ACE ID genotype affects blood creatine kinase response to eccentric exercise

Chen Yamin,1 Offer Amir,2 Moran Sagiv,1 Eric Attias,1 Yoav Meckel,1 Nir Eynon,1 Michael Sagiv,1 and Ruthie E. Amir1

1Department of Genetics and Molecular Biology, the Zinman College of Physical Education and Sport Sciences at the Wingate Institute, Netanya; and 2Department of Cardiology, Lady Davis Carmel Medical Center, Haifa, Israel

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Yamin C, Amir O, Sagiv M, Attias E, Meckel Y, Eynon N, Sagiv M, Amir RE. ACE ID genotype affects blood creatine kinase response to eccentric exercise. J Appl Physiol 103: 2057–2061, 2007. First published September 20, 2007; doi:10.1152/japplphysiol.00867.2007.—Unaccustomed exercise may cause muscle breakdown with marked increase in serum creatine kinase (CK) activity. The skeletal muscle renin-angiotensin system (RAS) plays an important role in exercise metabolism and tissue injury. A functional insertion (I)/deletion (D) polymorphism in the angiotensin I-converting enzyme (ACE) gene (rs4646994) has been associated with ACE activity. We hypothesized that ACE ID genotype may contribute to the wide variability in individuals’ CK response to a given exercise. Young individuals performed maximal eccentric contractions of the elbow flexor muscles. Pre- and postexercise CK activity was determined. ACE genotype was significantly associated with postexercise CK increase and peak CK activity. Individuals harboring one or more of the I allele had a greater increase and higher peak CK values than individuals with the DD genotype. This response was dose-dependent (mean ± SE U/L: II, 8,882 ± 2,362; ID, 4,454 ± 1,105; DD, 2,937 ± 753, ANOVA, P = 0.02; P = 0.009 for linear trend). Multivariate stepwise regression analysis, which included age, sex, body mass index, and genotype subtypes, revealed that ACE genotype was the most powerful independent determinant of peak CK activity (adjusted odds ratio 1.3, 95% confidence interval 1.03–1.64, P = 0.02). In conclusion, we indicate a positive association of the ACE ID genotype with CK response to strenuous exercise. We suggest that the II genotype imposes increased risk for developing muscle damage, whereas the DD genotype may have protective effects. These findings support the role of local RAS in the regulation of exertional muscle injury.

Address for reprint requests and other correspondence: R. Amir, Director, Dept. of Genetics and Molecular Biology, Zinman College of Physical Education and Sport Sciences at the Wingate Institute, Netanya 42902, Israel (e-mail: ruthieam@012.net.il).

There is growing evidence for a genetic contribution to the phenotypic responses of exertional muscle damage (8, 13).

The renin-angiotensin system (RAS) plays an important role in human body fluid homeostasis and left ventricular remodeling. The angiotensin-converting enzyme (ACE) is a key component in the RAS, generating the vasoconstrictor angiotensin (ANG) II and degrading vasodilator kinins (11). ACE is widely expressed in human tissues, including skeletal muscle, and may play a metabolic role during exercise (20). ANG II has known effects on metabolism (6) and is a recognized growth factor necessary for the hypertrophy of skeletal muscle in response to mechanical load (16). Most of its known physiological and pathophysiological activities are mediated through the ANG II type 1 receptor (AT1R), which is also the only ANG II receptor present in human skeletal muscle (20). A functional polymorphism of the human ACE gene has been identified in which the presence (insertion: I allele), rather than the absence (deletion: D allele), of a 287-bp Alu repeat element in intron 16 (rs4646994) is associated with lower enzyme activity in both serum and tissue (12, 39), resulting in greater production of ANG II and aldosterone and a decreased half-life of bradykinin (4, 52).

ACE ID polymorphism has been extensively studied in relation to human physical performance. Several reports from European and US Caucasian populations suggested the association of the I and the D alleles with endurance (1, 15, 30–32, 42) and sprint performance (31, 32, 53), respectively. However, some studies in which elite athletes were drawn from diverse sporting disciplines, requiring mixed skills, have failed to demonstrate any association with the ACE genotype (37, 46). Of note, in a different ethnic population of Israeli Caucasians, the ACE DD genotype has been associated with elite endurance athletes (2).

