THE CAUSES OF MUSCLE FATIGUE are complex, and not totally understood (1, 21). Depending on the circumstance, fatigue may result from disturbances in the central nervous system (26) and/or peripheral factors within the skeletal muscles (1–3, 21–23, 28, 29, 58). By definition, muscle fatigue is characterized by a loss of muscle power that results from a decline in force and velocity (16, 21). To distinguish fatigue from muscle weakness or damage, it is important to note that the loss of power with fatigue is reversible by rest (21). The etiology of muscle fatigue is an important question, as it can lead to serious limitations in muscle and whole body performance (21, 23), and in clinical situations to respiratory failure and death (24). While the exact causes of muscle fatigue and the relative importance of particular factors remains controversial (1, 21), depending on the circumstance, fatigue may result from disturbances in the central nervous system (26) and/or peripheral factors within the skeletal muscles (1–3, 21–23, 28, 29, 58). By definition, muscle fatigue is characterized by a loss of muscle power that results from a decline in force and velocity (16, 21). To distinguish fatigue from muscle weakness or damage, it is important to note that the loss of power with fatigue is reversible by rest (21). The etiology of muscle fatigue is an important question, as it can lead to serious limitations in muscle and whole body performance (21, 23), and in clinical situations to respiratory failure and death (34). While the exact causes of muscle fatigue and the relative importance of particular factors remains controversial (1, 21), it is clear that an individual’s state of fitness, dietary status, fiber type composition, and intensity and duration of the exercise all affect the process. For example, the factors causing fatigue during high-intensity exercise are distinctively different from those precipitating fatigue during prolonged activity (21). The problem is further complicated as muscle fatigue not only results from multiple factors acting at various sites but in many cases from synergistic actions of two or more agents. The purpose of this minireview is to describe the functional changes that characterize fatigue and then discuss the role of altered cross-bridge events in the fatigue process. Other factors such as central fatigue, and events affecting excitation-contraction coupling or energy supply, will not be considered.

THE CROSS-BRIDGE CYCLE

To understand fatigue mediated by alterations in cross-bridge events, it is important to review the steps involved in cross-bridge cycling and force generation. Although the molecular details of the cross-bridge cycle are not yet fully understood, Fig. 1 reproduced from Geeves et al. (27) shows a minimal mechanochemical scheme for the actomyosin cross-bridge cycle. The myosin and actomyosin ATPase steps are shown in the top and bottom line of Fig. 1A, respectively, with the predominant pathway for the actomyosin ATPase shaded. The scheme shown in Fig. 1B begins with the rigor complex (A•M, state a). As Geeves et al. (27) point out, each step shown has a series of substeps that involve protein conformational changes that alter the affinity of the myosin binding to actin, P, or ADP (50). When myosin initially binds to actin, the actomyosin is in a weakly bound low-force state (state c). With the subsequent release of P, (step d), the cross bridge transforms into strongly bound high-force state and goes through the power stroke (state e). ADP is then released (state f), returning the cross bridge to the rigor complex. The strongly bound, high-force states (states d, e, f, and a) are thought to be the dominant form during a maximal isometric contraction,
Fig. 1. Actomyosin ATPase cycle. A: minimal description of the myosin (M) and actomyosin (A·M) ATPase as defined in solution. Top line represents the myosin ATPase with the following events: ATP (T) binding, and ATP hydrolysis, followed by P_i release and then ADP (D) release. The equivalent steps for actomyosin are shown in the bottom line. Vertical arrows indicate the actin (A) association and dissociation from each myosin complex. In every case, the events shown can be broken down into a series of substeps involving one or more identifiable protein conformational changes. States outlined and with a shaded background represent the predominant pathway for the actomyosin ATPase. B: a minimal mechanoscheme for the actomyosin cross-bridge cycle. Starting from the rigor complex, A·M (state a), ATP binds to rapidly dissociate the complex, and the lever arm is reprimed to the pre-power stroke position (state b). This is followed by hydrolysis. The preceding 3 states have been well defined by crystallography, electron microscopy, and solution kinetics. The exact sequence of biochemical, structural, and mechanical events is more speculative. The M·D·P_i complex rebinds to actin, initially weakly (state c) and then strongly (state d). Binding to actin induces the dissociation of P_i and the power stroke (state e). The completion of the tail swing (state f) is followed by ADP release to return to the rigorlike complex (state a); in some myosins (e.g., smooth muscle myosin II; myo 1b, or myosin-V), ADP dissociation is associated with a further displacement of the lever arm. Actin monomers are shown as golden spheres. The motor domain is colored metallic gray for the free form, purple for the weakly bound form, and violet for the strongly bound form. The converter is shown in blue and the lever arm in orange. [Reproduced from Geeves et al. (27) with copyright permission of Birkhäuser Verlag, Basel, Switzerland.] whereas during isotonic shortening skeletal muscle myosin spends only 5% of the cycle time in strongly bound states (50). The rate of transition from the weakly bound low-force state to the strongly bound high-force state (difference between the forward and backward rate constants of the state c-to-state d transition in Fig. 1) is thought to limit the peak rate of force development (+dP/dt). In single cells, this parameter is determined by shortening and reextending a fully activated fiber and measuring the rate constant of tension redevelopment (k_tr). The k_tr is sevenfold higher in fast compared with slow fibers and shows Ca^{2+} sensitivity with lower values observed in suboptimal Ca^{2+} conditions (40). In contrast, the maximal unloaded shortening velocity (V_0) is highly correlated with the actomyosin ATP hydrolysis rate, which in turn appears limited by the ADP dissociation step (state f-to-state a transition in Fig. 1). This kinetic scheme has proven useful in assessing the cellular mechanisms of muscle fatigue for those factors acting at the cross bridge (21, 23).

