = 0.003; Fig. 2B). Plasma NEFA concentrations during exercise were significantly lower in the grain trial than in the feed-withholding and hay trials (P < 0.05).

Respiratory gas exchange and whole body substrate oxidation. Feeding type before exercise altered RER and the whole body rates of CHO ox and lipid oxidation during exercise. There was a small but significant (P < 0.001) increase in Vo2 during the 60 min of treadmill exercise, and Vo2 was similar among the three trials (Table 2). RER decreased significantly in all three trials over the 60 min of exercise (P < 0.001), and at 60 min of exercise grain trials had a significantly higher RER compared with hay trials (0.89 ± 0.01 and 0.84 ± 0.01, grain vs. hay; P = 0.021). TEE steadily increased over time from 126.1 ± 3.3, 127.8 ± 5.6, and 129.1 ± 4.1 kcal/min at 5 min to 143.1 ± 2.9, 138.3 ± 2.6, and 140.2 ± 2.6 kcal/min at 60 min for feed-withholding, hay, and grain trials, respectively (P < 0.001; Table 2). Total CHO ox decreased during exercise in feed-withholding and hay trials but remained stable in grain trials. Total CHO ox was higher in grain than in hay trials at 60 min of exercise (327 ± 26 vs. 239 ± 26 μmol·kg⁻¹·min⁻¹, grain vs. hay; P = 0.023). Conversely, fat oxidation was suppressed in grain trials compared with feed-withholding and hay trials, and at 60 min the rate of fat oxidation was significantly lower in grain trials (49 ± 4 μmol·kg⁻¹·min⁻¹) compared with hay trials (72 ± 8 μmol·kg⁻¹·min⁻¹; P = 0.016).

Glucose kinetics. Feeding type before exercise altered plasma glucose isotopic enrichment and glucose Ra before and during exercise, and it altered glucose R a before exercise as well as glucose MCR during exercise. Before exercise, plasma isotopic enrichment (Fig. 3) was lower in grain trials than in feed-withholding and hay trials (P < 0.001). During the first 5 min of exercise, plasma isotopic enrichment increased in all trials; however, in grain trials, plasma isotopic enrichment increased more steeply (P < 0.001).

Table 2. Rate of oxygen consumption, respiratory exchange ratio, rate of carbohydrate and lipid oxidation, calculated muscle glycogen (and lactate) oxidation, and rate of glucose disappearance during 60 min of exercise at 50 ± 0.7% of Vo₂ max

