HIGHLIGHTED TOPIC | Hypoxia

HIF and pulmonary vascular responses to hypoxia

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Shimoda LA, Laurie SS. Hif and pulmonary vascular responses to hypoxia. J Appl Physiol 116: 867–874, 2014. First published December 12, 2013; doi:10.1152/japplphysiol.00643.2013.—In the lung, acute reductions in oxygen lead to hypoxic pulmonary vasoconstriction, whereas prolonged exposures to hypoxia result in sustained vasoconstriction, pulmonary vascular remodeling, and the development of pulmonary hypertension. Data from both human subjects and animal models implicate a role for hypoxia-inducible factors (HIFs), oxygen-sensitive transcription factors, in pulmonary vascular responses to both acute and chronic hypoxia. In this review, we discuss work from our laboratory and others supporting a role for HIF in modulating hypoxic pulmonary vasoconstriction and mediating hypoxia-induced pulmonary hypertension, identify some of the downstream targets of HIF, and assess the potential to pharmacologically target the HIF system.

hypoxy inducible factor; pulmonary hypertension; hypoxic pulmonary vasoconstriction; digoxin

REDUCED OXYGEN AVAILABILITY is a common occurrence in the lung, arising in both physiological (i.e., fetal development) and pathological (i.e., chronic lung disease) settings. As oxygen sensing and swift response to oxygen deprivation are essential for survival, regulation of oxygen homeostasis has evolved into a complex system. In higher organisms, adaptation to hypoxia involves the synchronized transcriptional regulation of a variety of genes and is coordinated, in large part, by hypoxia-inducible factors (HIFs), oxygen-sensitive transcription factors that have been described as “master regulators of oxygen homeostasis” (53). In this review, we will describe the HIF system, the regulation of HIFs in the lung, and the mounting evidence supporting a role for HIFs in pulmonary vascular responses to hypoxia.

HYPOXIA-INDUCIBLE FACTORS

The discovery of the first HIF, HIF-1, a little over 20 years ago (67), paved the way for an explosion of research implicating activation of HIF as a vital step in a variety of physiological processes associated with hypoxia, including embryogenesis (13, 28, 34, 43, 59), wound healing (40), control of respiration (52), red blood cell production (64, 65), and glucose and energy metabolism (63, 65). HIF has also been shown to play a critical role in pathological scenarios, such as pulmonary (9, 100) and systemic (65) hypertension, cancer (31, 61), and ischemic preconditioning (62, 66).

Identified as a hypoxia-sensitive transcription factor that controlled the expression of erythropoietin (67), HIF-1 is now regarded as a highly conserved transcription factor that is present in almost all cell types and regulates the oxygen-dependent expression of hundreds of genes (63). HIF-1 exists as a heterodimer, formed from HIF-1α and HIF-1β subunits. Concerted research efforts have demonstrated that the activity of HIF-1 is tightly regulated by oxygen availability (30, 53, 65). Under conditions in which oxygen is abundant, HIF-1α is ubiquitously expressed, whereas HIF-1α protein expression is typically quite low or altogether absent. As oxygen levels fall, HIF-1α protein levels and HIF-1 DNA binding activity increase (63). For nearly a decade after the discovery of HIF-1, the mechanism by which cells sensed changes in oxygen concentration and transduced the signal into an increase in HIF-1 activity and induction of hypoxia-inducible genes was a matter of intense speculation and investigation. In 2001, several laboratories simultaneously reported that HIF-1α ubiquitination and targeting for degradation required hydroxylation of the protein (27, 29, 44, 101). Under normoxic conditions, hydroxylation of HIF-1α occurs at two proline residues (Pro402 and Pro564 in human HIF-1α) located within the oxygen-dependent degradation domain in a reaction catalyzed by prolyl hydroxylase domain proteins (PHDs) using molecular oxygen as a substrate (17, 27, 29). Four PHD isoforms have been identified, of which PHD1, PHD2, and PHD3 hydroxylate HIF-1α, with PHD2 responsible for the majority of HIF-1α hydroxylation in vivo (3, 7, 46). The PHDs appear to be developmentally regulated, with higher levels observed in adult compared with fetal animals (56). Despite low levels in the fetus, however, PHD2 is clearly required for in utero
development, since mice deficient for PHD2 die midgestation (85). In contrast, mice lacking PHD1 and PHD3 appear to develop normally and are viable (85).