Because there is considerable RAS activity in skeletal muscle that is involved in the regulation of muscle metabolism, vascular tone, and injury responses, we hypothesized that the ACE ID genotype may contribute to the development of exertional rhabdomyolysis and therefore may affect CK response to strenuous exercise. To test our hypotheses, in the present study, we used repetitive eccentric contractions as an exercise model for inducing exertional rhabdomyolysis in healthy young individuals. We investigated the association of ACE ID alleles with CK response to exercise.

MATERIALS AND METHODS

Subjects. Seventy healthy physical education students (42 men and 28 women; aged 22–32 yr) volunteered for the study. Participants

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were physically active, but none were well trained or competitive athletes. All were healthy nonsmokers and were not receiving any medical treatment. Exclusion criteria included an occupation that required heavy weight lifting, participating in a resistance training program in the previous 6 mo, baseline blood values outside of the normal range (men: 24–195 U/l; women: 24–170 U/l), known muscle disorders, and any existing myopathy. Participants were all Israeli Caucasians, with an equivalent ratio of Ashkenazi and Non-Ashkenazi descent.

The study was approved by the Institution Review Board (Helsinki Committee) of the “Hillel-Yafe” Medical Center, and all participants gave written, informed consent before inclusion in the study and the start of any study-related procedures.

Eccentric exercise protocol. Subjects performed one set of 50 maximal eccentric contractions of the elbow flexor muscles of their nondominant arm using the BIDOEX dynamometer (BIDOEX System 3). Subjects were seated with the arm supported and were stabilized at the wrist and the chest. Starting with the elbow flexed to 90° and opening at an angle of 170°, each subject performed 50 maximal eccentric movements of the elbow flexors at 120°/s (each contraction lasted 3 s). During each movement, subjects were verbally encouraged to produce a maximal effort to resist the ability of the dynamometer to extend the elbow. Subjects were given a 10-s rest between each contraction, during which time the dynamometer arm returned passively to the starting position.

Subjects were instructed to drink water before the exercise session and encouraged to maintain hydration and to monitor their urine color throughout the study. They were instructed to call the 24-h study phone to report whether their urine changed from clear or yellow to a brownish color (no subject experienced darkened urine). Subjects were instructed not to participate in any strenuous physical activity until their CK values had returned to near normal.

CK analysis. Blood samples were drawn from an antecubital vein by venipuncture for genotypic analysis and to determine whole blood CK activity pre- and postexercise. Subjects were followed-up and instructed not to participate in any strenuous physical activity throughout the study. They were instructed to call the 24-h study line if their urine color changed from clear or yellow to brownish color.

Genotyping for ACE ID polymorphism. Genomic DNA was extracted from peripheral blood leukocytes using a standard protocol (40). We used a PCR-based method for genotyping the ACE ID polymorphism, as previously described (26). Briefly, this method yields a PCR fragment of 319 bp and 597 bp in the presence of the D and the I alleles, respectively. PCR fragments were amplified from ∼20 ng of each DNA sample used as a template in 20-μl polymerase chain reaction (PCR) containing 0.2 U Taq polymerase, 1× reaction buffer, 0.2 mM concentration of each deoxynucleotide triphosphate, and 10 pmol of each of the following primers: GCCCTGCAAGGTGTTCGACGATGT (ACE sense) and GGATGGCTCTCCCGCCGCTGTCTCTC (ACE antisense). The initial denaturation at 95°C for 5 min was followed by 35 cycles of 94°C for 30 s, 58°C annealing for 30 s, and 65°C elongation for 45 s. To avoid misclassification of ID genotypes into DD genotypes, a second PCR was performed using l-specific primers: TGGGACCCACAGGCGCCACTAC (l-specific sense) and TGCCACGCCCTCCCATGGCCATAA (l-specific antisense). This PCR yields a 335-bp fragment only in the presence of the l allele and no product in sample homozygotes for the D allele. Genotyping was performed by experienced staff. PCR scores by two independent investigators who were blind to subject data correlated well (r² = 0.991).