<table>
<thead>
<tr>
<th>Time, min</th>
<th>5</th>
<th>15</th>
<th>30</th>
<th>45</th>
<th>60</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vo2, ml·kg⁻¹·min⁻¹</td>
<td>Feed withholding 60 ± 1 64 ± 1 67 ± 2 67 ± 2 69 ± 2</td>
<td></td>
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<tr>
<td>Hay 62 ± 3 63 ± 2 66 ± 1 68 ± 1 68 ± 1</td>
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<tr>
<td>Grain 61 ± 2 63 ± 2 66 ± 2 67 ± 2 67 ± 2</td>
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<tr>
<td>RER Feed withholding 0.92 ± 0.02 0.90 ± 0.01 0.88 ± 0.01 0.87 ± 0.01 0.87 ± 0.01</td>
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<tr>
<td>Hay 0.90 ± 0.01 0.89 ± 0.01 0.87 ± 0.01 0.85 ± 0.01 0.84 ± 0.01</td>
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<tr>
<td>Grain 0.93 ± 0.02 0.92 ± 0.02 0.90 ± 0.02 0.89 ± 0.01 0.89 ± 0.01</td>
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<tr>
<td>CHO oxidation, μmol·kg⁻¹·min⁻¹ (cal·kg⁻¹·min⁻¹) Feed withholding 339 ± 28 318 ± 24 297 ± 27 289 ± 18 287 ± 18</td>
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<tr>
<td>(226 ± 18) (212 ± 16) (198 ± 18) (193 ± 12) (191 ± 12)</td>
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<tr>
<td>Hay 313 ± 19 309 ± 28 282 ± 24 258 ± 26 239 ± 26</td>
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<tr>
<td>(209 ± 12) (206 ± 18) (188 ± 16) (172 ± 17) (159 ± 17)</td>
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<tr>
<td>Grain 352 ± 30 357 ± 32 337 ± 33 325 ± 26 327 ± 26</td>
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<tr>
<td>(234 ± 20) (238 ± 21) (225 ± 22) (217 ± 17) (218 ± 18)</td>
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<tr>
<td>Lipid oxidation, μmol·kg⁻¹·min⁻¹ (cal·kg⁻¹·min⁻¹) Feed withholding 32 ± 6 (76 ± 14) 44 ± 4 (106 ± 9) 56 ± 5 (134 ± 11) 59 ± 3 (140 ± 8) 63 ± 1 (150 ± 3)</td>
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<tr>
<td>Hay 41 ± 4 (97 ± 9) 46 ± 6 (109 ± 13) 58 ± 5 (139 ± 13) 67 ± 6 (160 ± 14) 72 ± 8 (172 ± 19)</td>
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<tr>
<td>Grain 30 ± 8 (72 ± 19) 33 ± 7 (79 ± 17) 45 ± 6 (103 ± 15) 48 ± 4 (115 ± 11) 49 ± 4 (117 ± 10)</td>
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<tr>
<td>Glycogen (and lactate) oxidation, μmol·kg⁻¹·min⁻¹ (cal·kg⁻¹·min⁻¹) Feed withholding 319 ± 26 281 ± 25 261 ± 27 260 ± 18 246 ± 17</td>
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<tr>
<td>(211 ± 18) (184 ± 17) (171 ± 18) (171 ± 12) (161 ± 11)</td>
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<tr>
<td>Hay 284 ± 15 269 ± 29 247 ± 25 230 ± 28 207 ± 26</td>
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<tr>
<td>(185 ± 8) (175 ± 22) (161 ± 16) (151 ± 19) (135 ± 19)</td>
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<tr>
<td>Grain 320 ± 31 301 ± 33 294 ± 32 290 ± 26 288 ± 24</td>
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<tr>
<td>(209 ± 17) (195 ± 22) (192 ± 21) (190 ± 17) (189 ± 17)</td>
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<tr>
<td>Rₐ, μmol·kg⁻¹·min⁻¹ (cal·kg⁻¹·min⁻¹) Feed withholding 20 ± 3 (15 ± 2) 37 ± 5 (28 ± 4) 36 ± 5 (27 ± 4) 29 ± 4 (22 ± 3) 38 ± 4 (29 ± 3)</td>
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<tr>
<td>Hay 39 ± 7 (30 ± 5) 40 ± 3 (30 ± 2) 35 ± 3 (27 ± 2) 28 ± 4 (22 ± 3) 33 ± 4 (25 ± 3)</td>
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<tr>
<td>Grain 49 ± 5 (37 ± 3)† 56 ± 4 (42 ± 3)† 43 ± 4 (32 ± 3) 36 ± 3 (27 ± 2) 39 ± 6 (39 ± 4)</td>
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</table>

Values are means ± SE for 6 horses. Horses had food withheld (food withholding), were fed a hay meal (hay), or were fed a grain meal (grain) 90 min before exercise. Rates of oxygen consumption (Vo2) and respiratory exchange ratio (RER) were calculated from 5-min average values. Rates of carbohydrate (CHO) and fat oxidation were calculated from respiratory gas exchange data, averaged over 5 min. Muscle glycogen (plus lactate) oxidation was estimated from the difference between total carbohydrate oxidation and rate of glucose disappearance. Values in parentheses are the relative caloric contributions to energy expenditure from oxidation of CHO, lipid, muscle glycogen (and lactate), and blood-borne glucose. *P < 0.05, grain vs. hay. †P < 0.05, grain vs. feed withholding. ‡P < 0.05, hay vs. feed withholding.