ALTERATIONS IN MUSCLE MECHANICS WITH FATIGUE

The functional correlates of fatigue observed in both animals and humans during prolonged endurance and high-intensity, short-duration exercise include a decline in twitch and tetanic force, maximal shortening velocity, and peak power (7, 17, 21, 23). While muscles do not contract in vivo with single Twitches, the evaluation of isometric twitches before and after fatigue has proven useful in identifying the cellular sites of fatigue (21). Features of a postfatigued twitch are reduced twitch force (P_0), prolonged contraction and half-relaxation times (CT and 1/2RT), and reductions in the peak rate of tension development (+dP/dt) and decline (−dP/dt). The low P_0 and prolonged CT and 1/2RT are known to reflect a reduction in the amplitude and an increase in the duration of the intracellular Ca^{2+} transient, respectively, and thus do not necessarily reflect any direct affect on the cross bridge (1, 2).

The reduced peak force (P_0) with fatigue can be explained by a decline in the force per cross bridge and/or the number of cross bridges in the high-force states (states d, e, f, and a in Fig. 1). The mechanism can involve a decline in the amplitude of the Ca^{2+} transient and agents that directly inhibit the cross bridges. Allen et al. (1) summarized the mechanisms involved in the reduction of isometric force with fatigue and suggested that the initial decline in force occurred as a direct result of an inhibition of the force-generating steps of the cross-bridge cycle. Simultaneously, there was a right shift of the force-Ca^{2+} relationship, but this had no consequence until later in fatigue when the amplitude of the intracellular Ca^{2+} transient decreased.

