Rehydration with fluid of varying tonicities: effects on fluid regulatory hormones and exercise performance in the heat

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1Departments of Kinesiology, University of Connecticut, Storrs, Connecticut; 2Department of Kinesiology, University of Rhode Island, Kingston, Rhode Island; 3Department of Biology, University of Puerto Rico at Cayey, Cayey, Puerto Rico; and 4US Army Research Institute of Environmental Medicine, Natick, Massachusetts

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Kenefick RW, Maresh CM, Armstrong LE, Riebe D, Echegaray ME, Castellani JW. Rehydration with fluid of varying tonicities: effects on fluid regulatory hormones and exercise performance in the heat. J Appl Physiol 102: 1899–1905, 2007. First published February 22, 2007; doi:10.1152/japplphysiol.00920.2006.—This study examined the effects of rehydration (Rehy) with fluids of varying tonicities and routes of administration after exercise-induced hypohydration on exercise performance, fluid regulatory hormone responses, and cardiovascular and thermoregulatory strain during subsequent exercise in the heat. On four occasions, eight men performed an exercise-dehydration protocol of ∼185 min (33°C) to establish a 4% reduction in body weight. Following dehydration, 2% of the fluid lost was replaced during the first 45 min of a 100-min rest period by one of three random Rehy treatments (0.9% saline intravenous; 0.45% saline intravenous; 0.45% saline oral) or no Rehy (no fluid) treatment. Subjects then stood for 20 min at 36°C and then walked at 50% maximal oxygen consumption for 90 min. Subsequent to dehydration, plasma Na+, osmolality, aldosterone, and arginine vasopressin concentrations were elevated (P < 0.05) in each trial, accompanied by a ∼4% hemococoncentration. Following Rehy, there were no differences (P > 0.05) in fluid volume restored, post-rehydration (Post-Rehy) body weight, or urine volume. Percent change in plasma volume was 5% above pre-Rehy values, and plasma Na+, osmolality, and fluid regulatory hormones were lower compared with no fluid. During exercise, skin and core temperatures, heart rate, and exercise time were not different (P > 0.05) among the Rehy treatments. Plasma osmolality, Na+, percent change in plasma volume, and fluid regulatory hormones responded similarly among all Rehy treatments. Neither a fluid of greater tonicity nor the route of administration resulted in a more rapid or greater fluid retention, nor did it enhance heat tolerance or diminish physiological strain during subsequent exercise in the heat.

fluid regulation; hydration state; dehydration; environment; osmotic load

HYPOHYDRATION INDUCED by exercise-heat stress decreases plasma volume (PV) and increases plasma sodium concentration ([Na+]') and osmolality (Osm) (4, 23, 26). Concomitantly, exercise following hypohydration raises plasma concentrations of arginine vasopressin ([AVP]) (2, 7, 15) and aldosterone ([ALD]) (2, 8, 12) relative to euvhated control conditions. However, it is not evident whether the lower PV or higher [Na+]' caused by hypohydration influenced the exercise-induced changes in [AVP] and [ALD]. Furthermore, rehydration (Rehy) during or following exercise generally lowers [AVP] and [ALD] (2, 5, 17, 28), but the mechanisms for this are unclear.

Several investigations have addressed the roles of PV, [Na+]', and Osm during Rehy on fluid-regulating hormones. Moses et al. (18) used dehydration (Dehy) and intravenous (IV) infusion of 5% saline to examine mediators of AVP release and suggested that PV expansion following infusion may serve to attenuate the osmotic stimulus for AVP secretion. This finding implies that AVP would remain the same if IV fluids of varying tonicities were used during Rehy, if PV was restored similarly. Nose et al. (21) reported that, during recovery from Dehy, rehydrating orally with a Na+-containing drink (compared with drinking water) better restored PV and lowered plasma renin activity and ALD to a greater extent, suggesting PV changes have a larger role than osmotic changes in mediating renin and ALD responses after oral Rehy (21). They stated that an inferior Rehy occurs when drinking hypotonic water, because it removes the osmotic drive for drinking and increases free water clearance, suggesting that Rehy with fluids of greater tonicity would better maintain PV compared with fluids of lesser tonicity. It is unknown whether tonicity differences would also cause differential PV and ALD responses after exercise-induced Dehy, if Rehy fluid were delivered via IV, rather than orally.