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values remained stable, whereas in feed-withholding and hay trials, plasma isotopic enrichment steadily decreased despite the threefold increase in tracer infusion rate.

At rest, the mean glucose $R_a$ (Fig. 4A) and $R_d$ (Fig. 4B) during the grain trials were both ~70% greater than during feed-withholding and hay trials (both $R_a$ and $R_d, P < 0.005$). Glucose $R_a$ increased steadily in all trials from $7.9 \pm 0.7$, $8.1 \pm 0.6$, and $13.6 \pm 1.6$ mol·kg$^{-1}$·min$^{-1}$ to reach a peak of $60 \pm 6$, $52 \pm 4$, and $53 \pm 7$ mol·kg$^{-1}$·min$^{-1}$ for feed-withholding, hay, and grain trials, respectively, at 30 min of exercise ($P < 0.001$). During the second half of the exercise trial, $R_a$ remained stable or decreased slightly in all trials.

During exercise, there was a significant trial effect ($P = 0.007$) for glucose $R_d$. Whereas glucose $R_d$ increased with the onset of exercise in all trials, the increase was much larger in grain trials (Fig. 4B). In feed-withholding and hay trials, glucose $R_d$ increased from $7.9 \pm 0.7$ and $8.1 \pm 0.6$ mol·kg$^{-1}$·min$^{-1}$ before exercise to reach a peak of $37 \pm 5$ and $40 \pm 3$ mol·kg$^{-1}$·min$^{-1}$, respectively, at 15 min of exercise ($P < 0.001$) and remained stable or decreased slightly during the rest of the exercise period. In grain trials, glucose $R_d$ increased sharply from $13.6 \pm 1.6$ mol·kg$^{-1}$·min$^{-1}$ before exercise to reach a peak of $56 \pm 4$ mol·kg$^{-1}$·min$^{-1}$ at 15 min of exercise ($P < 0.001$), and it steadily decreased during the rest of the exercise. Glucose MCR demonstrated a similar pattern (Fig. 5). During exercise MCR was ~50–100% higher in grain trials than in feed-withholding trials ($P < 0.05$ at 5, 15, and 30 min), whereas only at 5 min was MCR higher in hay trials compared with feed-withholding trials ($P < 0.05$).

Muscle glycogen. Preexercise muscle glycogen concentration was similar among the three trials (Fig. 6). Net muscle glycogen utilization during exercise was similar ($P = 0.6$) among the three trials ($47 \pm 9$, $44 \pm 11$, and $35 \pm 8$ mmol/kg wet muscle in feed-withholding, hay, and grain trials, respectively). Postexercise muscle glycogen concentration was lower compared with preexercise samples in all trials ($P < 0.001$). The calculated rates of muscle glycogen oxidation (total $\text{CHO}_{\text{ox}} - \text{glucose } R_d$) during exercise were not affected by meal type (Table 2).

Hematocrit, plasma total protein, and lactate. Feeding type before exercise altered hematocrit and plasma total protein concentration before exercise but did not affect plasma lactate concentration. Hematocrit was significantly increased by 5 min of exercise and remained elevated throughout exercise in all trials (35–37% immediately before exercise and 49–50% at 5 min of exercise; $P < 0.001$; Table 3). During the resting period in hay meal trials, hematocrit was higher than in grain meal and feed-withholding trials (feeding × time, $P < 0.059$). Plasma total protein concentration followed a pattern similar to that for hematocrit. Plasma total protein concentration increased signifi-
Gen was attenuated in grain trials, and a tendency was observed in grain trials to have a higher muscle glycogen contribution compared with hay trials ($P = 0.11$). In hay trials, during the second half of exercise, the relative caloric contribution of fat oxidation was higher compared with grain trials ($48 \pm 4$ and $34 \pm 4\%$; $P < 0.05$, hay vs. grain).