With contraction, the forward rate constant for the weakly bound, low-force state c (Fig. 1) to the strongly bound, high-force state d of the cross bridge (Fig. 1) is accelerated by both cytoplasmic Ca^{2+} and the number of strongly bound cross bridges, both of which decrease with fatigue. In high-intensity exercise, the decline in +dP/dt with fatigue could also be mediated by an increase in H^+ and P_i. The observation that +dP/dt divided by P_0 ([+dP/dt]/P_0) was not altered by fatigue suggests that the reduced +dP/dt resulted primarily from the decline in P_0, which in turn was caused by the fatigue-induced reduction in the number and force of the strongly bound, high-force cross bridges (21, 36). The latter is dependent on the
concentration of cytoplasmic Ca$^{2+}$, pH, and P$_i$. In contrast, $-dP/dt$ declines considerably more than force, and more in fast than slow fibers. This suggests a slower dissociation of actin from myosin in fatigued muscle cells, which could reflect a direct effect on the cross-bridge detachment rate and/or a reduced rate of Ca$^{2+}$ reuptake by the SR pumps (56, 57). The rate of Ca$^{2+}$ dissociation from troponin is thought to be too fast to be rate limiting (1). Relaxation from a tetanus generally occurs in three phases, where force initially shows no change following the final action potential (phase 1); then exhibits a slow, linear drop (phase 2); and finally a rapid exponential decline (phase 3). With fatigue, each of these phases is slowed, and the distinction between them is less clear (1, 60). During phase 2, the sarcomere length remains constant, and the force decline is thought to reflect the rate of cross-bridge detachment. The time course of the decline in force is delayed compared with the predicted force determined from the force-pCa relationship. It has been hypothesized that the increased difference between the measured and predicted force in fatigued muscle fibers is due to a slowed relaxation caused in part by a reduced cross-bridge dissociation rate (1, 56, 57).

While peak force has been shown to decline before and to a greater extent than velocity (15), both are known to contribute to the loss of power with fatigue. Peak power shows a greater decline with fatigue in fast muscles because of a greater reduction in both force and velocity (23). The decline in velocity at peak power in fast muscles is in part due to a fatigue-induced increase in the curvature (i.e., lower a/P$_0$ ratio) of the force-velocity relationship (23, 31). This is particularly important as peak power is elicited at approximately 20–30% of peak force and maximal velocity, where the force-velocity relationship has the greatest curvature (23, 31).

While force always declines first, the reduction in maximal velocity determined from the force-velocity relationship ($V_{max}$) may ultimately drop by an amount equal to that observed for force (1). However, muscles contracting in vivo never experience zero loads; thus it is important to assess whether the curvature as reflected by the a/P$_0$ ratio changes with fatigue. In fast muscles, the a/P$_0$ ratio is three- to fourfold higher compared with slow muscle. Consequently, individuals with a high percentage of fast-twitch fibers can generate higher velocity and power at a given relative load than those with predominantly slow-twitch fibers. There are little data assessing the effect of fatigue on the a/P$_0$ ratio (23). Jones et al. (31) produced fatigue in the fast-twitch human adductor pollicis muscle and observed the a/P$_0$ ratio to decline with a similar time course to the loss of peak power. The authors concluded that the loss of force and the reduced a/P$_0$ ratio contributed equally to the decline in peak power. To my knowledge, there is no comparable information on predominately slow-twitch muscles. Since the extremes of the force-velocity relationship ($V_{max}$ and P$_0$) in response to a given contractile paradigm are more resistant to change in slow compared with fast muscles, one might expect a similar difference in the stability of the a/P$_0$ ratio (1, 21, 23).