In addition, whether IV Rehy following exercise-induced hypohydration should be isosmotic or hypotonic has practical implications, if multiple bouts of exercise in the heat are performed and if the type of IV Rehy solution is found to differentially affect PV restoration or plasma [Na+]' and Osm. Numerous studies have shown that either lower PV or higher Osm causes the sweating rate or cutaneous blood flow (9) to be lower at any given core temperature and for these thermoregulatory effector responses to begin at a higher core temperature (22). Changes in these thermoregulatory responses subsequent to different IV treatments could potentially affect core temperature and exercise performance during repeated exercise bouts.

The primary purpose of this study was to examine the effects of hypovolemia and [Na+]' on fluid-regulatory hormone responses to exercise in the heat, subsequent to exercise-induced hypohydration. To test the hypothesis that differences in either PV or [Na+]' would affect plasma [AVP] and [ALD] during exercise heat stress, measurements were made following Dehy with two IV treatments (0.9 and 0.45% NaCl), an oral Rehy treatment (0.45% Oral), and a no-fluid control treatment (NF). We hypothesized that, following Dehy: 1) IV Rehy with a fluid

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of greater tonicity (0.9% saline) would increase the plasma [Na⁺] above that of a fluid of lesser tonicity (0.45% saline); 2) the greater [Na⁺] associated with the 0.9% IV treatment would subsequently elevate plasma Osm and stimulate AVP secretion above that of the 0.45% IV or Oral treatments; 3) due to the more rapid restoration of PV in the IV treatment and the greater fluid retention in the 0.9% IV treatment (a result of higher [AVP]), there will be less cardiovascular and thermoregulatory strain, greater heat tolerance, and exercise duration during subsequent exercise in the heat, compared with the 0.45% Oral and IV saline treatments. A secondary focus was to determine whether the route of fluid administration (Oral vs. IV) would result in any physiological differences pertaining to fluid-regulatory responses.

METHODS

Subjects. Eight men, unacclimatized to heat, volunteered to participate in this investigation. Physical characteristics (mean ± SE) were as follows: age, 22.1 ± 0.8 yr; height, 179.6 ± 1.5 cm; weight, 73.6 ± 2.4 kg; maximal oxygen consumption (V̇O₂ max), 57.9 ± 1.6 ml·kg⁻¹·min⁻¹; percent body fat, 7.7 ± 0.9%. Before participation, each subject completed a written, informed consent document and a medical history questionnaire after being informed of the purpose of the experiment and possible risks. The study protocol was approved in advance by the Committee on the Use of Human Subjects in Research at the University. Subjects were paid for their participation.

Preliminary measures. V̇O₂ max was determined using a continuous treadmill running test, as previously described (9). Briefly, the V̇O₂ max test was preceded by a 7-min warm-up run at ~160–200 m/min, 0% grade. During the maximal oxygen test, speed was kept constant for 160–220 m/min, 0% grade, for the first 4 min, after which the grade was raised to 4%. Thereafter, the grade was increased by 2% every 2 min until volitional exhaustion. The hydrostatic weighing technique described by Katch and Katch (14) was utilized to determine body density. Body fat was calculated from the formula of Brozek et al. (3). Measurement of residual lung volume was proved in advance by the Committee on the Use of Human Subjects in Research at the University. Subjects were paid for their participation.