Once hydroxylated, HIF-1α can interact with the tumor suppressor, von Hippel-Lindau (VHL) protein (Fig. 1), which recruits an E3 ubiquitin-protein ligase, allowing HIF-1α to be polyubiquitinated and targeted for 26S proteasomal degradation (41, 63). At reduced oxygen concentrations, PHD activity decreases, with almost complete inhibition of activity when oxygen levels fall below 2% (14). The consequent decline in HIF-1α hydroxylation results in protein stabilization and translocation into the nucleus, where it binds HIF-1β and recruits coactivator proteins to form a transcriptional complex at the HIF binding site (5’RCGTG3’) within the hypoxia response element, modulating the transcription of various target genes (26). Increased HIF-1α protein generally correlates with increased transcriptional activity, although HIF-1 transactivation is regulated by hydroxylation of HIF-1α at an asparagine residue (Asp803) within the COOH-terminal transactivation domain via factor inhibiting HIF-1 (FIH-1), which causes a conformational change in the protein and blocks the binding of the transcriptional coactivators CREB binding protein and p300 (18, 42).

Both PHD and FIH-1 hydroxylation activity depend on oxygen availability (Fig. 1), with one atom of dioxygen incorporated in the prolyl or asparagyl residue, respectively, providing a molecular basis for the oxygen-sensing function of these enzymes (30, 53). Analysis of the kinetics of PHD and FIH-1 activity suggests that these enzymes are ideally suited as oxygen sensors, with in vitro derived KM values for oxygen binding of ~100 μM and 70 μM for PHDs and FIH-1, respectively (33). This range of oxygen dependence has been confirmed by measurement of PHD activity in cellular extracts under different oxygen concentrations (6). Thus, based on their affinities for oxygen, PHDs and FIH-1 are well positioned to sense oxygen within the range of physiological tissue concentrations (30).

Later work revealed the existence of paralogs of the HIF-1α subunit. HIF-2α was identified based on its sequence similarity to HIF-1α (16, 87) and is regulated by the same PHD-dependent degradation pathways. HIF-2α also dimerizes with HIF-1β, but, in contrast to HIF-1α, exhibits a restricted, cell-specific expression pattern (53, 63). A third paralog, HIF-3α, has also been identified, but appears to function primarily as a negative regulator of HIF-1 (53).

Early studies revealed the oxygen dependence and kinetics of HIF-1 activation in the lung (95, 99), with in vivo hypoxic induction of HIF-1α mRNA occurring within 30 min (95). Using an isolated, ventilated, perfused ferret lung preparation, where rapid biopsies of hypoxic lung tissue could be readily obtained, significant HIF-1α protein accumulation was observed within 1 h of initiating hypoxic exposure, continued to increase for the duration of the hypoxic challenge, and decreased within 1 min on reoxygenation (99). With more reduced preparations, it became evident that HIF-1α protein levels and DNA binding increased in both endothelial and smooth muscle cells from pulmonary arteries (99). The rapid induction of HIF-1α observed in pulmonary arterial smooth muscle cells (PASMCs) exposed to 1% oxygen is most certainly a direct consequence of substrate-limited inhibition of PHD activity. Moderate levels of hypoxia (4% oxygen) also induce HIF-1α protein, but through a different mechanism, requiring longer duration of hypoxic exposure and involving production of endothelin-1 (ET-1) and downregulation of PHD expression (51).