MUSCLE FATIGUE AND THE CROSS-BRIDGE CYCLE

Inhibition of the high-force states by H$^+$ and P$_i$. Intense exercise induces high rates of ATP hydrolysis and glycolysis, and corresponding increases in cell H$^+$, P$_i$, and ADP. The first two are known to directly reduce the force output of the high-force states, while the former by itself would increase force but slow velocity (8, 9, 12, 43). Intracelluar pH can reach levels as low as 6.2 (from a resting value of 7.0), while P$_i$ may increase to 30 mM (7, 10, 61). The extent of the change in these ions is directly related to the intensity of the work and fiber type, with the fast glycolytic fibers (type IIx and IIb) showing the lowest pH and highest P$_i$ values (21). Metzger and Moss (37) found fast-twitch fibers from the predominantly type IIb and IIx superficial region of the vastus lateralis of the rat to be more sensitive than slow soleus fibers to the depressive effects of low pH. This difference was not observed when slow type I and fast type IIa fibers were compared (30). The observation that the depressive effects of low pH on peak force are still apparent in the presence of saturating levels of free Ca$^{2+}$ indicates that the mechanism is not simply the result of H$^+$ inhibition of Ca$^{2+}$ binding to troponin (37). The fact that rigor tension (state a in Fig. 1) was reduced at pH 6.2 compared with 7.0 suggests a direct effect of H$^+$ on the actomyosin cross bridge (19). The effect could involve a decline in the number and/or the force per bridge of the high-force strongly bound cross bridges (Fig. 1). At 15°C, a drop in fiber pH has been shown to reduce the force per cross bridge in both fast- and slow-twitch fibers, while the number of strongly bound (high force) cross bridges was reduced only in fast fibers (38, 39). The latter attributed to a H$^+$-mediated inhibition of the forward rate constant for the transition between the weakly and strongly bound cross-bridge states. The observation that stiffness (a marker of the number of strongly bound cross bridges) did not decline in slow fibers, and declined less than force in fast fibers, suggests that the primary means by which protons inhibit force is a reduction in the force per bridge.

Similar to H$^+$, an increased intracellular P$_i$ is correlated with fatigue in contracting muscles (7, 10, 61) and is known to depress force in maximally activated skinned fibers (12–14, 25, 33, 43, 46). Also, like H$^+$, it is thought to act by inhibiting the transition from the weakly bound (state c; Fig. 1) to the strongly bound (state d; Fig. 1) cross bridge, and/or a reduction in the force of the strongly bound states (13, 44). It has been suggested that high P$_i$ reduces force and increases $k_{rx}$ by accelerating the reverse rate constant of the low- to high-force state transition (44). The observation that fiber stiffness is less sensitive to P$_i$ than force suggests that either a part of the force depression is due to less force per strongly bound bridge or that an additional intermediate low-force, strongly bound state exists. This state of the actomyosin would contain both P$_i$ and ADP and would increase in the presence of high P$_i$ (13, 50).

Effect of temperature on the inhibition of force. Up until the mid-1990s, experiments assessing the effect of H$^+$ and P$_i$ on fiber force were performed at temperatures below 25°C (32, 33, 37–40, 43, 46). Recently, a number of labs, including mine (9, 12, 13, 45, 59), have shown the deleterious effects of high H$^+$ and P$_i$ on the P$_0$ of maximally activated fibers to be reduced at 30°C compared with lower temperatures. The reduced effect of these ions is thought to result from the forward rate constant of the force-generating step being more temperature sensitive than the reverse rate constant (62). Zhao and Kawai (62) used sinusoidal analysis to demonstrate that the forward rate constant was greatly accelerated as temperature increased. The observation that P$_0$ increased with temperature is consistent
with the acceleration of the forward rate constant, and the production of more bridges in the high-force state. In our experiments, for both slow and fast fibers, the inhibiting effect of pH 6.2 on fiber $P_0$ was reduced from $\sim 30$ to 11% with a temperature increase from 15 to 30°C (30). These results were consistent with the data of Pate et al. (45) and Westerblad et al. (59), who found low pH to decrease $P_0$ by 18% and 10%, respectively, at temperatures between 30 and 32°C. The effect of $P_i$ on $P_0$ was also tempered by increasing temperature but, unlike $H^+$, showed a fiber type dependency. At 15°C, 30 mM $P_i$ depressed fiber $P_0$ by $\sim 50\%$ in fast and slow fibers, while at 30°C the decline was only 5% in fast fibers compared with 19% in slow fibers (12). The mechanism of this difference is not understood but could be due to fiber type-dependent effects of temperature on the forward rate constant of the low-to-high-force transition (62). The lack of a fiber type difference in response to low pH (at least between fast type Ila and slow type I fibers) suggests that the mechanism by which $P_i$ is depressed is fundamentally different for high $P_i$ and low pH. If these ions act at different sites in the cross-bridge cycle, their effects should be additive. Using the skinned fiber, Nosek et al. (43) showed this to be the case as 30 mM $P_i$ and pH 6.2 depressed force considerably more than pH 6.2 and zero added $P_i$.