Experimental design. Testing involved three treatments, each consisting of three stages, as follows: 1) exercise-induced Dehy, 2) Rehy, and 3) moderate exercise in the heat. The protocols were randomly assigned and separated by at least 14 days. The Rehy treatments were as follows: 0.9% IV infusion (0.9% IV); 0.45% IV infusion (0.45% IV); 0.45% Oral Rehy (0.45% Oral), and no Rehy (NF). Subjects were asked to consume similar diets during the 3 days before each experimental trial, verified by a 3-day dietary record. These food records were then analyzed for kilocalories, carbohydrate, fat, protein, sodium, and potassium content (Food Processor II, ESHA Research, Salem, OR). Subjects were asked to refrain from any recreational activities or exercise training for 24 h before experimental testing. They were also instructed to drink 450 ml of water the night before testing, and 450 ml of water the morning of testing, and to abstain from eating for 12 h before each experimental treatment.

Experimental treatments. On arrival at the laboratory (0700), subjects provided a urine sample for determination of urine-specific gravity (U₉g; Spartan Refractometer, model A 300 CL). A U₉g of 1.023 ± 0.006 (1) was used to verify that the subject was adequately hydrated before each trial. Hydration status was additionally verified by a pre-Dehy plasma Osm value of ±286 ± 3 mosmol/kgH₂O (1). Subjects were then fitted with a heart rate monitor (UNIQ heartwatch, Computer Instrument, Hempstead, NY), and a flexible thermistor (Yellow Springs Instruments, series 401, Yellow Springs, OH) was inserted 10 cm beyond the external anal sphincter to monitor rectal temperature (Tre). A Teflon catheter was then inserted into a superficial forearm vein, and a male luer adapter (model 5877, Abbott Hospital, Chicago, IL) was inserted into the catheter port for acquisition of subsequent blood samples. The catheter port and male luer adapter were kept patent with heparin Lock Flush Solution. The subject then entered the environmental chamber (model 2000, Minus-Eleven, Malden, MA) and stood quietly during a 20-min equilibration period. A 26-ml blood sample (baseline) was taken, and subjects then consumed a standard breakfast of one bagel, one banana, and 240–350 ml (depending on body weight) of fruit juice.

Immediately before each of the four experimental trials, subjects performed a Dehy protocol in 33°C air, to reduce body weight by ~4%. The Dehy protocol consisted of alternating stationary cycling (117 ± 9 W; model 818E, Monark, Sweden) and treadmill walking (1.6 ± 0.1 m/s; 5 ± 1% grade; Quinton, Seattle, WA) at a 25-to-5-min ratio (exercise/rest) for each modality. Body weight was measured during each rest interval. Urine was collected throughout the Dehy period and was included as part of the weight loss. Subjects continued exercising until the desired weight loss was achieved. The last exercise mode before the 4% weight loss was always walking, to ensure an upright posture. The time to achieve the 4% decrease in body weight before each of the three treatments was consistent for each individual subject (Table 1). The mean exercise intensity for the four Dehy trials ranged from 49.8 to 51.1% of V̇O₂ max. The mean ambient temperature and percent relative humidity were 33.0 ± 0.1°C and 47.6 ± 0.5%, respectively. Airflow (2.3 m/s) was generated by a fan directed at the subject.

Following the exercise-induced Dehy protocol, a 26-ml blood sample was taken, and the subject exited the environmental chamber to a 25.5 ± 0.2°C environment. Subjects assumed a recumbent position, and after a 15-min rest period received one of the three Rehy treatments or NF control over a 45-min period. An experienced IV nurse, using a butterfly