Data analysis. The SPSS statistical package version 13.0 was used for statistical evaluation (SPSS, Chicago, IL). Pre- and postexercise CK values were analyzed for their association with ACE genotypes. A χ² test was used to confirm that observed genotype frequencies were in Hardy-Weinberg equilibrium. Normality of each quantitative variable was tested using the Kolmogorov-Smirnov normality test. Genotype subtype comparisons were made by ANOVA and the Kruskal-Wallis test (asymmetrical data distribution). Continuous variables were compared according to genotype group by linear ANOVA. Genotype distribution across levels of CK response was compared by χ² for linear trend. Physiological parameters and genotype data were used in multivariate analysis by the use of forward stepwise regression to determine the model that best predicted CK response to eccentric exercise and to evaluate whether the number of ACE alleles carried by each subject had statistical influence on laboratory parameters. To account for any possible effects of age, sex, or body mass index (BMI), analyses were covaried for these parameters. Asymmetrically distributed variables were log transformed before regression analysis. Continuous data are presented as means ± SD or as means ± SE. Square multiple correlation coefficients (R²) were calculated. Two-tailed values of P < 0.05 were considered statistically significant.

RESULTS

Subjects were 25 ± 3 yr in age, 171 ± 8 cm in height, and 67 ± 10 kg in mass. The data on allele and genotype frequencies in the study population are shown in Table 1. There was no deviation from the Hardy-Weinberg equilibrium (allele frequency ACE ID = 0.37/0.63, expected genotype frequencies % II/DD/DD = 14%/47%/39%; χ² = 0.03; P = 0.98). Ashkenazi and Non-Ashkenazi descendants did not differ by ACE genotype. There was no relationship of age or body mass to ACE ID genotypes (Table 1). We, like others (8, 17), analyzed both sexes. Of the study population, 42 (60%) were men and 28 (40%) were women, and there was no difference in sex distribution among any of the genotype subtypes (Table 1). CK activity in response to the eccentric exercise regimen for the study population is shown in Fig. 1. Briefly, the baseline CK values were 157 ± 11 U/l (mean ± SE), and the average peak CK activity, which was noted 96 h postexercise (in all sub-

| Table 1. ACE ID allele and genotype frequencies in the study group with subjects’ physical characteristics |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
|                | ACE ID          | ACE ID          | ACE DD          | Allele I        | Allele D        | P Value |
| All, n = 70     | 10 (0.14)       | 32 (0.46)       | 28 (0.40)       | 52 (0.37)       | 88 (0.63)       | 0.18    |
| Men, n = 42     | 5 (0.12)*       | 21 (0.50)*      | 16 (0.38)*      | 31 (0.37)†      | 53 (0.63)†      | 0.91    |
| Women, n = 28   | 5 (0.18)*       | 11 (0.39)*      | 12 (0.43)*      | 21 (0.37)†      | 35 (0.63)†      | 0.93    |
| Age, yr         | 24 ± 3          | 25 ± 3          | 25 ± 3          | 25 ± 3          | 0.06 ± 0.03     | 0.96    |
| Height, cm      | 171 ± 6         | 171 ± 7         | 67 ± 10         | 67 ± 10         | 0.98            |         |
| Weight, kg      | 67 ± 9          | 67 ± 10         | 67 ± 10         | 67 ± 10         | 0.98            |         |
| LBM, kg         | 55 ± 11         | 55 ± 11         | 54 ± 11         | 54 ± 11         | 0.93            |         |
| BMI, kg/m²      | 22.6 ± 1.6      | 22.7 ± 1.6      | 22.8 ± 2.9      | 22.8 ± 2.9      | 0.96            |         |

Values are absolute and relative (in parentheses) frequencies, and means ± SD for continuous variables. ACE, angiotensin-converting enzyme; I, insertion allele; D, deletion allele; DD, homozygotes for the deletion allele; II, homozygotes for the insertion allele; ID, heterozygotes. *χ² = 0.93; degrees of freedom = 2; P = 0.62 for genotype frequencies in men vs. women. †χ² = 0.01; degrees of freedom = 1; P = 0.91 for allele frequencies in men vs. women.
Fig. 1. Creatine kinase (CK) activity in response to the eccentric exercise regimen for the study population. Subjects’ average serum CK activity at baseline and at different points in time postexercise.