Due to problems with fiber stability, studies using skinned fibers are limited to temperatures at or below 30°C. Intramuscular temperature is known to vary with environmental temperature (48) and exercise (20) and thus may vary between environmental and core body temperature. It is difficult to predict the effect on high $H^+$ and $P_i$ at the high limit of muscle temperatures (37°-39° C). However, the observation that $P_0$ is unaffected and $V_0$ only moderately affected by temperature increases from 30°C to 35°C (47) suggests that the inhibiting effects of $H^+$ and $P_i$ at high muscle temperatures might be relatively the same as those measured at 30°C.

**Inhibition of fiber velocity.** An increase in myoplasmic $H^+$ concentration has been shown to depress the maximal shortening velocity as determined by the slack test ($V_o$) in both living and skinned fibers (15, 37, 52). This effect has been reported to be greater in fast- compared with slow-twist fibers (37). Recently, we observed this fiber type difference to be reduced at high compared with low temperature. At 15°C, we observed pH 6.2 to inhibit $V_0$ by 9 and 27% in slow type I and fast type Ila fibers, respectively, while the inhibition was 25 and 32%, respectively, at 30°C (30). Importantly, the $H^+$ inhibition of the slow fiber $V_0$ was increased at the higher (near in vivo) temperature. Since $V_0$ is thought to be limited by the rate of ADP dissociation from the myosin head (state $f$-to-state $a$ transition, Fig. 1), it is apparent that low pH inhibits this transition as well as the weakly to strongly bound cross-bridge transition (state $c$ to state $d$, Fig. 1). When the pH effects on maximal velocity were determined from the force-velocity relationship ($V_{max}$), the results were qualitatively the same, but the degree of inhibition was less than observed for $V_0$ (30). The mechanism by which low pH inhibits velocity is unknown. It may directly inhibit the rate-limiting step (transition from state $f$ to state $a$, Fig. 1) and/or cause a decrease in the filament lattice spacing and corresponding increase in internal drag (30, 53).

Controversy exists as to the relative importance of low pH in inhibiting fiber velocity (30, 59). Westerblad et al. (59) studying living fast fibers at 32°C found no significant effect of pH 6.7 on $V_0$, and Bruton et al. (6) concluded that acidosis to pH 6.6 (produced by 30% CO$_2$ exposure) did not accelerate the development of fatigue during tetanic stimulation of fast fibers. In contrast, Edman and Mattiazzi (15) observed a significant decline in $V_0$ and $P_0$ following the acidification of frog fibers by $\sim 1$ pH unit. Collectively, these results suggest that the inhibiting effects of protons on $V_0$ and $P_0$ do not occur until pH drops below 6.7. Intense exercise has been shown to reduce intracellular muscle pH to as low as 6.2 (61); however, in many studies the extent of acidosis is less (see Table 1 of Ref. 21). Clearly, the contribution of $H^+$ to fatigue will depend on the extent of acidosis, muscle temperature, and the amplitude of the intracellular Ca$^{2+}$ transient. The latter is important as $H^+$ is known to depress myofibrillar Ca$^{2+}$ sensitivity.