Table 1. Selected Dehy and Rehy variables

<table>
<thead>
<tr>
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<th>0.45% IV</th>
<th>0.45% Oral</th>
<th>0.9% IV</th>
<th>NF</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dehy %V̇O₂ max, ml·kg⁻¹·min⁻¹</td>
<td>51 ± 2</td>
<td>50 ± 2</td>
<td>50 ± 3</td>
<td>51 ± 2</td>
</tr>
<tr>
<td>Exercise time, min</td>
<td>190 ± 12</td>
<td>182 ± 8</td>
<td>179 ± 12</td>
<td>180 ± 10</td>
</tr>
<tr>
<td>Post-Dehy urine volume, liter</td>
<td>0.34 ± 0.12</td>
<td>0.40 ± 0.10</td>
<td>0.44 ± 0.11</td>
<td>0.48 ± 0.09</td>
</tr>
<tr>
<td>Post-Rehy urine volume, liter</td>
<td>0.06 ± 0.02</td>
<td>0.08 ± 0.02</td>
<td>0.08 ± 0.02</td>
<td>0.08 ± 0.02</td>
</tr>
<tr>
<td>Post-Rehy, %weight loss</td>
<td>4.4 ± 0.07</td>
<td>5.1 ± 0.1</td>
<td>4.5 ± 0.07</td>
<td>4.6 ± 0.07</td>
</tr>
<tr>
<td>Post-Rehy %weight loss</td>
<td>2.1 ± 0.10</td>
<td>2.3 ± 0.1</td>
<td>2.1 ± 0.10</td>
<td>4.6 ± 0.04*</td>
</tr>
</tbody>
</table>

Values are means ± SE; n = 8. IV, intravenous; Oral, oral rehydration; NF, no fluid control; Dehy, dehydration; V̇O₂ max, maximal oxygen consumption; Post-Dehy, post-dehydration; Post-Rehy, post-rehydration. *Difference from 0.9% IV, 0.45% IV, and 0.45% Oral treatments (P < 0.05).
catheter (1.5 in., 16 gauge; Abbot Laboratories, North Chicago, IL), administered the IV infusions in the arm opposite to the indwelling venous cannula. During the NF trials, a cannula was not placed in the opposite arm during the Rehy period. The rate of IV infusion was 0.56 ml·kg⁻¹·min⁻¹. Oral Rehy consisted of 4 g of Sugar Free Tropical Punch Kool-Aid dissolved in 889 ml of 0.45% NaCl and 111 ml of distilled, deionized water chilled to 4°C for palatability. The composition of 0.45% Oral fluid was 78.6 ± 1.0 meq/l Na⁺, 0.96 ± 0.02 meq/l K⁺, and 2.54 ± 0.07 meq/l Ca²⁺. The Osm was 145.6 ± 1.1 mosmol/kgH₂O. Fluid given in the 0.45% Oral trial was weighed before each drink and was given every 5 min over a 45-min period. The fluid volumes given were not different (P > 0.05) and were 1,890 ± 45, 1,856 ± 65, and 1,889 ± 62 ml for the 0.45% IV, 0.45% Oral, and 0.9% IV treatments, respectively. The rate of IV infusion and oral Rehy over 45 min was chosen to allow for a fluid volume restoration to be similar to the upper range for orally ingested fluids after exercise-induced Rehy (20). Subjects then stood for 55 min in the laboratory to allow for equilibration of the fluid and then reentered the environmental chamber.

Subjects reentered the climatic chamber, equilibrated (standing for 20 min), and a 26-ml blood draw was performed. Subjects then consumed 1 g/kg body wt of a commercial carbohydrate product (Skittles, M and M Mars, Hackettstown, NJ) and 100 ml of distilled, deionized water. Carbohydrates were given before the second exercise bout to offset the possible loss of muscle glycogen during the Rehy protocol. Subjects then performed a 90-min exercise bout of treadmill walking, up a 3–5% grade at ~50% of V̇O₂max. The exercise bout could be stopped either by subjects (volitional exhaustion), or by the researchers, due to heart rate (>180 beats/min for 5 consecutive min) or Tₑₑ (>39.5°C) criteria or symptoms of heat exhaustion. Walking speed was verified for each test with a hand-held tachometer (model 8204-20, Cole-Parmer Instrument, Chicago, IL). A 26-ml blood draw was performed at the completion of the exercise bout. The mean temperature and relative humidity were 35.9 ± 0.1°C and 46.6 ± 2%, respectively, with airflow at 2.3 m/s.

Physiological measures. During all Rehy bouts, the subjects’ heart rate and Tₑₑ were measured every 15 min to monitor physiological strain. Oxygen consumption was measured once every 30 min during the Rehy protocol for a 7- to 10-min period.