**DISCUSSION**

The present study examined the effects of a HGM (grain trial), an approximately isocaloric LGM (hay trial), or feed-withholding before exercise on CHO and lipid metabolism in horses during moderate-intensity exercise. The main findings were: 1) a ~50% increase in whole body $R_d$ occurs during the first 30 min of moderate-intensity exercise after feeding a corn meal before exercise, compared with exercise after withholding feed for 18 h; 2) feeding a hay or grain meal, irrespective of the glycemic index of a meal fed before exercise, decreases plasma glucose concentration during exercise, compared with exercise after withholding feed; 3) augmentation of $\text{CHO}_{ox}$ and attenuation of lipid oxidation occurred during exercise in trials preceded by corn feeding; and 4) feeding status before exercise does not affect net muscle glycogen utilization.

As expected, feeding a HGM before exercise resulted in marked hyperglycemia (Fig. 1A) and hyperinsulinemia (Fig. 1B) before the exercise bout. These results are consistent with findings of previous studies of the effect of feeding of different grains and/or roughages on plasma metabolite and hormone concentrations in horses (29, 30, 34, 43). Stull and Rodiek (42) showed that a corn meal that provides 25% of the energy requirements induces a ~50% increase in plasma glucose concentration that peaks between 2 and 3 h after eating and a sixfold increase in serum IRI concentration that peaks 2 h after eating. However, in the same study, an isocaloric amount of alfalfa hay did not induce an increase in plasma glucose and serum IRI concentrations. This is in contrast to our findings, in which hay feeding did not result in an increase in

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**Fig. 5.** Glucose metabolic clearance rate (MCR) at rest and during 60 min of exercise at $50 \pm 0.7\%$ of maximal oxygen uptake after withholding feed, feeding hay, or feeding grain 90 min before exercise. Values are means ± SE for 6 horses. *Grain meal significantly different from withholding feed, $P < 0.05$. #Hay meal significantly different from withholding feed, $P < 0.05$.

**Fig. 6.** Muscle glycogen concentrations before (Pre) and after (Post) 60 min of exercise at $50 \pm 0.7\%$ of maximal oxygen uptake after withholding feed, feeding hay, or feeding grain 90 min before exercise. Values are means ± SE for 6 horses. ww, Wet weight. *Significant difference compared with preexercise values, $P < 0.05$. **
plasma glucose but hay consumption resulted in a 60% increase in serum IRI concentration. The meals fed in this study achieved our aim of offering meals of different glycemic indexes to horses before exercise.

In this study, the increase in plasma glucose concentration induced by corn feeding was accompanied by increments in glucose Ra and Rd. Glucose Ra was similar in horses that had food withheld and that were fed hay and was similar to previous estimations by intravenous [2-3H]glucose infusion of the total glucose production in two ponies fed hay (9.8 μmol·kg⁻¹·min⁻¹) (39). During exercise, glucose Rd was significantly higher in trials preceded by ingestion of a grain meal compared with trials preceded by a hay meal or withholding feed. Similarly, in humans, ingestion of a HGM (mashed potatoes) or glucose solution before moderate-intensity exercise results in higher glucose Ra and Rd before and during exercise, compared with LGM (muesli), placebo, or fasting (8, 22, 27). However, preexercise glucose ingestion markedly decreases hepatic glucose production during exercise, and the increase in glucose Ra reflects ongoing intestinal uptake of glucose (27). In the present study, we did not measure the contribution of gut-derived Ra (intestinal absorption of glucose) to the total Ra. Nonetheless, it is likely that continued absorption of glucose from the gastrointestinal tract contributed to the higher glucose Ra before exercise in horses fed a grain meal. During the first half of exercise in horses fed grain, because the

Table 3. Hematocrit and plasma total protein and lactate concentrations before and during 60 min of exercise at 50 ± 0.7% of VO₂max