Table 2. Subjects’ phenotypes in relation to ACE genotype

<table>
<thead>
<tr>
<th></th>
<th>ACE II (n = 13)</th>
<th>ACE ID (n = 32)</th>
<th>ACE ID + II (n = 42)</th>
<th>ACE DD (n = 28)</th>
<th>ANOVA P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baseline CK activity, U/l</td>
<td>147±27</td>
<td>158±15</td>
<td>155±13</td>
<td>161±19</td>
<td>0.920/0.82* 0.71†</td>
</tr>
<tr>
<td>Peak CK activity, U/l</td>
<td>8,882±2,361</td>
<td>4,454±1,105</td>
<td>5,508±1,040</td>
<td>2,937±2,753</td>
<td>0.020/0.07* 0.009†</td>
</tr>
<tr>
<td>Delta CK activity, U/l</td>
<td>8,735±2,352</td>
<td>4,296±1,108</td>
<td>5,353±1,041</td>
<td>2,778±2,757</td>
<td>0.020/0.07* 0.009†</td>
</tr>
</tbody>
</table>

Values are reported as means ± SE. *P value for CK activity in ACE II + ID vs. ACE DD. †P value for linear trend.
association studies and may be attributable to different experimental designs and study cohorts. Data suggest that the effect of ACE genotype on physical performance may depend on the type of exercise (28). Moreover, it is well known that the frequencies of the ACE ID alleles vary considerably among different control populations (3), and the influence of varying genetic background may obscure a true association. To support our findings, we have recently demonstrated in Israeli elite athletes an association of the ACE D allele and DD genotype with endurance performance (2). Of note, our individuals were physically active, but none were well trained or competitive athletes. More importantly, they were not engaged in any resistance training programs and were not highly active. Thus it seems that in the Heled et al. study (17) participants were very active and their fitness level was higher than ours. This is also supported by the relatively lower average increase in CK level of their high responders group compared with our average delta CK levels (1,048 vs. 4,480 U/L). Given that fitness level influences the degree of exercise-induced muscle damage (10) and since the effect of ACE polymorphism depends on individuals’ fitness levels as well (28), the association between ACE genotype and CK response to exercise may become prominent only in sedentary individuals performing a highly intense effort, as in our study.

Finally, it is still possible that ACE ID polymorphism is one of many genetic variants contributing to the observed variance in muscle CK response or that it is in strong allelic association with functional variants in adjacent genes and that these are responsible for the observed associations with ACE genotype (36). Likewise, it has been proposed that the higher ACE activity associated with the ACE DD genotype (12, 39) is perhaps caused by an Alu-associated transcription silencer (47, 48) or by an identified variant of the of the ACE gene that is in linkage disequilibrium with the Alu insertion/deletion polymorphism (21, 38, 49). However, the medical literature is still debating whether this candidate polymorphism is located in the promoter region (38, 49) or in downstream sequences of the ACE gene (21). The inconsistency regarding ACE haplotypes represents some of the difficulties in studying different populations, and the sequencing of coding and noncoding regions of the ACE gene in other populations with different evolutionary histories may reveal alternative polymorphisms. Rieder et al. (38) found some differences in sequence variation between African-Americans and European-Americans. In his analysis, a major genetic subdivision in the deletion haplotypes was evident only in European-Americans, further supporting the notion that some haplotypes that are present in one population may not apply to other populations. Clearly, more work on large sample sizes is needed to confirm our observations and to better clarify the pathways through which ACE polymorphism and the regulation of exertional muscle injury.


REFERENCES