Expired gas samples were analyzed in each breath (Medical Graphics CPX-D system, Medical Graphics, St. Paul, MN). During exercise in the heat, thermocouples (series 400, Yellow Springs Instruments) secured on the chest, arm, thigh, and calf were used to measure skin temperature (Tₕₕ), computed from a weighted mean of the four local skin measurements (20).

Analysis of blood samples. Blood measures were analyzed at four time points: pre-dehydration (Pre-Dehy), post-dehydration (Post-Dehy), preexercise in the heat (Pre-Ex), and postexercise in the heat (Post-Ex). Blood was transferred from sterile syringes to tubes containing EDTA, lithium heparin, or SST gel and clot activator for serum separation, depending on the analytical specifications for each blood variable. Tubes were centrifuged (Marathon 12KBR Refrigerated Centrifuge, Fisher Scientific, Pittsburgh, PA) for 15 min using 760 g at 4°C. Serum or plasma was separated from each sample and stored at −80°C for later analysis. Samples of whole blood were taken for analysis of hemoglobin (Hb) and hematocrit (Hct). Following centrifugation, plasma was separated and analyzed for Osm and Na⁺. Absolute changes in plasma AVP, ALD, Na⁺, Osm, and percent change of PV (%ΔPV) were calculated and analyzed relative to the Pre-Dehy time point.

Hct was determined in triplicate by the microcapillary technique following centrifugation for 4 min at 9,500 g. Values were not corrected for trapped plasma. Hb was determined in triplicate by the cyanomethemoglobin method (kit 525, Sigma Chemical, St. Louis, MO). %ΔPV was calculated using the equation of Dill and Costill (9a) from appropriate Hct and Hb values. All %ΔPV values were calculated using Post-Dehy as the initial time point. Plasma Osm was measured in triplicate via freezing point depression (MicroOsmometer model 3MO, Advanced Instruments, Needham Heights, MA). Plasma and urine [Na⁺] were determined in duplicate by selective ion-sensitive electrodes (model 984-S AVL Scientific, Roswell, GA).

Endocrine measures. After extraction on silica columns (DiaSorin, Stillwater, MN), plasma AVP was determined in duplicate by a commercially available radioimmunoassay kit (DiaSorin). AVP values were not corrected for extraction recovery, which was 91.3%. The limit of the detection for this assay was ~1.4 pg/ml. Serum levels of ALD were determined in duplicate by radioimmunoassay (Diagnostics Products, Los Angeles, CA). Assay sensitivity was 11.0 pg/ml. Within- and between-assay coefficients of variability for both assays were <5%. [AVP] and [ALD] were not corrected for %ΔPV.

Statistical analysis. A 4 x 3 (treatment x time) repeated-measures ANOVA was used to compare differences among trials. A Newman-Keuls post hoc analysis was employed to determine significant differences within and between conditions. The 0.05 level of significance was selected. Studies examining fluid-regulating hormones during Dehy have a coefficient of variation of 5%. A power analysis selecting conventional α (P < 0.05) and β (0.20) parameters showed that eight subjects would provide sufficient power to detect an effect greater than or equal to the anticipated coefficient of variation. All data are presented as means ± SE.

RESULTS

Dehy. Pre-Dehy dietary carbohydrate, protein, fat, sodium, and total kilocalorie intakes were similar over the 3 days before each experimental treatment. Pre-Dehy body weights were not different (P > 0.05) and were 75 ± 2, 74.3 ± 2.6, 74 ± 3, and 74 ± 3 kg for the 0.45% IV, 0.45 oral, 0.9% IV, and NF treatments, respectively. The exercise intensity during the Dehy phase across the four experimental conditions ranged from 49.8 to 51.1% of V̇O₂max. There were no differences (P > 0.05) among the three treatments and the NF trial for Dehy percent weight loss, urine volume, and exercise intensity (%V̇O₂max) during Dehy (Table 1).