High $P_i$ (up to 30 mM) has no effect on slow type I or fast type II fiber $V_{max}$ at cold or near in vivo temperatures (8, 12). Recently, Franks-Skiba et al. (24) reported that at 30°C and in the presence of 0.5 mM of the phosphate analog vanadate, myosin light chain phosphorylation inhibited fast fiber $V_{max}$. No effect of vanadate was observed in dephosphorylated myosin light chain conditions. The authors suggest that since the vanadate and $P_i$ occupy the same cross-bridge states, a $P_i$ build-up coupled to an increased myosin light chain phosphorylation could contribute to the decline in fiber velocity in fatigued muscles. The mechanism is not clear but might be mediated by a slower release of ADP or to an increased drag caused by weakly bound cross bridges (24).

**Force-velocity relationship and peak power.** The extent of muscle fatigue is perhaps best related to changes in peak power, as body movement is dependent on the capacity to generate power. Since power is a function of both force and velocity, it is considerably higher in fast fibers and greatly affected by temperature (12, 30). Peak power is elicited at contraction velocities and loads between 20 and 40% of $V_{max}$ and $P_0$. Given their effects on force and velocity, it is not surprising that both low pH and high $P_i$ depress peak power (Fig. 2). For $P_i$, the inhibition of power is reduced but still significant at 30°C compared with 15°C (12). At the higher temperature, peak power was reduced by 26 and 18% of the control condition (zero added $P_i$) for slow and fast fibers, respectively (Fig. 2, B and D). High temperature not only increases peak force, $V_0$, and peak power, but the $a/P_0$ ratio allowing fibers to generate more force at a given velocity. At 30°C, high $P_i$ reduces peak power by depressing force and reducing the $a/P_0$ ratio such that less force is generated at a given velocity (12, 31). The latter is quite substantial. At 30°C, high $P_i$ depressed the $a/P_0$ ratio by 38 and 30% in fast and slow fibers, respectively (12). The observation that $P_i$ had no effect on the $a/P_0$ ratio at 15°C suggests that the mechanism cannot by due to an increased internal drag caused by a $P_i$-induced increase in the number of weakly bound low-force cross bridges (state $c$, Fig. 1) as that effect is greater at the lower temperature. The most likely explanation is that $P_i$ inhibits the cross-bridge detachment rate. In support of this hypothesis, Westerblad and Allen (56, 57) observed a marked slowing of the peak relaxation rate ($-dP/dt$) with fatigue of the mouse flexor brevis muscle, which they attributed to an altered cross-bridge kinetics. The observation that high $P_i$ depresses the $a/P_0$ ratio but not $V_{max}$ suggests that the step limiting velocity in the cross-bridge cycle (Fig. 1) may be different in loaded vs. unloaded contractions.
In contrast to Pi, low pH inhibits the peak power of the slow fiber to a greater extent at 30°C than at colder temperatures (30). This effect is attributed to a greater depression of velocity at the higher temperature. Unlike colder temperatures where fast fibers are more sensitive to low pH, at near in vivo temperatures, slow fibers show a greater pH-dependent depression of peak power (compare Fig. 2, A and C). These data on fiber type susceptibility should be interpreted with caution as the skinned fiber preparation may not reflect the conditions of contracting living fibers. If as suggested by Franks-Skiba et al. (24), myosin light chain phosphorylation exacerbates the inhibitory actions of H⁺ and/or Pi, the effect might be greater in fast fibers known to have higher myosin light chain kinase activities.

Low pH depressed the $a/P_0$ ratio in slow but not fast fibers. Thus the fatigue-induced increase in the curvature of the force-velocity relationship (reduced $a/P_0$ ratio) observed in fast muscles studied in situ or in vivo might be induced by an increased Pi but does not appear to be caused by low pH (23, 30, 31).