HIF AND ACUTE PULMONARY RESPONSES TO HYPOXIA

In the systemic circulation, hypoxia causes vasodilation to increase oxygen delivery and meet the metabolic demands of the tissue. Conversely, the pulmonary circulation contracts as alveolar and arterial oxygen content decrease, as initially described in detail by von Euler and Lillestrand in 1946 (90). The exact reason for this phenomenon, termed hypoxic pulmonary vasoconstriction (HPV), remains in debate, although HPV is most commonly believed to be a residual feature from fetal development, where the hypoxic environment maintains a closed pulmonary circulation until birth, or a mechanism to match ventilation and perfusion by diverting blood flow from poorly oxygenated portions of the lung. In either case, decrements in alveolar oxygen tension give rise to an immediate, reversible contraction that varies in intensity by level of hypoxia, sex, and species (84). The exact mechanisms underlying oxygen sensing in the pulmonary vasculature are still under investigation, but are considered to require a coordinated response between the endothelial and smooth muscle cells (1, 84). In PASMCs, hypoxia leads to production of mitochondrial...
reactive oxygen species, calcium release from the sarcoplasmic reticulum, opening of calcium-permeable nonselective cation channels, inhibition of voltage-gated potassium channels, membrane depolarization, and subsequent activation of calcium influx through voltage-gated calcium channels (84). Increased production of vasoconstricting, and/or reduced production of vasodilating, factors by the neighboring endothelial cells (ECs) augment PASMC responses to hypoxia (84), indicating that, while the main contractile mechanics underpinning HPV reside in PASMCs, the endothelium certainly plays a modulatory role in the response.

The fact that the pulmonary circulation contracts almost immediately upon a reduction in oxygen tension would appear to rule out a role for regulation of the acute response at the level of transcription, and thus eliminate the possibility that HIF mediates HPV. However, the finding that individuals with Chuvash polycythemia, a rare genetic condition where germ-line hypomorphic mutations in VHL result in augmented HIF expression under nonhypoxic conditions, exhibited enhanced HPV (79) suggested a possible role for HIF in regulating acute pulmonary vascular responses to hypoxia. The proposal that HIF could regulate HPV finds further support in studies of humans with gain-of-function mutations in HIF-2α who also exhibit enhanced HPV (20). Determining whether the role of HIF lies in modulating targets believed to be directly involved in the contractile response (i.e., voltage-gated potassium or calcium channels) or in circulating factors known to be required for enhancement or priming of the smooth muscle during HPV (i.e., ET-1), remains to be determined.

HIF AND PULMONARY HYPERTENSION

Pulmonary hypertension is defined as a resting mean pulmonary artery pressure ≥ 25 mmHg and results from various etiologies, including chronic hypoxia (CH) associated with several lung diseases (74). Pathophysiology results from sustained active vasoconstriction and pulmonary vascular remodeling, the latter of which is characterized by medial thickening, enhanced muscularity, and migration and proliferation of PASMCs (70, 82). Ultimately, right ventricular hypertrophy becomes a serious complication with the potential for ensuing right ventricular failure. The majority of work addressing the effects of CH on the pulmonary circulation has been performed in adult animals, even though increased pulmonary arterial pressure, remodeling, and contraction also occur in infants exposed to perinatal hypoxia. It is widely recognized that regulation of pulmonary arterial pressure is influenced by postnatal age, and it is unclear which mechanisms identified in the adult translate to the neonate. The focus of this section will be on mechanisms identified in adult animals.

Rodents exposed to CH develop vasoconstriction and vascular remodeling reminiscent of humans with hypoxia-associated pulmonary hypertension and are, therefore, useful models to identify cellular modifications contributing to disease pathogenesis (70, 82). PASMCs isolated from chronically hypoxic rats exhibit elevated intracellular calcium concentration ([Ca²⁺]i) (68), which in turn leads to enhanced cell contraction (73), migration (36), and proliferation (47). Initial reports of downregulated voltage-gated potassium channels (77, 91) and membrane depolarization (83) suggested that voltage-gated Ca²⁺ channels might be responsible for the increased [Ca²⁺]i; however, it is now well established that sustained calcium influx is mediated primarily by upregulation of nonselective cation channels (68), which are composed of canonical transient receptor potential (TRPC) proteins (39, 92). Instead, reduced voltage-gated potassium channel expression, which is HIF dependent (8, 71, 94), may contribute to remodeling by reducing apoptosis (55).