<table>
<thead>
<tr>
<th>Time, min</th>
<th>0</th>
<th>90</th>
<th>150</th>
<th>155</th>
<th>165</th>
<th>180</th>
<th>195</th>
<th>210</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hematocrit, %</td>
<td>Feed withholding</td>
<td>38 ± 2</td>
<td>35 ± 1</td>
<td>35 ± 2</td>
<td>49 ± 1</td>
<td>50 ± 1</td>
<td>49 ± 1</td>
<td>48 ± 1</td>
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<tr>
<td>Hay</td>
<td>39 ± 1</td>
<td>39 ± 1</td>
<td>37 ± 1</td>
<td>49 ± 1</td>
<td>50 ± 1</td>
<td>49 ± 1</td>
<td>49 ± 1</td>
<td>49 ± 1</td>
</tr>
<tr>
<td>Grain</td>
<td>36 ± 1</td>
<td>34 ± 2</td>
<td>35 ± 1</td>
<td>50 ± 1</td>
<td>49 ± 1</td>
<td>48 ± 1</td>
<td>49 ± 1</td>
<td>50 ± 1</td>
</tr>
<tr>
<td>Plasma total protein, g/dl</td>
<td>Feed withholding</td>
<td>7.2 ± 0.1</td>
<td>7.1 ± 0.1</td>
<td>7.0 ± 0.1</td>
<td>7.6 ± 0.1</td>
<td>7.5 ± 0.1</td>
<td>7.4 ± 0.2</td>
<td>7.4 ± 0.1</td>
</tr>
<tr>
<td>Hay</td>
<td>7.5 ± 0.2</td>
<td>7.3 ± 0.2</td>
<td>6.9 ± 0.1</td>
<td>7.3 ± 0.1</td>
<td>7.4 ± 0.1</td>
<td>7.3 ± 0.2</td>
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<td>7.6 ± 0.2</td>
</tr>
<tr>
<td>Grain</td>
<td>6.9 ± 0.1</td>
<td>6.9 ± 0.1</td>
<td>6.8 ± 0.1</td>
<td>7.4 ± 0.2</td>
<td>7.2 ± 0.2</td>
<td>7.1 ± 0.2</td>
<td>7.3 ± 0.1</td>
<td>7.4 ± 0.2</td>
</tr>
<tr>
<td>Lactate, mM</td>
<td>Feed withholding</td>
<td>0.4 ± 0.0</td>
<td>0.4 ± 0.0</td>
<td>0.5 ± 0.0</td>
<td>0.7 ± 0.1</td>
<td>1.6 ± 0.1</td>
<td>2.9 ± 0.3</td>
<td>3.9 ± 0.4</td>
</tr>
<tr>
<td>Hay</td>
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<td>0.6 ± 0.0</td>
<td>0.6 ± 0.1</td>
<td>1.0 ± 0.1</td>
<td>2.0 ± 0.2</td>
<td>3.6 ± 0.6</td>
<td>4.2 ± 0.7</td>
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</tr>
<tr>
<td>Grain</td>
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<td>0.6 ± 0.1</td>
<td>0.6 ± 0.0</td>
<td>0.9 ± 0.1</td>
<td>1.5 ± 0.2</td>
<td>3.1 ± 0.4</td>
<td>4.0 ± 0.5</td>
<td>4.2 ± 0.5</td>
</tr>
</tbody>
</table>

Values are means ± SE for 6 horses. Values during exercise are from 155 to 210 min. VO₂max, maximal VO₂. *P < 0.05, grain vs. hay. †P < 0.05, hay vs. feed withholding; ‡P < 0.05, 165–210 min vs. 150 min. Hematocrit and plasma total protein from 155 to 210 min are higher than 150 min (P < 0.001) in all trials.

Fig. 7. Relative caloric contributions from oxidation of other carbohydrate (CHO; muscle glycogen and lactate), lipid, and blood glucose during the 5- to 30-min and 35- to 60-min periods of exercise after withholding feed, feeding hay, or feeding grain 90 min before exercise. Values are means ± SE for 6 horses. *Grain meal significantly different from withholding feed, P < 0.05. #Hay meal significantly different from withholding feed, P < 0.05. &Grain meal significantly different from hay meal, P < 0.05. For all trials, the percent energy expenditure of each substrate oxidation is significantly different at 5–30 min vs. 35–60 min (P < 0.05), except glucose utilization in trials after withholding feed.
increase in glucose \( R_d \) was not accompanied by a similar increase in glucose \( R_a \). Plasma glucose concentration decreased sharply. However, plasma glucose concentrations during exercise in horses fed grain were not different from in horses fed hay before exercise. This is in contrast to previous studies performed in horses (34, 43) in which horses fed corn before exercise had lower plasma glucose concentrations during exercise compared with those of horses fed alfalfa hay before exercise.