Pre- and Post-Dehy plasma and urine values. Pre-Dehy plasma AVP, ALD, Osm, and [Na⁺] were not different (P > 0.05) among treatments. In addition, Pre-Dehy urine Osm, [Na⁺], and Uₕₕ values were not different (P > 0.05) among treatments. Following Dehy, plasma values of AVP, ALD, Osm, and [Na⁺] were elevated (P < 0.05) above Pre-Dehy values but were not different among the Rehy treatments and NF trial Post-Dehy. Post-Dehy urine Osm, [Na⁺], and Uₕₕ were also not different (P > 0.05) from Pre-Dehy values.
(Table 2). In addition, the %ΔPV calculated from the Pre-Dehy was not different (P > 0.05) among the Rehy treatments and the NF trial; the change from Pre- to Post-Dehy averaged −5.2% for the Rehy treatments and the NF trial (see Fig. 2A).

Rehydration and exercise. There were no differences in the volume of IV or Oral fluid (0.9% saline IV, 0.45% saline IV, and 0.45% saline Oral) given during the Rehy period (Table 1). Post-Rehy urine volume was similar among the Rehy and NF treatments. The percentage of weight loss Post-Rehy (compared with the Pre-Rehy body weight) was similar among the 0.45% IV, 0.45 Oral, and 0.9% IV treatments and were significantly lower (P < 0.05) than in the NF trial (Table 1).

The absolute change from Pre-Rehy in plasma [Na⁺] was lower (P < 0.05) in the 0.45% Oral treatment compared with both the 0.9% IV and NF treatments after Rehy (Pre-Ex time point). However, Post-Ex in the heat, there were no differences (P > 0.05) in the absolute change in Na⁺ among any of the treatments or the NF trial (Fig. 1A). The absolute change in plasma Osm in the NF trial was greater than that observed in the 0.45% IV and Oral treatments following Rehy (Pre-Ex time point); however, following exercise in the heat, none of the Rehy treatments or the NF trial were different (P > 0.05) (Fig. 1B).

Following Rehy, at the Pre-Ex and Post-Ex time points, the absolute change in plasma AVP was greater (P > 0.05) in the NF trial compared with the Rehy treatments. Following exercise in the heat, there was an increase in the absolute plasma AVP in all three treatments and the NF trial, but this change was only different (P < 0.05) from Pre-Ex in the NF trial (Fig. 1C).

Subsequent to Rehy, the %ΔPV at the Pre-Ex time point was not different (P > 0.05) among the Rehy treatments, but was greater (P < 0.05) than in the NF trial. Following exercise in the heat, there was a significant hemococoncentration in the Rehy treatments (P < 0.05) relative to the Pre-Ex time point. In the NF trial, the %ΔPV did decrease slightly Post-Ex; however, there was no difference (P > 0.05) in the %ΔPV Post-Ex among the NF trial and Rehy treatments (Fig. 2A).

At the Pre-Ex and Post-Ex time points, the absolute change in plasma ALD was greater (P > 0.05) in the NF trial compared with the Rehy treatments. Following exercise in the heat, there was an increase (P < 0.05) in the absolute change in plasma ALD in all three treatments and the NF trial (Fig. 2B).

Selected Pre- and Post-Ex thermoregulatory and exercise performance time. Exercise performance times were longer (P < 0.05) in 0.45% IV, 0.45% Oral, and 0.9% IV treatments compared with NF (Table 3). Exercise trials in the heat were terminated for the following reasons: during the 0.45% IV, three subjects reached 39.5°C, one reached the heart rate limit of >180 beats/min for 5 consecutive min, one subject developed signs and symptoms of heat exhaustion, and three completed 90 min of exercise; during the 0.45% Oral trial, two subjects reached 39.5°C, two subjects exhibited symptoms of fatigue, and four completed 90 min of exercise; during the 0.9% IV trial, two subjects reached 39.5°C, two subjects exhibited symptoms of fatigue, and four completed 90 min of exercise; during the NF trial two subjects reached 39.5°C, four subjects exhibited symptoms of fatigue, and two completed 90 min of exercise.