PASMC proliferation is also regulated by Na⁺/H⁺ exchanger (NHE) activity, which extrudes H⁺ ions for Na⁺ ions and controls intracellular pH (pHi) (54). Moreover, NHE form 1 (NHE1) expression and activity, and thus pHi, are elevated in PASMCs isolated from chronically hypoxic animals (58, 69), and NHE inhibitors prevented CH-induced pulmonary vascular remodeling and development of pulmonary hypertension (54). The role of NHE1 in hypoxia-induced pulmonary hypertension was confirmed using NHE1 null mice, which were protected from CH-induced elevations in right ventricular pressure (used as an estimate of pulmonary arterial pressure) and pulmonary vascular remodeling (103), and in vitro silencing of NHE1 by short interfering RNA, which inhibited PASMC proliferation and migration in response to hypoxia (102).

The role of HIF-1 in regulating the development and maintenance of hypoxia-induced pulmonary hypertension has been investigated using transgenic mice. Indistinguishable from their wild-type littermates under normoxia, mice with a loss-of-function mutation in one allele encoding the HIF-1α protein (Hif1a+/-) (28) are partially protected from developing CH-induced PH, exhibiting reduced vascular remodeling and attenuated increases in right ventricular pressure and hypertrophy (100). In Hif1a+/- mice, CH also fails to upregulate expression of TRPCs or NHE1, elevate [Ca²⁺]i, alkalinize pHi, or reduce voltage-gated potassium channel expression or activity (69, 71, 93, 94). Consistent with these findings, gain-of-function studies demonstrated that expressing HIF-1α under nonhypoxic conditions mimicked the effects of hypoxia, resulting in downregulated voltage-gated potassium channel subunits, increased TRPC and NHE1 expression, enhanced NHE activity, and elevated pH (69, 93, 94).

Other HIF target genes may also participate in the development of hypoxic PH. First isolated from cultured porcine aortic ECs in 1988 (97), ET-1 is a HIF target, potent vasoconstrictor, and stimulator of vascular smooth muscle cell proliferation (23). CH increases lung, pulmonary artery, and plasma ET-1 levels, as well as the expression of ET-1 receptors (37, 72). In animal models, ET-1 receptor antagonists partially prevent and/or reverse established CH-induced pulmonary hypertension (23, 72). In addition to being a HIF target, as described earlier in this review, ET-1 also upregulates HIF-1α in PASMCs (38, 51), creating a feedforward mechanism to enhance HIF-1 expression and potentiate the development of pulmonary hypertension (Fig. 2). Consistent with this hypothesis, ET-1 receptor antagonism prevented upregulation of HIF-1α in PASMCs and rats exposed to hypoxia (51). In contrast, ET-1 failed to augment HIF-1α in aortic smooth muscle cells, suggesting a feature not shared by all vascular smooth muscle.

HIF-2α, with predominant lung expression in ECs and epithelial cells, also appears to play a role in the pathogenesis of CH-induced pulmonary hypertension, as mice with heterozygous deficiency for HIF-2α exhibited blunted right ven-
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Fig. 2. Proposed role of HIF-1 in hypoxic pulmonary hypertension. Initially, hypoxia induces the synthesis and release of endothelin-1 (ET-1) from pulmonary endothelial and arterial smooth muscle cells (PASMCs). ET-1 binds to receptors on the PASMCs, increasing Ca2+ sensitivity of the contractile apparatus and upregulating HIF-1α, which induces ET-1 transcription and results in a feedforward mechanism to augment HIF activation. Downstream targets of HIF-1 include voltage-gated K+ channels (including Kv1.5 and Kv2.1), Na+/H+ exchanger isofrom 1 (NHE1), and canonical transient receptor potential (TRPC) proteins (TRPC1 and TRPC6), which coordinate to regulate PASMC contraction, growth/survival, and migration.

**Potential for Therapeutic Targeting of HIF**

Until the last 5 years, pharmacological targeting of HIF was quite difficult. However, in a study published in 2008, Zhang et al. (104) reported the use of a high-throughput screening assay for HIF activity to identify a number of clinically used compounds in the Johns Hopkins Drug Library that inhibited HIF activity. Of the identified compounds, several have now been used in preclinical studies to evaluate the ability to target HIF pharmacologically (35, 60, 96, 98, 104) and to determine the effect of such targeting on hypoxic responses in the lung (2).