In horses that had feed withheld, blood glucose concentrations steadily increased during the exercise bout and were higher than in horses fed before exercise. These findings are consistent with previous studies in horses (24, 25, 29, 30, 43) but not in humans (8, 23, 37, 38, 40, 44). Endurance-trained human athletes cycling at a higher relative exercise intensity (85% \( \dot{V}O_2 \text{max} \)) but a similar absolute exercise intensity (58 ml \( O_2 \cdot kg^{-1} \cdot min^{-1} \)) had similar responses to exercise than the horses in this study (35). Human athletes exercising at an intensity that elicits a similar \( \dot{V}O_2 \) for the horses in this study had also a steady increase in blood glucose concentrations during the exercise bout (~3.9 mM before and ~8.2 mM during exercise) and had similar glucose \( R_d \) before and during exercise (~10 \( \mu \text{mol} \cdot kg^{-1} \cdot \text{min}^{-1} \) before and ~50 \( \mu \text{mol} \cdot kg^{-1} \cdot \text{min}^{-1} \) during exercise) and similar relative and absolute contributions from different substrates to energy expenditure during exercise (~180 cal \( \cdot kg^{-1} \cdot \text{min}^{-1} \) from muscle glycogen oxidation, ~80 cal \( \cdot kg^{-1} \cdot \text{min}^{-1} \) from muscle triglyceride and plasma NEFA oxidation, and ~40 cal \( \cdot kg^{-1} \cdot \text{min}^{-1} \) from plasma glucose oxidation) (35). These differences between human subjects and horses may be associated with the greater aerobic capacity of horses; in other words, when horses exercise at the same work intensity relative to \( \dot{V}O_2 \text{max} \) as human athletes, the \( \dot{V}O_2 \) and the workload are much greater in horses than in human athletes. (46) Therefore, when some metabolic responses to exercise are compared, it may be more meaningful to compare human athletes and horses when they are performing exercise tasks that elicit similar \( \dot{V}O_2 \).

Previous studies in horses have led to recommendations of feeding before exercise based on changes in substrate and hormone concentrations in plasma or serum (29, 30). Because of the exercise-induced decrease in plasma glucose concentration and reduced NEFA availability in horses fed grain before exercise, it has been recommended not to feed a grain meal before exercise (29, 30). In humans, ingestion of a HGM before and/or during exercise has resulted in enhanced (13, 23, 37, 38), decreased (44), or unchanged submaximal exercise performance (8, 9, 18, 40, 47). In horses, the effect of CHO ingestion on performance during moderate-intensity exercise has not been determined. Therefore, recommendations on interval and type of feeding before exercise can only be offered based on the metabolic responses induced by the interval of feeding and the composition of the meal. Determination of plasma or serum concentration of metabolites and substrates has been the sole means for providing recommendations of dietary manipulations in horses before exercise. These studies have provided little insight into the influence of feeding status before exercise on substrate supply and utilization during exercise. The combination of estimation of whole body \( \text{CHOox} \) and fat oxidation rates (by indirect calorimetry measurements) and estimation of rates of appearance and disappearance of metabolites in plasma (by measurement of infused metabolites labeled with stable isotopes) is the only means to determine the contribution of extramuscular and intramuscular substrates to energy production. To the best of our knowledge, the present study is the first one that estimates the contribution of extramuscular and intramuscular substrates to energy production in horses fed different meals or denied access to food before exercise.