Pre-Ex measures of Tₑₑ were lower (P < 0.05) in the Rehy treatments compared with the NF trial. Immediate Post-Ex, Tₑₑ were higher (P < 0.05) than Pre-Ex temperatures for all Rehy treatments and the NF trial, but were not different (P > 0.05) among each other. Pre-Ex Tₕₕ were also lower (P < 0.05) in the Rehy treatments compared with NF; however, after exercise, Tₕₕ were not different among the 0.9% IV, 0.45% IV, and Oral treatments and the NF trial. NF Pre-Ex heart rates were greater (P < 0.05) compared with the Rehy treatments, but Post-Ex heart rates were not different (P > 0.05) among treatments. All Pre-Ex Rehy treatment body weights were greater (P < 0.05) than NF. Only the NF Post-Ex body weights were significantly lower (P < 0.05) than Pre-Ex. In addition, the NF Post-Ex body weights were lower (P < 0.05) compared with Rehy Post-Ex values.

DISCUSSION

In the present study, a 4–5% exercise-induced hypohydration caused subjects to become hyperosmotic, hypovolemic, and elevated circulating [AVP] and [ALD] (5, 8). The results of the present investigation demonstrate that, following partial Rehy, there were no overall differences in %ΔPV or absolute change of plasma sodium, Osm, [AVP], and [ALD] among the Rehy treatments. There was no greater fluid restoration associated with the 0.9% IV compared with the 0.45% IV treatment, nor was there a greater fluid restoration imparted by IV compared with oral Rehy. As a result, there were no cardiovascular, thermoregulatory, or performance advantages during

Table 2. Selected plasma and urine variables pre- and post-Dehy

<table>
<thead>
<tr>
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<th>Pre-Dehy</th>
<th>Post-Dehy</th>
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<tr>
<td></td>
<td>0.45% IV</td>
<td>0.45% Oral</td>
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<tr>
<td><strong>Plasma</strong></td>
<td></td>
<td></td>
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<tr>
<td>AVP, pg/ml</td>
<td>5.5 ± 0.8</td>
<td>5.8 ± 0.9</td>
</tr>
<tr>
<td>ALD, pg/ml</td>
<td>410 ± 69</td>
<td>327 ± 57</td>
</tr>
<tr>
<td>Osm, mosmol/kgH₂O</td>
<td>284 ± 1</td>
<td>282 ± 2</td>
</tr>
<tr>
<td>Na⁺, meq/l</td>
<td>146 ± 4</td>
<td>145 ± 10</td>
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<tr>
<td><strong>Urine</strong></td>
<td></td>
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<tr>
<td>Volume (ml)</td>
<td>499 ± 117</td>
<td>487 ± 76</td>
</tr>
<tr>
<td>Osm, mosmol/kgH₂O</td>
<td>61 ± 9</td>
<td>75 ± 14</td>
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<tr>
<td>Na⁺, meq/l</td>
<td>1.015 ± 0.003</td>
<td>1.017 ± 0.003</td>
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Values are means ± SE; n = 8. Pre-Dehy, pre-dehydration; AVP, arginine vasopressin; ALD, aldosterone; Osm, osmolality. *Difference from corresponding 0.45% IV, 0.45% Oral, 0.9 % IV, and NF Pre-Dehy value (P < 0.05).
subsequent exercise in the heat associated with either a particular IV solution or administration route. Plasma [Na⁺], Osm, and AVP responses. Prolonged exercise in the heat without Rehy increases plasma [AVP] (2, 7, 16). As plasma Osm has been identified as the primary mediator of AVP release (18), we hypothesized that infusion of the 0.9% IV saline infusion would raise the plasma [Na⁺] and plasma Osm above that of the 0.45% IV treatment, further stimulating AVP secretion in 0.9% IV treatment. The present results do not agree with our hypothesis. In the present study, during a 75-min post-Rehy recovery (55-min post-Rehy + 20-min equilibration), the 0.9% IV [Na⁺] were elevated in the 0.9% IV compared with the 0.45% IV treatment at the Pre-Ex time point, but was not significantly different, despite the greater tonicity of the 0.9% IV fluid. As a result, the absolute change in plasma Osm and [AVP] was also not different between the IV treatments. Thus the greater [Na⁺] in the 0.9% IV treatment may not have been large enough to cause differences in plasma Osm between the 0.9% IV and the 0.45% IV or Oral treatments.