**MUSCLE FATIGUE AND THE FORCE-pCa RELATIONSHIP**

Both H⁺ and Pi not only directly inhibit cross-bridge force but also shift the force-pCa relationship to the right such that higher free Ca²⁺ is required to reach a given tension. As long as the amplitude of the Ca²⁺ transient remains high (pCa > 5.5), peak force will be unaltered by the reduced Ca²⁺ sensitivity (Fig. 3). However, late in fatigue, Allen et al. (2) have shown a precipitous drop in the amplitude of the Ca²⁺ transient. Under these conditions (pCa < 6.0), the reduced Ca²⁺ sensitivity induced by Pi and H⁺ would be expected to make a significant contribution to the decline in force (Fig. 3). For low pH, this effect is mediated in part by competitive inhibition of Ca²⁺ binding to troponin-C (55). In addition, the decline in the number of strongly bound, high-force cross bridges (caused by both H⁺ and Pi) reduces the thick filament-mediated cooperativity between regulatory thin filament sites, and this is thought to contribute to the right shift in the force relationship. Additionally, at less than maximal Ca²⁺ concentrations, the pH and Pi-induced right shift in the force-pCa relationship would

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**Fig. 2.** Composite force-power curves for soleus type I fibers determined at pH 7.0 and 6.2 and zero added Pi (A), and at zero and 30 mM added Pi, and pH 7.0 (B). Composite force-power curves for gastrocnemius type II fibers determined at pH 7.0 and 6.2 and zero added Pi (C), and at zero and 30 mM added Pi, and pH 7.0 (D). W/l = watts/liter. Adapted from data plots published in Debold et al. (12) and Knuth et al. (30); all data were obtained at 30°C.
also contribute to the decline in $k_{ir}$ (and presumably $+dP/dt$) as the forward rate constant of state c-to-state d transition (Fig. 1) would be slowed by the reduction in the number of strong-binding, high-force cross bridges (39).

The $Ca^{2+}$ sensitivity of force development is fiber type dependent with fast fibers activating at a higher free $Ca^{2+}$ concentration but with a greater degree of cooperative binding. A rise in muscle temperature has been shown to increase $Ca^{2+}$ sensitivity (shift the force-pCa relationship to the left) in both slow and fast fibers (13, 49, 51). An important finding from the perspective of muscle fatigue is the recent observation by Debold et al. (13) that high $P_i$ depresses $Ca^{2+}$ sensitivity more at 30°C compared with 15°C. Figure 3 shows the extent of the right shift in the force-pCa relationship caused by 30 mM $P_i$ at 30°C for both slow type I and fast type II fibers (13). Although not yet studied, it is likely that the depressive effect of $H^+$ on $Ca^{2+}$ sensitivity is also temperature dependent. These results suggest that when SR $Ca^{2+}$ release is compromised (as it is in high-intensity muscle contraction), the deleterious effects of high $P_i$ and $H^+$ on muscle force are likely to be more important than previously thought. The steeper force-pCa relationship in fast fibers contributes to the greater susceptibility of the fast fiber type to fatigue. For example, in the Debold et al. (13) study, 30 mM $P_i$ resulted in a twofold greater drop in force in fast compared with slow fibers when pCa went from 5.0 to 6.0 (Fig. 3).

**REACTIVE OXYGEN SPECIES AND CROSS-BRIDGE FUNCTION**

The production of reactive oxygen species (ROS) is known to increase with exercise, and a number of studies have suggested that increases in ROS contribute to the development of fatigue (4, 5, 11 41, 42, 54). Moopanar and Allen (41) found the contribution of ROS to fatigue to be considerably greater at 37° compared with 22° C. Edwards et al. (18) recently confirmed the temperature dependence of ROS production. They found the rate of superoxide ($O_2^{-}$) production to show little increase between 22°C and 32°C, while it increased fivefold when temperature increased from 22°C to 37°C. Moopanar and Allen (41) observed fatigue elicited by repeated isometric tetani of the flexor digitorum brevis in vitro to be accelerated at body temperature compared with room temperature. Since the effect was blocked by the ROS scavenger Tiron (5 mM), the authors concluded that the fatigue was caused by ROS. The increased rate of fatigue at the higher temperature was not associated with any change in the amplitude of the $Ca^{2+}$ transient or differences in the extent of decline in maximal $Ca^{2+}$-activated force. Rather, the ROS-mediated fatigue was attributed to a depression in myofibrillar $Ca^{2+}$ sensitivity. ROS production at 37°C without fatigue had no effect on $Ca^{2+}$ sensitivity (18, 42). The exact ROS responsible for the reduced $Ca^{2+}$ sensitivity is unknown. The observation that the superoxide dismutase mimetic Tempol prevented the ROS-induced fatigue eliminates hydrogen peroxide as a causative agent as it is a product of the Tempol reaction with $O_2^{-}$ (18). The site of oxidative damage of the myofibrillar proteins has not been elucidated. Logical candidates are troponin-C and -I as alterations in either could yield a reduced myofibrillar response to $Ca^{2+}$ (5, 55). Other possible targets of oxidative damage are tropomyosin, actin, and myosin (42). An important unanswered question is whether oxidative damage of myofibrillar proteins, shown to occur with stimulation of in vitro preparations, actually occurs with fatigue during in vivo exercise (42).