Ingestion of corn before exercise resulted in a significant decrease in plasma glycerol concentration during exercise (Fig. 2A) and plasma NEFA concentration before and during exercise (Fig. 2B). Lower plasma NEFA concentrations in grain trials also may have contributed to the higher rate of glucose utilization in this trial compared with hay or feed withholding trials. In humans, preexercise CHO ingestion inhibits lipolysis during moderate-intensity exercise and suppresses the increase in plasma NEFA concentration (6, 20). Similarly, in horses, preexercise ingestion of corn attenuates the increase in fatty acid concentration during exercise (25, 42).

Feeding corn before exercise resulted in a greater relative contribution to energy expenditure from blood-borne glucose during the initial half of a 60-min moderate-intensity exercise event, compared with withholding feed, and it resulted in greater \( \text{CHOox} \) and lower lipid oxidation during the second half of the exercise bout, compared with hay feeding before exercise (Fig. 7). Similarly to the results in our present study, increased glucose availability during 60 min of running at 55% \( \dot{V}O_2 \text{max} \) by preexercise intragastric administration of glucose [2 g/kg, 10 kcal/kg DE (41.6 kJ/kg DE)] to horses resulted in enhanced \( \text{CHOox} \) and utilization of blood-borne glucose during moderate-intensity exercise but did not alter muscle glycolysis utilization (15). In studies performed in human subjects, ingestion of glucose or HGMs before exercise results in enhanced \( \text{CHOox} \) and utilization of blood-borne glucose, and the rate of glycolysis may be enhanced (18) or unchanged (9, 17, 22, 23). The same controversy is apparent in regard to the effect of increased glucose availability on muscle glycolysis in studies performed in horses. Lawrence et al. (24) reported that increased glucose availability by ingestion of a HGM before exercise enhances muscle glycolysis in exercising horses, compared with feed withholding (~46 vs. 18 mmol/kg wet wt in ~15 min exercise test of 1.6 km at 6 m/s, 0.8 km at 1.9 m/s (treadmill grade changed to 2%), 0.4-km speed-up period, and a final 1.6 km at 11 m/s]. This is in contrast to the findings of this study (Fig. 6), previous studies performed in our laboratory (14, 15), and studies by others (7), in which increased glucose availability by preexercise ingestion...
of grain, intragastric glucose administration, or CHO-rich diet, respectively, did not alter muscle glycogenolysis during exercise. However, in our study, a nonsignificant reduction of muscle glycogenolysis of ~25% was noted when horses were fed grain before exercise compared with feed withholding.

TEE during 60 min of exercise was ~19.3 kcal/kg (~80.4 kJ/kg), which is ~33.8 kcal/kg DE (~141 kJ/kg DE) assuming an efficiency of utilization of DE of 57% (31). When this is translated into National Research Council nutritional requirements (28) and the maintenance energy requirements are added (~33.1 kcal/kg DE (~138 KJ/kg DE)), there is an average total daily energy requirement of ~66.9 kcal/kg DE (~278.8 kcal/kg DE), an increase over maintenance of ~102%. Rose et al. (36) reported similar increases in daily energy requirement over maintenance energy requirements are added [278.8 kcal/kg DE (~112%) for horses exercised at 50% V˙O2 max](36); an increase over maintenance of 67% for 450-kg horses exercised for 2 h on a flat surface at 4.2 m/s by a 75-kg rider. These estimates of the increase in energy requirements for exercising horses may be useful in the future to develop more accurate guidelines of the nutritional requirements of these animals.

In summary, this study has demonstrated that feeding of a HGM before exercise augments CHOox and utilization of blood-borne glucose in horses during moderate-intensity exercise but does not alter muscle glycogen usage. Conversely, feeding of an isocaloric LGM with roughage before exercise, compared with a HGM, augments lipid oxidation in horses during moderate-intensity exercise but does not alter muscle glycogen usage. These findings support that feeding a HGM 2 h before exercise might be indicated in exercise bouts of moderate intensity in which CHOox is responsible for supplying more than half of the energy requirement for energy transduction during exercise.

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