We previously reported a reduction in plasma AVP after 55 min of Rehy with 0.9% and 0.45% IV fluid (15). In that study, plasma [Na⁺] in the 0.9% Rehy trial were significantly elevated above that of the 0.45% at 55 min post-Rehy. Also, while not significantly different, plasma AVP was greater after 55 min post-Rehy in the 0.9% Rehy trial compared with the 0.45% trial. Our current results (reduced [AVP] and [ALD]) agree with previous studies where subjects received water or an electrolyte solution before or during exercise in a warm environment (2, 12, 13). Follenius et al. (10) found that progressive Rehy, by either acid isotonic solution or neutral isotonic solution, equally maintained PV and blunted the AVP response. In contrast, Thompson et al. (27) reported that, in addition to increasing blood volume, infusion of hypertonic solutions...
saline resulted in a significant rise in plasma sodium, Osm, and plasma AVP. Differences in the findings of our study and that of Thompson et al. may be due to time of observation, as Thompson et al. observed responses immediately following IV infusion and our observations were following 75 min. Restoration of PV was achieved in the present study by IV solutions infused and our observations were following 75 min. Restor- 
plasma AVP. Differences in the findings of our study and that
Selected Pre-Ex and exercise performance variables

Table 3. Selected Pre-Ex and exercise performance variables

<table>
<thead>
<tr>
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<th>0.45% IV</th>
<th>0.45% Oral</th>
<th>0.9% IV</th>
<th>NF</th>
<th>0.45% IV</th>
<th>0.45% Oral</th>
<th>0.9% IV</th>
<th>NF</th>
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<tbody>
<tr>
<td>Exercise time, min</td>
<td>77±5</td>
<td>84±2</td>
<td>76±6</td>
<td>58±8†</td>
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</tr>
<tr>
<td>Tsk, °C</td>
<td>37.5±0.1</td>
<td>37.4±0.1</td>
<td>37.5±0.1</td>
<td>37.8±0.1*</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tsa, °C</td>
<td>31.0±0.2</td>
<td>31.1±0.1</td>
<td>31.1±0.2</td>
<td>31.8±0.1†‡</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Heart rate, beats/min</td>
<td>102±5</td>
<td>93±4</td>
<td>97±4</td>
<td>113±2*</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Body weight, kg</td>
<td>73±2.3</td>
<td>72±2.7</td>
<td>73±2.7</td>
<td>71.4±2.3*</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Values are means ± SE; n = 8. Tsk, rectal temperature; Tsa, skin temperature. *Difference from 0.45% IV and 0.9% IV values (P < 0.05). †Difference from 0.45% Oral value (P < 0.05). ‡Difference from corresponding Pre-Ex value (P < 0.05).

In summary, the results of the present investigation demonstrate that Rehy with 0.9% IV saline did elevate plasma sodium, Osm, and [AVP] concentrations above levels observed following 0.45% IV Rehy. However, while Rehy with fluid containing sodium has been shown to result in greater restoration of PV compared with water (20), it appears that the greater toxicity of the 0.9% fluid did not result in greater restoration of PV, enhanced heat tolerance, or diminished physiological strain during subsequent exercise in the heat compared with 0.45% saline fluid.

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DISCLAIMER

The views, opinions, and/or findings in this report are those of the authors and should not be construed as official Department of the Army position, policy, or decision, unless so designated by other official designation. All experiments were carried out in accordance with state and federal guidelines.

REFERENCES