**SUMMARY AND FUTURE RESEARCH DIRECTIONS**

Force, velocity, and power are ultimately determined by the molecular factors controlling the number and force of the strongly bound cross bridges, and the rate of cross-bridge cycling (Fig. 1). With high-intensity muscle contraction, the force per strongly bound, high-force bridge is reduced by both $P_i$ and $H^+$. In the case of fast fibers, the force reduction is further exacerbated by a decline in the number of strongly bound bridges (Fig. 1, states d, e, f, and a). Elevated $H^+$ ($pH < 6.7$) has the added effect of inhibiting velocity, presumably by slowing the rate of ADP release (Fig. 1 state f-to-state a transition). With fatigue, the decline in peak power is due to inhibition of both force and velocity, and at least in fast fibers to a depressed $a/P_o$ ratio. The latter is thought to be caused by $P_i$ inhibition of cross-bridge detachment. Equally important in
the fatigue process is that both $P_i$ and $H^+$ reduce myofibrillar Ca\(^{2+}\) sensitivity, which becomes important late in fatigue when the amplitude of the Ca\(^{2+}\) transient is depressed. The reduction in myofibrillar Ca\(^{2+}\) sensitivity is greater at 30°C compared with 22°C, which suggests that the deleterious effects of high $P_i$ and $H^+$ may be greater than originally thought from experiments conducted at cold temperatures (10–22°C).

Many of the studies reviewed were conducted using the chemically skinned fiber preparation. A major advantage of this technique is the ability to determine how known quantities of a particular compound ($P_i$, $H^+$, ADP, etc.) either individually or collectively alter fiber force, velocity, and power. Importantly, the effects of a particular fatigue agent can be assessed independently of changes in intracellular Ca\(^{2+}\). However, the chemically skinned fiber has limitations in that reliable studies cannot be performed at temperatures >30°C, soluble proteins such as parvalbumin may diffuse out of the fiber, and the phosphorylation state of proteins (such as myosin light-chain 2) may not reflect that existing in living fibers. For these reasons, future studies need to be performed on both skinned (chemically and mechanically peeled) and living fibers.

While it is clear that alterations in myofibrillar events contribute to fatigue (particularly fatigue resulting from high-intensity exercise), the relative importance of increases in $P_i$ and $H^+$, and the combined effects of high $P_i$, $H^+$, and ADP, on cross-bridge properties and the fatigue of slow and fast fiber types is yet to be determined. Future studies are needed to establish the mechanisms by which these compounds inhibit force and power, and how $P_i$ depresses the $a/P_{0}$ ratio. Since the effects of fatigue agents acting at the level of the myofibrillar proteins are clearly temperature dependent, it will be important to conduct studies at near in vivo temperatures (≥30°C). Finally, while ROS have been shown to depress myofibrillar Ca\(^{2+}\) sensitivity in muscles contracting in vitro, future studies are needed to determine whether this effect occurs during in situ or in vivo muscle contraction; and if it does, the mechanism and site of action of the ROS needs to be established.

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