Finally, inflammation due to CH is another component likely contributing to the pathogenesis of pulmonary hypertension. In human patients and animal models, inflammation is an early consequence of hypoxic exposure (10, 21, 89), and studies in chronically hypoxic rats revealed that mast cell accumulation, increased inflammatory cell infiltrates, and recruitment of circulating monocytes and progenitor cells preceded and/or contributed to vascular remodeling (10, 50). Following recruitment, these cells can exert paracrine effects on the vasculature via release of vasoactive, pro-proliferative, and chemotactic metabolites (21, 49) and/or stimulate production of collagen and facilitate endothelial-mesenchymal transdifferentiation (81). The proinflammatory environment created in the pulmonary vasculature during CH involves several known HIF target genes, including nuclear factor (NF)-κB (89), stromal cell-derived factor-1 (10), and vascular endothelial growth factor (89). Taken together, these data suggest that, in addition to direct effects in ECs and PASMCs, HIF may also contribute to the development of hypoxia-induced pulmonary hypertension by promoting an inflammatory milieu.
remodeling, and normal PASMC intracellular Ca\(^{2+}\) and pH\(_i\) values (2). Interestingly, digoxin reduced, but did not normalize, right ventricular hypertrophy, perhaps suggesting a direct effect of hypoxia on the right ventricle independent of HIF-1 or the pulmonary vascular effects of hypoxia.

In mice beginning digoxin treatment after hypoxia-induced pulmonary hypertension had already developed, pH\(_i\), and [Ca\(^{2+}\)], were still normalized; however, vascular remodeling was unaltered (2). In rats, the smooth muscle that appears in the small intra-acinar pulmonary vessels during exposure to CH can be reversed with return to normoxic conditions for 6 wk (76), suggesting that de-remodeling is possible if the inciting conditions are removed. Whether the lack of de-remodeling observed with digoxin reflects insufficient time to reverse the changes in muscularity of the small vessels, or, once established, vascular remodeling is maintained via HIF-independent mechanisms, remains to be determined. Right ventricular systolic pressures were also reduced, indicating a slowing of the progression of pulmonary hypertension (2). The ability of digoxin to attenuate further increases in right ventricular pressure in the face of maintained vascular remodeling suggests that vasoconstriction likely plays a more prominent role in maintaining increased pulmonary arterial pressure with longer exposures to CH. This notion is supported by reports showing that these drugs could have exerted their effects independent of HIF inhibition. That HIF target genes were reduced in the lungs of digoxin-treated animals, and acriflavine inhibits HIF activity by a distinct molecular mechanism (35) and does not have other known mechanisms of action that overlap with digoxin, suggests the effects of these drugs were likely due to pharmacological inhibition of HIF; nevertheless, caution should be used when attributing the effects solely to HIF inhibition.

Another method to potentially target the HIF system is via iron supplementation, since PHD activity is iron sensitive (29). Indeed, infusion of iron reduced, whereas iron chelation increased, the elevation in pulmonary arterial pressures in response to both acute (78) and sustained hypoxia (80). As with the drugs discussed above, it is not possible to rule out nonspecific effects of iron supplementation; however, these data are consistent with the proposal that HIF inhibition could be protective during hypoxic pulmonary hypertension.

CONCLUSIONS

The unique responses of the pulmonary vasculature to hypoxia have been widely studied, yet the exact mechanism(s) underlying these responses remains to be fully elucidated. Data from humans and animal models implicate HIF as both a modulator of acute HPV and a mediator of the development of hypoxia-associated pulmonary hypertension. In patients, digoxin has been used in the treatment of right ventricular failure during pulmonary hypertension (35, 45, 57); however, without data supporting a positive effect of digoxin on survival and pulmonary arterial pressure, the use of this drug in patients with pulmonary hypertension remains controversial. Future studies using HIF inhibitors, including digoxin and acriflavine, will be needed to verify the role of HIF in HPV and to determine whether these agents will prove useful in treating pulmonary hypertension.

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DISCLOSURES

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REFERENCES


